



A research and development partnership between Africa and Australia

Genetic diversity and structure in Cameroon native goat populations using microsatellites

Meutchieye, F., Nongo, P.J., Djikeng, A., Ommeih, S., Agaba, M., Yacouba, Y.

Univ. Dschang, Cameroon ; Univ. N'dere, Cameroon; BecA-Hub, ILRI, Nairobi, Kenya



64th

EAAP 2013

ANNUAL MEETING
OF THE EUROPEAN FEDERATION OF ANIMAL SCIENCE

AUGUST 26TH - 30TH, 2013
NANTES, FRANCE



Background

Indigenous/local goat breeds are critical for subsistence of smallholders in the world (Pollet and Wilson, 2009).

In most developing countries, goats breeds have been merely considered and documented (Wilson, 1990) .

=> **Consequences**: low performances, unpredictable crossings and genetic erosion (Groenevald *et al.*, 2010).

Genetic status of majority of livestock in developing world is still very scanty (Guimarães *et al.*, 2007).

Background

With about 5 millions heads, Cameroon hosts a variety of goats types (MINEPIA, 2010)

Common names from literature: *Cameroon Dwarf*, *West African Dwarf Goat*-Chèvre Naine de Guinée, *Djallonke Goat*, *Nigerian Goat*, *Pygmy Goat*, *Dwarf Goat*-Chèvre Naine, *Fouta Djallon Goat et chèvre Kirdi* (Epstein, 1962; Devendra and Burns, 1982; Lauvergne *et al.*, 1993).

These various names = possible genetic variability in goats from the coastal Atlantic ocean => northern arid Sahelian region?

Background

Genomic application in farm animal is at its initial stage in Cameroon.

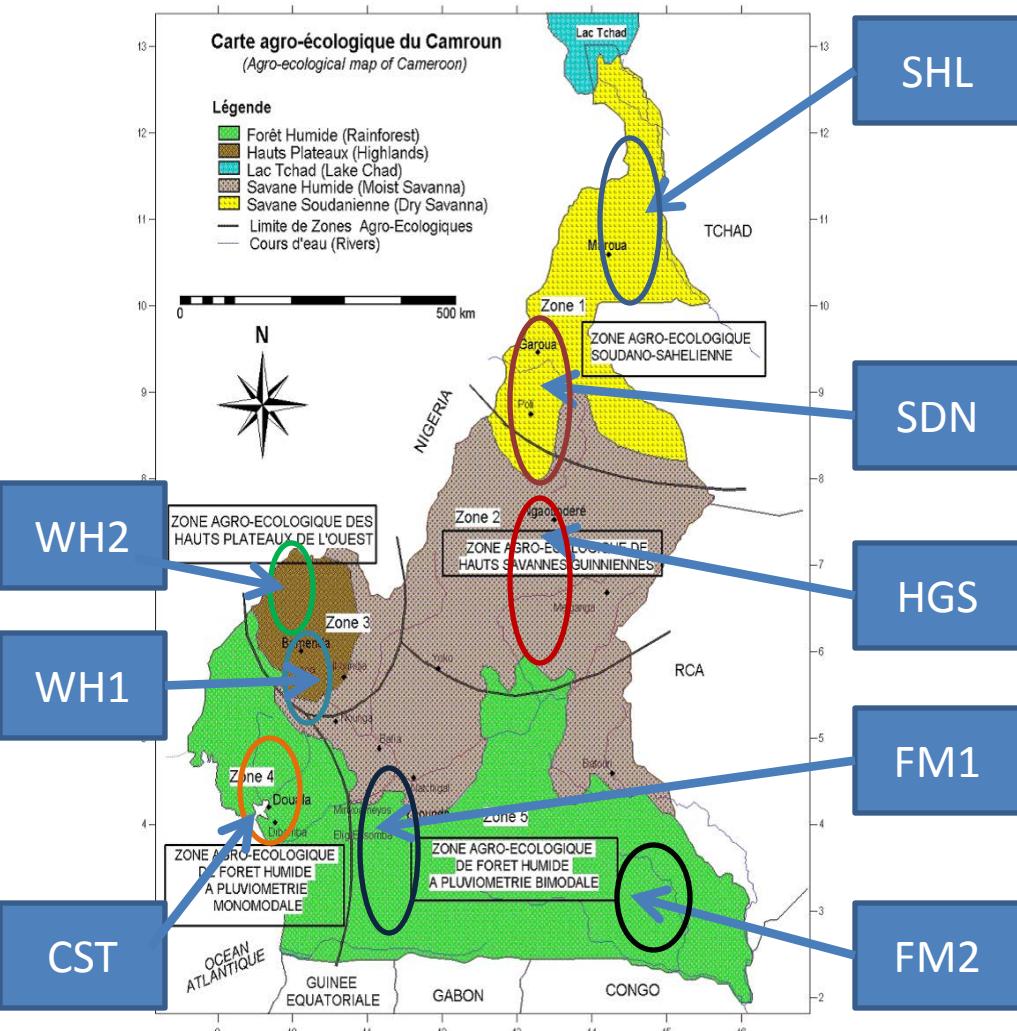
Short tandem repeat (STR) or short sequence repeats (SSR) markers or microsatellites developed did not take into account sub saharan goat populations variability (Hanotte et al., 2004).

Their application are popularized in tropics as a tool to assess in very efficient way the diversity in livestock

Research Objective

Characterize the genetic relationships between 08 Cameroonian goats ecotypes by the application of a panel of 17 SSR markers

Sampling approach



- 190 goats (M/F) not related and randomly
- Dry spot blood on FTA Whatmann
- Georeferencing with GPS
- gDNA extraction using Invitrogen kit
- 35 cycles for 25µl total volume Rx
- Genotyping under ABI 3130 and GeneMapper v.4

Figure 1: Agro ecological regions and sampled ecotypes

**SHL = Sahelian**

Longipes

HW \geq 65cm; LBW=20-45kg

Variable coat color

Meat, skin and milk

SDN = Kirdi

Longipes-Mediopes

HW \approx 60cm; LBW=20-35kg

Variable coat color

Meat, skin and milk

**WH1 = Djallonke**

Mediopes-Naine

HW \leq 65cm; LBW=15-30kg

Variable coat color

Meat and goatlings

HGS=Djallonke

Longipes-Mediopes

HW \leq 65cm; LBW=15-40kg

Variable coat color

Meat and milk

**WH2 = Djallonke**

Mediopes-Dwarf

HW \leq 65cm; LBW=15-30kg

Variable coat color

Meat

FM2=Djallonke

Dwarf

HW \leq 55cm; LBW=15-25kg

Variable coat color

Meat

**FM1 = Djallonke**

Dwarf

HW \leq 50cm;LBW=15-25kg

Variable coat color

Meat

CST= Djallonke

Dwarf

HW \leq 50cm;LBW=15-25kg

Variable coat color

Meat



Figure 2: Main features of local goat ecotypes

Table 1: Microsatellites Markers

N°	Microsatellite	Allele size (bp)	Annl. T°c	Dye
1	ILSTS029	148-170	56.6	NED
2	SRCRSP3	98-122	56.6	NED
3	ETH10	200-210	57.0	PET
4	CSRD247	220-247	62.2	PET
5	TCRVB6	217-255	59.2	PET
6	SRCRSP7	117-131	50.7	PET
7	SRCRSP9	99-125	58.8	PET
8	McM527	167-247	59.0	VIC
9	ILSTS005	172-208	60.3	VIC
10	MAF209	100-104	56.6	VIC
11	OarFCB20	80-130	54.0	VIC
12	MAF065	116-158	62.5	VIC
13	DRBP1	195-229	63.0	VIC
14	INRA063	164-186	63.3	VIC
15	MAF70	134-178	60.6	6-FAM
16	BM6444	118-200	65.5	6-FAM
17	SPS113	134-158	62.5	6-FAM

Some statistical parameters:

- ✓ Allele frequency and Heterozygosity
- ✓ Diversity and Polymorphism information content
- ✓ AMOVA under GenAIEX 6.5 (Peakall and Smouse 2012)
- ✓ FST values amongst populations (Arlequin)
- ✓ K probability, K = sub populations given by Evanno et al., (2005) under STRUCTURE
- ✓ Admixture estimation of caprine population following Pritchard et al., (2000) ; et Falush et al., (2003 et 2007).

Results

Table 2: Statistical values obtained from microsatellites analyses

Markers	Maj. Allele Freq.	Genotype Nbr	Obs. Nbr	Allele Nbr	Gene Diversity	He	PIC
BM6444	0.241	28	93	15	0.864	0.258	0.851
CSRD247	0.317	29	93	13	0.811	0.602	0.788
DRBP1	0.301	20	138	10	0.820	0.239	0.799
ETH10	0.357	10	157	5	0.677	0.561	0.608
ILSTS005	0.869	12	176	7	0.239	0.102	0.232
ILSTS029	0.237	43	150	12	0.853	0.793	0.836
INRA063	0.632	11	125	7	0.543	0.376	0.496
MAF065	0.383	26	64	10	0.782	0.453	0.758
MAF70	0.341	42	167	14	0.794	0.677	0.769
MAF209	0.424	11	170	5	0.677	0.529	0.615
MCM527	0.368	24	167	10	0.749	0.515	0.712
OarFCB20	0.237	33	173	10	0.821	0.555	0.798
SPS113	0.421	22	87	8	0.809	0.713	0.783
SRCPSP3	0.513	15	140	6	0.682	0.557	0.628
SRCRSP7	0.286	8	79	5	0.545	0.443	0.444
SRCRSP9	0.340	38	119	12	0.846	0.597	0.831
TCRVB6	0.386	25	119	9	0.776	0.697	0.743
Mean	0.386	23.35	130.412	9.294	0.723	0.510	0.688

Results

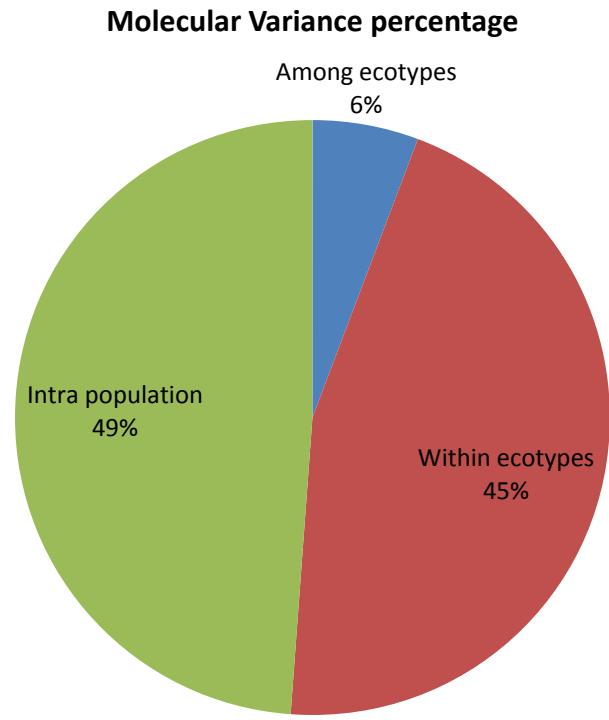


Figure 3: Molecular variance in Cameroon goat population

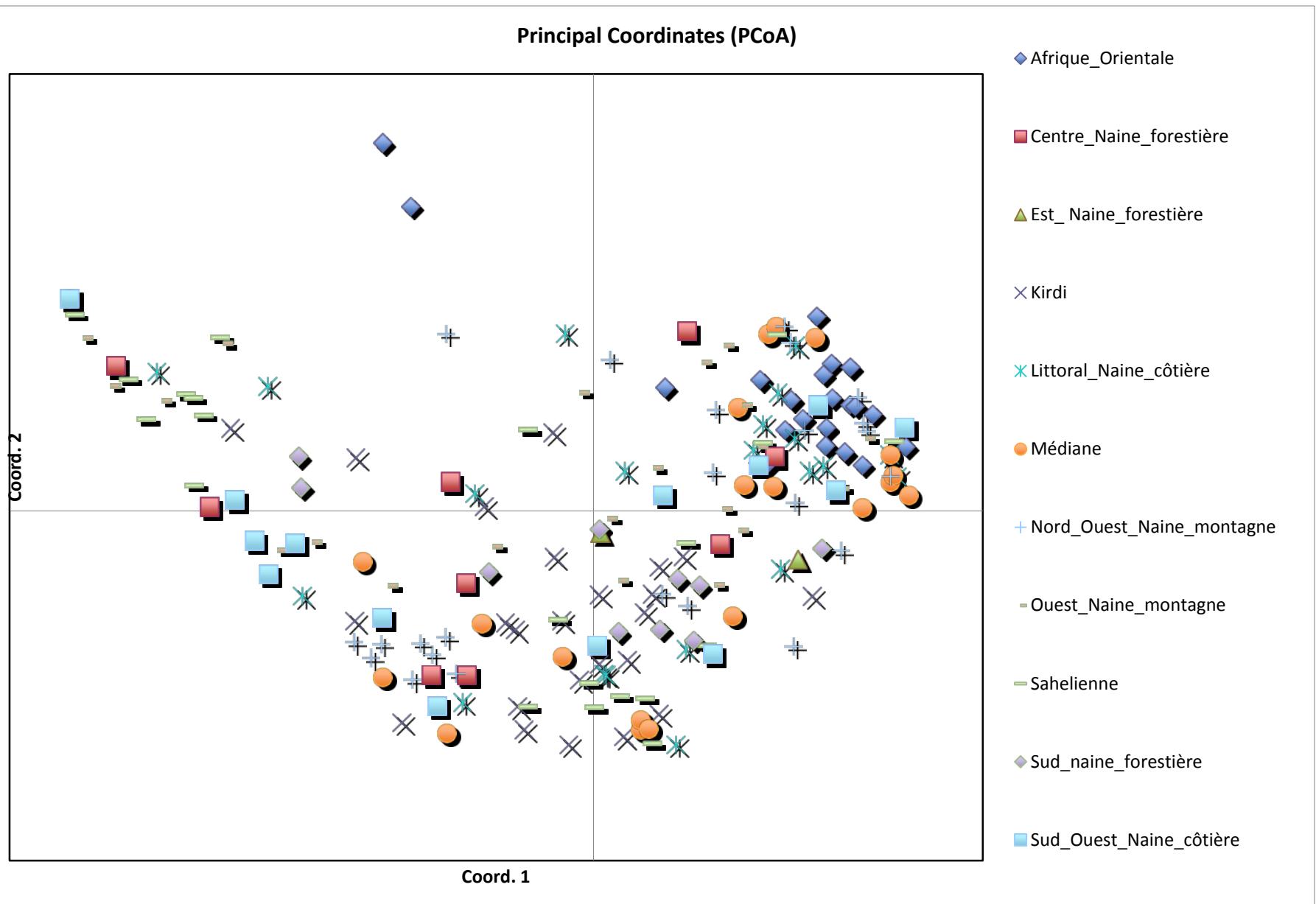


Figure 4: Principal Component Analysis in Cameroon goat population

Results

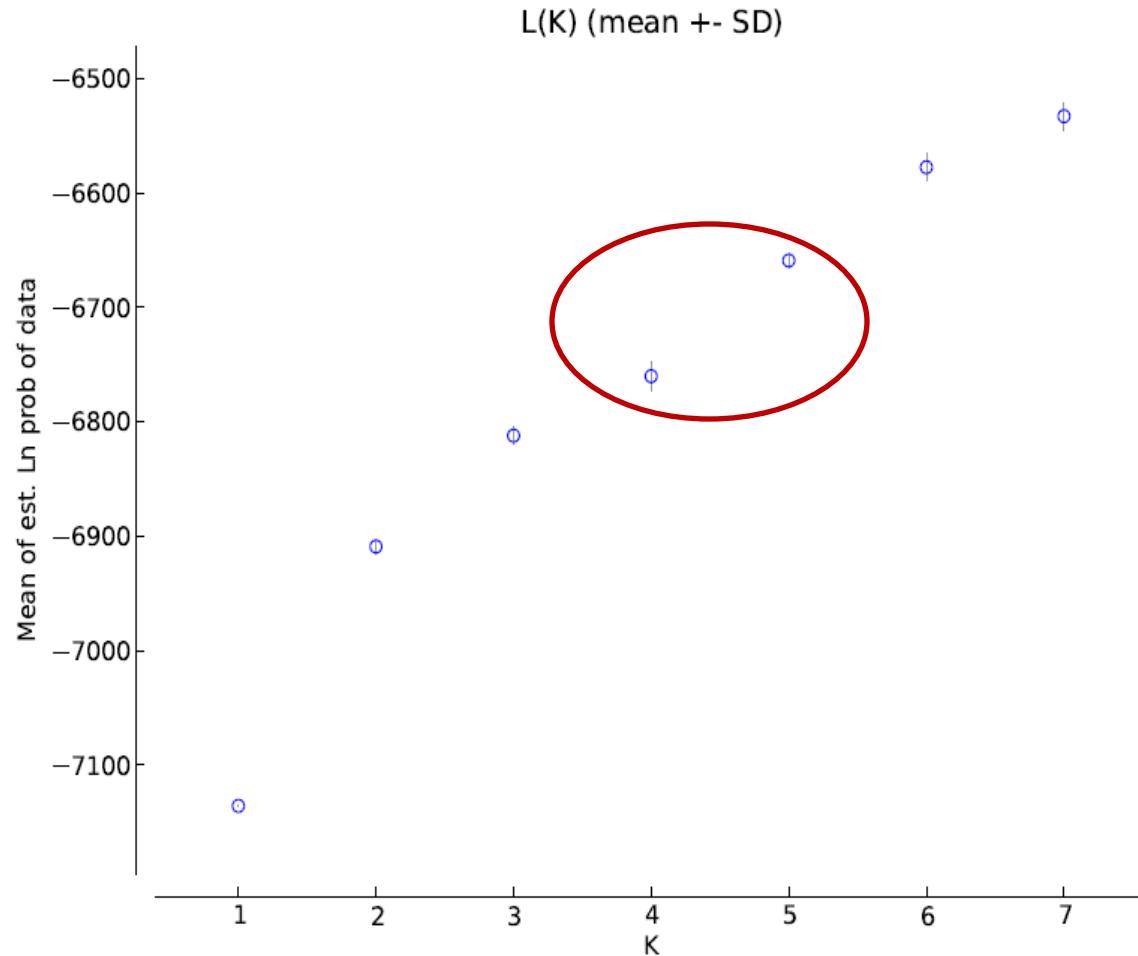


Figure 5 : Estimation of Cameroon population structure by Evanno's procedure

Results

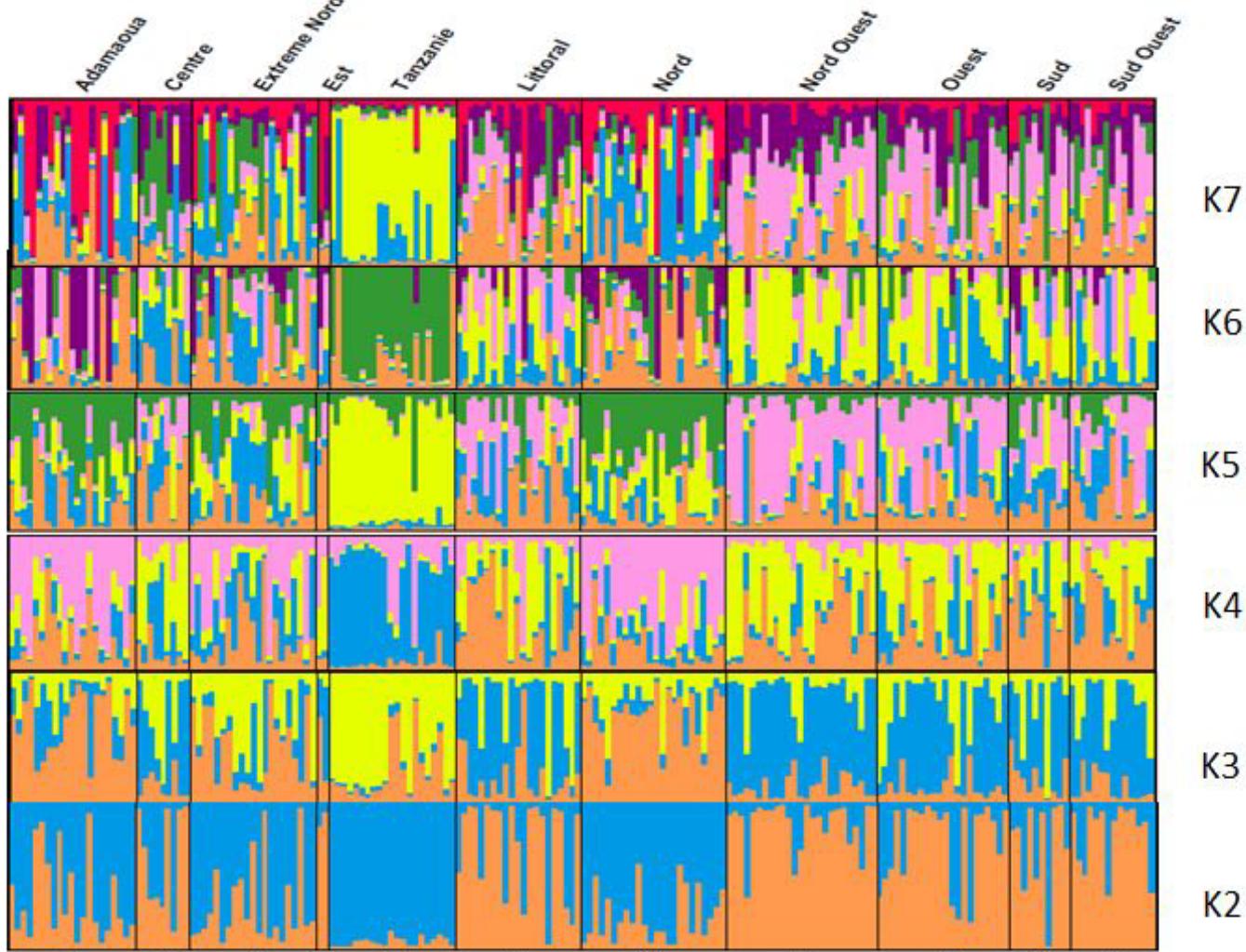


Figure 6 : Admixture trend in Cameroon goat population

Conclusion

- All 17 SSR are polymorphic in different Cameroon goat ecotypes.
- Results suggest variability among Cameroon goat population
- At least 4 putative populations exist
- Admixture tendency => high

Further analysis and more specific SSR applications are needed.

Some measures are needed to avoid massive dilution of native goat population

Acknowledgements

- The BecA/ILRI-CSIRO partnership
- Small Ruminant Support Program - Cameroon
- AusAID for funding the research



Thank You!!!