



### Farmed animal genomes – status and where next

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### Acknowledgements

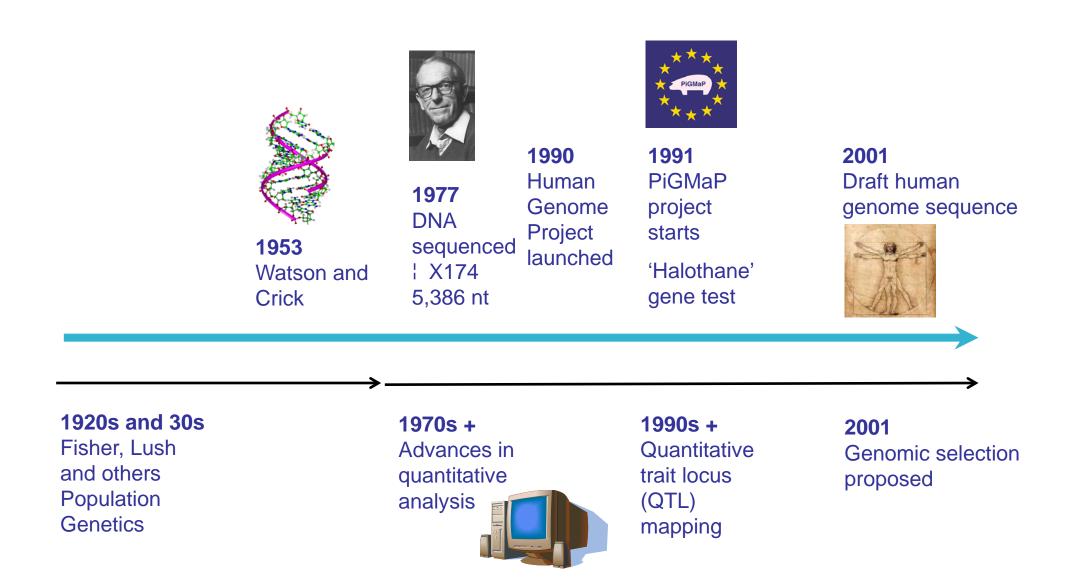
- Swine Genome Sequencing Consortium
- International Sheep Genome Consortium
- Chicken and other avian genome consortia
- Many others
- Funders
  - Many sources, including
    - EC FP7 Quantomics-222664
    - EC FP7 3SR-245140





### **From Sequence to Consequence**

### **Tools for the Exploitation of Livestock Genomes**









# Reference genome sequence as a key resource and framework for biological research

- Genetics
  - Variation (SNPs, indels, CNVs)
    - SNP chips, Genotype-by-Sequence
  - Genome-Wide Association Studies (GWAS)
  - Genetic improvement
- Functional genomics
  - incl. physiology, immunology,.....
  - Genome-wide analysis of responses to perturbation
    - Gene expression, methylation, ......
    - Microarrays, Assay-by-sequence

#### 2003

2002 sequence Mouse "finished" draft \$3 billion genome sequence



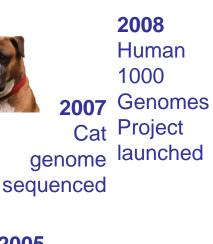
Human genome 2004 Chicken genome sequenced 2005



Dog

genome

sequenced



2010 Turkey genome sequenced

2009 Cattle genome sequenced Horse genome

Sequenced

Mouse genome "finished"





2009 2008 Pig 60K SNP Bovine 50K chip SNP chip

> Sheep 60K **SNP** chip



2010 750K bovine SNP chips





**2013** Goat genome sequenced



**2013** Duck genome sequenced



**2013 onwards** Animal ENCODE

**2013 onwards** Genotype-bysequence

2012 Chicken 600K SNP chip



2013 Salmon SNP chip

#### Fish: Tilapia, Cod, Salmon,.....





**2012** Pig genome sequenced

**2012** Pig gene expression atlas

BMC Biology

**2013** Sheep genome sequenced





### **Genome variation**

- From a reference to multiple genomes per species
- Enables
  - Discovery of SNP and structural variation (& SNP chips)
  - Analysis of natural and artificial selection
  - Identification of causal variation
- Visualising genetic variation (Ensembl Variation)
- Predicting consequences (e.g. Variant Effect Predictor, SIFT,...)

# **Multiple genomes**

- Human 1000 Genomes Project
  - ~4-6x coverage / individual
  - revealing genetic burden
  - ~1-200 potential Loss of Function mutations per person
- Human genetics studies
  - 10's of thousands per study
  - ICQG 2012
    - 30K sequenced genomes in a study

# **Multiple animal genomes**

- Pooled samples
  - 10-15x coverage
  - Chickens, cattle, pigs
  - SNP discovery
  - Signatures of selection
  - Signatures of domestication
- Individual genomes
  - 4-10x coverage
  - €3,000 per genome

# **Multiple animal genomes**

- 1000 Bull Genomes Project
  - Collaborative, Cloud data repository
  - Nnn bulls, average coverage ~11x
  - Data analysis cycles for genomic prediction
- Pigs
  - Groenen (Wageningen) ~300 individual pigs
  - Korean ~60 individual pigs
  - China ?? Pigs
- Sheep
  - ISGC 75 individual sheep
- Chickens
  - 10's of individuals (e.g. 10 individual J line brown egg layers)

# Visualising genome variation

#### Variation

What can I find? Short sequence variants and longer structural variants; disease and other phenotypes



More about variation in Ensembl





Variant Effect Predictor



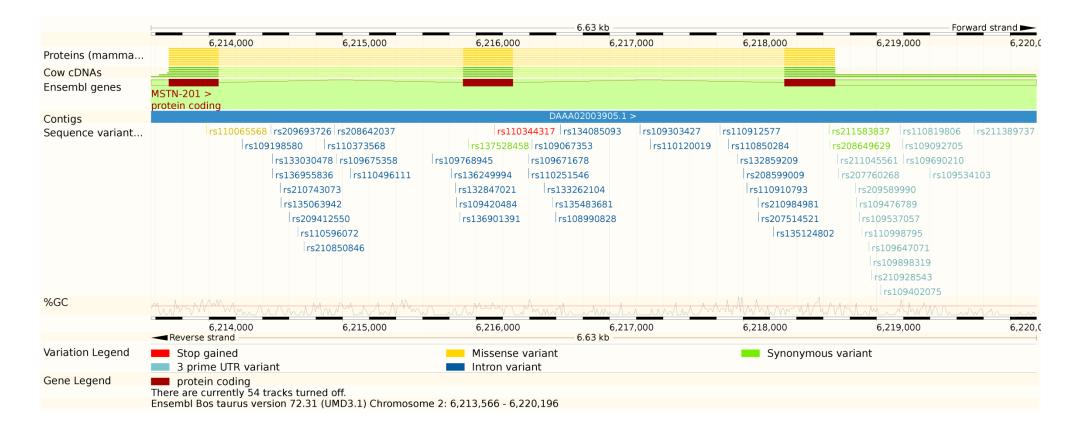


Example variant



Example phenotype

# **GDF8 - SNPs**



# **DGAT1 - variation**

#### Summary of variation consequences in ENSBTAG0000026356

Show All v entries					Filter
Number of variant consequences				Туре	Description
	0	-		Transcript ablation	A feature ablation whereby the deleted region includes a transcript feature (SO:0001893)
	0	-		Splice donor variant	A splice variant that changes the 2 base region at the 5' end of an intron (SO:0001575)
	0	-		Splice acceptor variant	A splice variant that changes the 2 base region at the 3' end of an intron (SO:0001574)
	0	-	•	Stop gained	A sequence variant whereby at least one base of a codon is changed, resulting in a premature stop codon, leading to a shortened transcript (SO:0001587)
	0	-		Frameshift variant	A sequence variant which causes a disruption of the translational reading frame, because the number of nucleotides inserted or deleted is not a multiple of three (SO:0001589)
	0	-	•	Stop lost	A sequence variant where at least one base of the terminator codon (stop) is changed, resulting in an elongated transcript (SO:0001578)
	0	-		Initiator codon variant	A codon variant that changes at least one base of the first codon of a transcript (SO:0001582)
	0	-		Transcript amplification	A feature amplification of a region containing a transcript (SO:0001889)
	0	-		Inframe insertion	An inframe non synonymous variant that inserts bases into in the coding sequence (SO:0001821)
	0	-		Inframe deletion	An inframe non synonymous variant that deletes bases from the coding sequence (SO:0001822)
	5	Show	v	Missense variant	A sequence variant, that changes one or more bases, resulting in a different amino acid sequence but where the length is preserved (SO:0001583)
	1	Show	v	Splice region variant	A sequence variant in which a change has occurred within the region of the splice site, either within 1-3 bases of the exon or 3-8 bases of the intron (SO:0001630)
	0	-		Incomplete terminal codon variant	A sequence variant where at least one base of the final codon of an incompletely annotated transcript is changed (SO:0001628)
	1	Show	v	Synonymous variant	A sequence variant where there is no resulting change to the encoded amino acid (SO:0001819)
	0	-		Stop retained variant	A sequence variant where at least one base in the terminator codon is changed, but the terminator remains (SO:0001567)
	0	-		Coding sequence variant	A sequence variant that changes the coding sequence (SO:0001580)
	0	-		Mature miRNA variant	A transcript variant located with the sequence of the mature miRNA (SO:0001620)
	0	-		5 prime UTR variant	A UTR variant of the 5' UTR (SO:0001623)

Switch to tree view 2

# **DGAT1 – missense variants**

#### Missense variant consequences 🗉

#### [back to top]

Show/hide of	columns							Fi	lter	
D	Chr: bp	Alleles	Class	Source	Evidence	Туре	AA	AA co- ord	<u>SIFT</u>	Transcript
rs134083952	14:1803973	T/C	SNP	dbSNP	ī	Missense variant	F/L	370	0	ENSBTAT00000 037423
rs135329220	14:1804495	T/G	SNP	dbSNP		Missense variant	V/G	464	0.01	ENSBTAT00000 037423
<u>rs137745035</u>	14:1801941	A/G	SNP	dbSNP		Missense variant	T/A	187	0.04	ENSBTAT00000 037423
rs109234250	14:1802265	G/A	SNP	dbSNP	6 😺	Missense variant	A/T	232	0.58	ENSBTAT00000 037423
rs109326954	14:1802266	C/A	SNP	dbSNP		Missense variant	A/E	232	1	ENSBTAT00000 037423

cf. K232E Grisart et al 2003 PNAS 101: 2398

flawed link





### **Genetic variation**

- Several important 'mutations' missing
  - RYR1 (HAL), MSTN/GDF8 (double muscling), DGAT1 (milk yield)
- Indels missing
- Predictions limitations





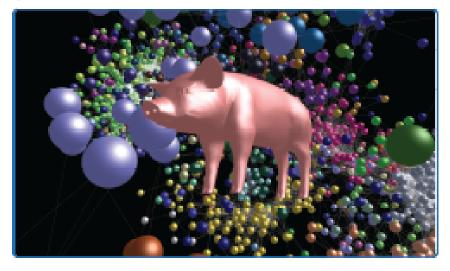
# Q

### **Gene expression**

• From sequence to consequence

- From microarrays to RNAseq
- Expression atlases
  - Pig: microarray (published), RNAseq (in progress)
  - Sheep: RNAseq (in progress)
  - Chicken: RNAseq (partial)





#### A gene expression atlas of the domestic pig

Freeman et d.

Freeman et al. BMC Biology 2012, 10:90 http://www.biomedcentral.com/1741-7007/10/90



#### RESEARCH ARTICLE

**Open Access** 

#### A gene expression atlas of the domestic pig

Tom C Freeman<sup>1\*</sup>, Alasdair Ivens<sup>26</sup>, J Kenneth Baillie<sup>1</sup>, Dario Beraldi<sup>1,7</sup>, Mark W Barnett<sup>1</sup>, David Dorward<sup>1</sup>, Alison Downing<sup>1</sup>, Lynsey Fairbairn<sup>1</sup>, Ronan Kapetanovic<sup>1</sup>, Sobia Raza<sup>1</sup>, Andru Tomoiu<sup>1</sup>, Ramiro Alberio<sup>3</sup>, Chunlei Wu<sup>4</sup>, Andrew I Su<sup>4</sup>, Kim M Summers<sup>1</sup>, Christopher K Tuggle<sup>5</sup>, Alan L Archibald<sup>1\*</sup> and David A Hume<sup>1\*</sup>

#### Abstract

**Background:** This work describes the first genome-wide analysis of the transcriptional landscape of the pig. A new porcine Affymetrix expression array was designed in order to provide comprehensive coverage of the known pig transcriptome. The new array was used to generate a genome-wide expression atlas of pig tissues derived from 62 tissue/cell types. These data were subjected to network correlation analysis and clustering.

**Results:** The analysis presented here provides a detailed functional clustering of the pig transcriptome where transcripts are grouped according to their expression pattern, so one can infer the function of an uncharacterized gene from the company it keeps and the locations in which it is expressed. We describe the overall transcriptional signatures present in the tissue atlas, where possible assigning those signatures to specific cell populations or pathways. In particular, we discuss the expression signatures associated with the gastrointestinal tract, an organ that was sampled at 15 sites along its length and whose biology in the pig is similar to human. We identify sets of genes that define specialized cellular compartments and region-specific digestive functions. Finally, we performed a network analysis of the transcription factors expressed in the gastrointestinal tract and demonstrate how they sub-divide into functional groups that may control cellular gastrointestinal development.

**Conclusions:** As an important livestock animal with a physiology that is more similar than mouse to man, we provide a major new resource for understanding gene expression with respect to the known physiology of mammalian tissues and cells. The data and analyses are available on the websites http://biogps.org and http://www.macrophages.com/pig-atlas.

Keywords: pig, porcine, Sus scrofa, microarray, transcriptome, transcription network, pathway, gastrointestinal tract



# Expression array, atlas

- Tool for monitoring gene expression
- Inferring function of unknowns
  - Inform genome annotation
- Comparative functional genomics
  - Is pig kidney more/less like human kidney than mouse kidney?
- Microarray-based atlas
- RNAseq atlas in progress





### Affymetrix Porcine Snowball Array content

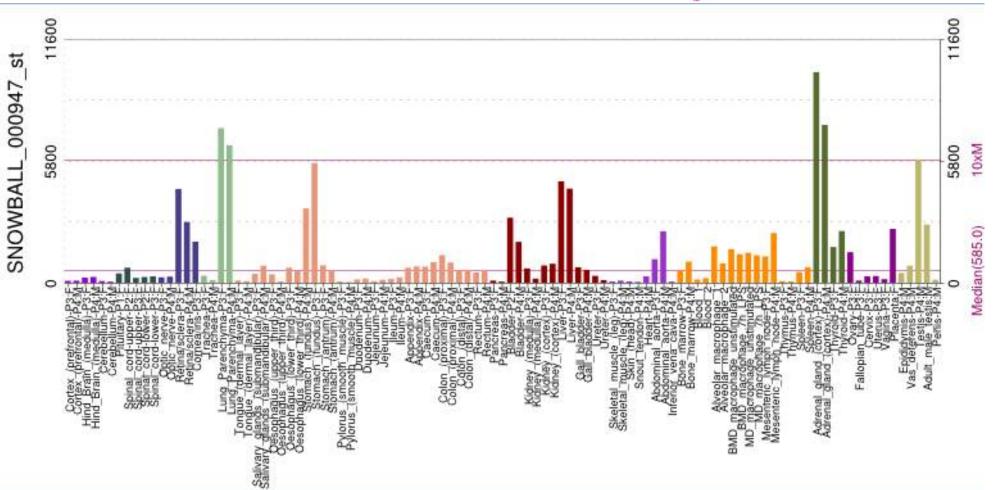


- 123 Affy controls
- 35 virus genomes (tiled 17 bp spacing)
- 1,857 miRNA probes
- 37 MT-mRNA
- 45,927 mRNA
  - 37,842 with annotation
    - 6,767 LOC annotations
    - 16,626 unique genes with official symbol/description









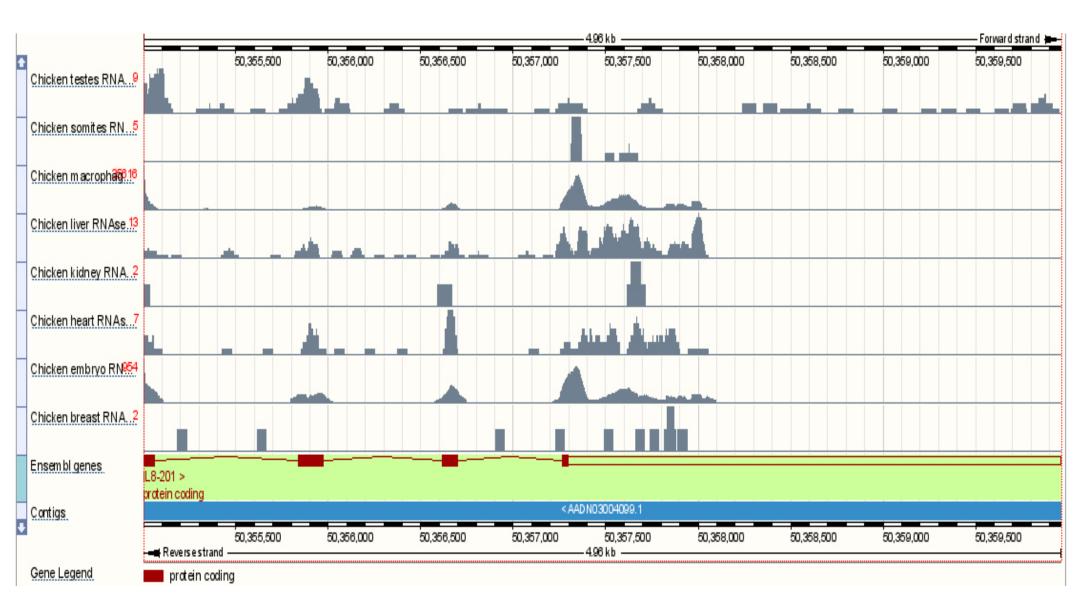
**Expression profiles** 



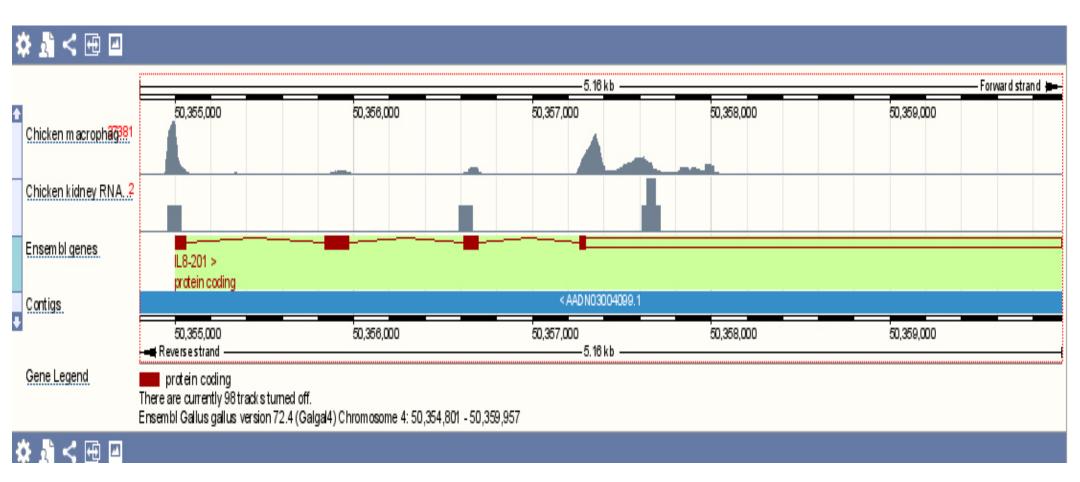
### http://biogps.org



# Chicken IL8 – RNAseq profiles



# Chicken IL8 – RNAseq profiles



# Sheep gene atlas

- Texel (ram, ewe, ewe lamb, 16d embryo)
- 50+ tissues per animal, whole embryo
  - Samples acquired, RNA prepared
- Illumina paired ends (2 x 150 bp)
  - > 1 Tb RNAseq data
- Ensembl RNAseq gene models (in progress)
- Funded 3SR, RoslinFoundation
- Poster 419





Cerebrum	Abomasum	Skeletal muscle, biceps	Testes, epididymis
Brain stem	Rumen	Skeletal muscle, longissimus dorsi	Corpus luteum, ovary, ovarian follicles
Tonsil	Duodenum	Skin (side/back)	Uterus, cervix, placenta
Cerebellum	Omentum	Spleen	Mammary gland
Hypothallamus	Caecum	Mesenteric lymph node	
Pituitary gland	Colon	Precapuslar lymph node	
Adrenal gland	Rectum	Peyer's patch	Whole embryo
Thyroid gland		Alveolar macrophages	
		Bone marrow	

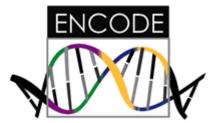




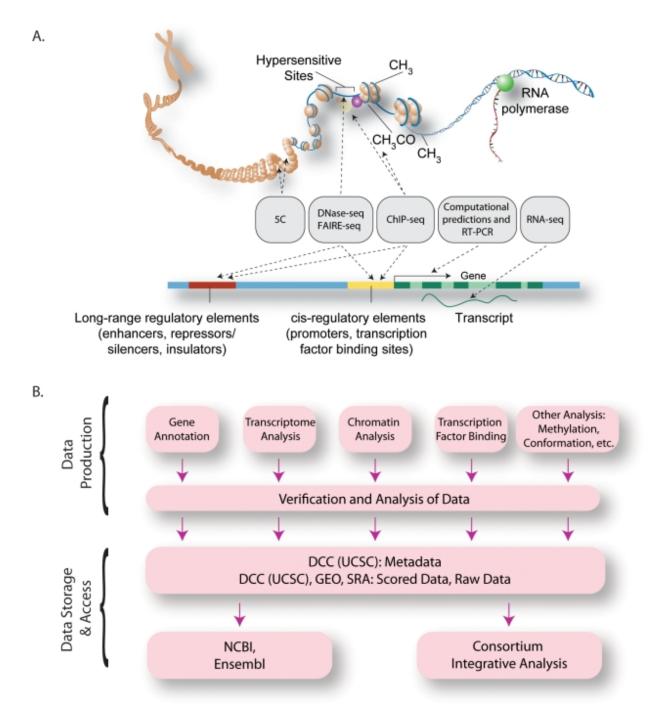
### Enabling the reading of farmed animal genomes

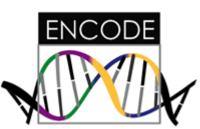
- Annotation of functional sequences
  - Protein coding
  - Non-coding RNA sequences
  - Regulatory sequences
- cf. human ENCODE project
  - <u>Encyclopedia of DNA elements in the human genome</u>

# ENCODE



#### ENCODE **Encyclopedia of DNA Elements** About ENCODE Data Human Data The Encyclopedia of DNA Elements (ENCODE) Consortium is an international collaboration c Summary Research Institute (NHGRI). The goal of ENCODE is to build a comprehensive parts list of fun that act at the protein and RNA levels, and regulatory elements that control cells and circumstant Search Downloads ENCODE data are now available for the entir available for immediate use via : Genome Browser **RNA** polymerase <u>Search</u> for displayable tracks and down (hg18) Download of data files Genome





## ARTICLE

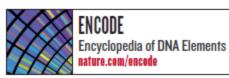
doi:10.1038/nature11247

# An integrated encyclopedia of DNA elements in the human genome

The ENCODE Project Consortium\*

The human genome encodes the blueprint of life, but the function of the vast majority of its nearly three billion bases is unknown. The Encyclopedia of DNA Elements (ENCODE) project has systematically mapped regions of transcription, transcription factor association, chromatin structure and histone modification. These data enabled us to assign biochemical functions for 80% of the genome, in particular outside of the well-studied protein-coding regions. Many discovered candidate regulatory elements are physically associated with one another and with expressed genes, providing new insights into the mechanisms of gene regulation. The newly identified elements also show a statistical correspondence to sequence variants linked to human disease, and can thereby guide interpretation of this variation. Overall, the project provides new insights into the organization and regulation of our genes and genome, and is an expansive resource of functional annotations for biomedical research.

The human genome sequence provides the underlying code for human biology. Despite intensive study, especially in identifying protein-coding genes, our understanding of the genome is far from complete, particularly with



95% of the genome lies within 8 kilobases (kb) of a DNA-protein interaction (as assayed by bound ChIP-seq motifs or DNase I footprints), and 99% is within 1.7 kb of at least one of the biochemical events measured by ENCODE.

# Headlines

- It is possible to correlate quantitatively RNA sequence production and processing with both chromatin marks and transcription factor binding at promoters
- indicating that promoter functionality can explain most of the variation in RNA expression

# Headlines

- Single nucleotide polymorphisms (SNPs) associated with disease by GWAS are enriched within non-coding functional elements, with a majority residing in or near ENCODE-defined regions that are outside of protein-coding genes.
- In many cases, the disease phenotypes can be associated with a specific cell type or transcription factor.
  - See also Hindorff et al 2009 PNAS 106: 9362
    - 88% of trait associated SNPs are intronic / intergenic





### **ENCODE for farmed animals - Why?**

- Understanding the genetic control of complex traits
- From sequence to consequence







### **ENCODE for farmed animals**

- Genetic variation underlying trait variation
  - Coding sequence
    - RYR1, DGAT1,
  - Regulatory sequence
    - IGF2, callipyge
    - likely to be more important / common
- Current annotation limted
  - cDNA, EST-based gene models (now RNAseq models too)
  - SNP variation

## **ENCODE for farmed / companion animals**

### How

- By-product of biology-led research
  - development, differentiation, responses to perturbation
- Focus on target tissues
  - musco-skeletal
  - immune tissues
- Limited assays
  - DNasel, FAIREseq
  - histone marks (promoters, enhancers)
  - methylation
  - RNAseq (stranded), CAGE

Histone modification or variant	Signal characteristics	Putative functions
H2AZ	Peak	Histone protein variant (H2A.Z) associated with regulatory elements with dynamic chromatin
H3K4me1	Peak/region	Mark of regulatory elements associated with enhancers and other distal elements, but also enriched downstream of transcription starts
H3K4me2	Peak	Mark of regulatory elements associated with promoters and enhancers
H3K4me3	Peak	Mark of regulatory elements primarily associated with promoters/transcription starts
H3K9ac	Peak	Mark of active regulatory elements with preference for promoters
H3K9me1	Region	Preference for the 5' end of genes
H3K9me3	Peak/region	Repressive mark associated with constitutive heterochromatin and repetitive elements
H3K27ac	Peak	Mark of active regulatory elements; may distinguish active enhancers and promoters from their inactive counterparts
H3K27me3	Region	Repressive mark established by polycomb complex activity associated with repressive domains and silent developmental genes
H3K36me3	Region	Elongation mark associated with transcribed portions of genes, with preference for 3' regions after intron 1
H3K79me2	Region	Transcription-associated mark, with preference for 5' end of genes
H4K20me1	Region	Preference for 5' end of genes

#### Table 2 | Summary of ENCODE histone modifications and variants

6 SEPTEMBER 2012 | VOL 489 | NATURE | 59

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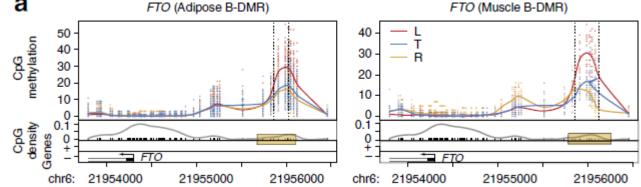
#### ARTICLE

Received 7 Oct 2011 Accepted 19 Apr 2012 Published 22 May 2012

DOI: 10.1038/ncomms1854

### An atlas of DNA methylomes in porcine adipose and muscle tissues

Mingzhou Li<sup>1,\*</sup>, Honglong Wu<sup>2,\*</sup>, Zonggang Luo<sup>1,3</sup>, Yudong Xia<sup>2</sup>, Jiuqiang Guan<sup>1</sup>, Tao Wang<sup>1</sup>, Yiren Gu<sup>4</sup>, Lei Chen<sup>5</sup>, Kai Zhang<sup>1,†</sup>, Jideng Ma<sup>1</sup>, Yingkai Liu<sup>1</sup>, Zhijun Zhong<sup>1</sup>, Jing Nie<sup>1</sup>, Shuling Zhou<sup>1</sup>, Zhiping Mu<sup>1</sup>, Xiaoyan Wang<sup>1</sup>, Jingjing Qu<sup>1</sup>, Long Jing<sup>1</sup>, Huiyu Wang<sup>1</sup>, Shujia Huang<sup>2</sup>, Na Yi<sup>2</sup>, Zhe Wang<sup>2</sup>, Dongxing Xi<sup>2</sup>, Juan Wang<sup>2</sup>, Guangliang Yin<sup>2</sup>, Li Wang<sup>2</sup>, Ning Li<sup>2</sup>, Zhi Jiang<sup>2</sup>, Qiulei Lang<sup>6</sup>, Huasheng Xiao<sup>7</sup>, Anan Jiang<sup>1</sup>, Li Zhu<sup>1</sup>, Yanzhi Jiang<sup>1</sup>, Guoqing Tang<sup>1</sup>, Miaomiao Mai<sup>1</sup>, Surong Shuai<sup>1</sup>, Ning Li<sup>8</sup>, Kui Li<sup>9</sup>, Jinyong Wang<sup>5</sup>, Xiuqing Zhang<sup>2</sup>, Yingrui Li<sup>2</sup>, Haosi Chen<sup>10</sup>, Xiaolian Gao<sup>10</sup>, Graham S. Plastow<sup>11</sup>, Stephen Beck<sup>12</sup>, Huanming Yang<sup>2</sup>, Jian Wang<sup>2</sup>, Jun Wang<sup>2</sup>, Xuewei Li<sup>1</sup> & Ruiqiang Li<sup>2,†</sup> **a** *FTO* (Adipose B-DMR)



## Cell

Volume 149, Issue 6, 8 June 2012, Pages 1381-1392

#### Resource

### Comparative Epigenomic Annotation of Regulatory DNA

Shu Xiao<sup>1, 2, 6</sup>, Dan Xie<sup>1, 2, 6</sup>, Xiaoyi Cao<sup>2, 3, 6</sup>, Pengfei Yu<sup>2, 3, 6</sup>, Xiaoyun Xing<sup>6</sup>, Chieh-Chun Chen<sup>1, 2</sup>, Meagan Musselman<sup>1</sup>, Mingchao Xie<sup>6</sup>, Franklin D. West<sup>4</sup>, Harris A. Lewin<sup>2</sup>, Ting Wang<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Mingchao Xie<sup>6</sup>, Franklin D. West<sup>4</sup>, Harris A. Lewin<sup>2</sup>, Ting Wang<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Mingchao Xie<sup>6</sup>, Franklin D. West<sup>4</sup>, Harris A. Lewin<sup>2</sup>, Ting Wang<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Mingchao Xie<sup>6</sup>, Franklin D. West<sup>4</sup>, Harris A. Lewin<sup>2</sup>, Ting Wang<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Mingchao Xie<sup>6</sup>, Franklin D. West<sup>4</sup>, Harris A. Lewin<sup>2</sup>, Ting Wang<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Mingchao Xie<sup>6</sup>, Franklin D. West<sup>4</sup>, Harris A. Lewin<sup>2</sup>, Ting Wang<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Mingchao Xie<sup>6</sup>, Franklin D. West<sup>4</sup>, Harris A. Lewin<sup>2</sup>, Ting Wang<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Mingchao Xie<sup>6</sup>, Franklin D. West<sup>4</sup>, Harris A. Lewin<sup>2</sup>, Ting Wang<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Xiao Ying<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Xiao Ying<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, Mingchao Ying<sup>6</sup>, Sheng Zhong<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Xiao Ying<sup>6</sup>, Sheng<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Xiao Ying<sup>6</sup>, Sheng<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Xiao Ying<sup>6</sup>, Sheng<sup>1, 2, 3</sup>, A. Wasselman<sup>1</sup>, Xiao Ying<sup>1, 2, 3</sup>, Xiao Ying<sup>1, 2, 3</sup>, Xiao Ying<sup>1, 2, 3</sup>, Xiao Ying<sup>1, 2, 3</sup>, Ying<sup>1, 2, 3</sup>, Xiao Ying<sup>1, 2, 3</sup>, Xiao

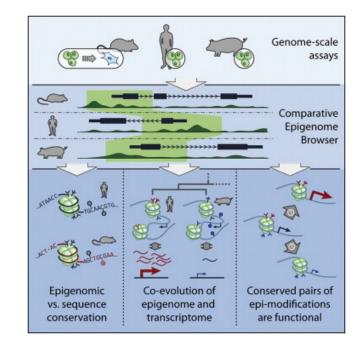
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<sup>1</sup> Department of Bioengineering, University of Illinois at Urbana-Champaign, 1304 West Springfield Avenue, Urbana, IL 61801, USA

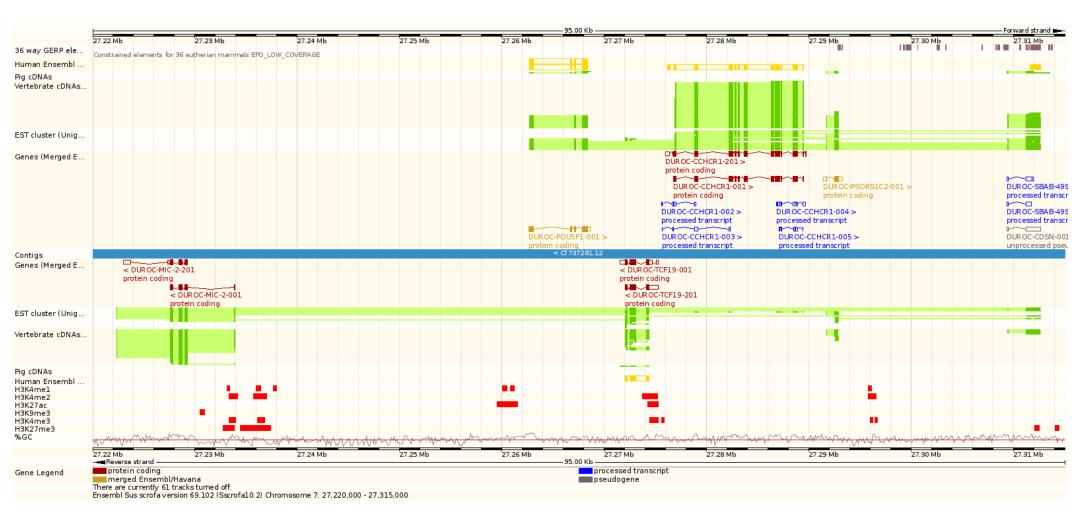
<sup>2</sup> Institute for Genomic Biology, University of Illinois at Urbana-Champaign, 1304 West Springfield Avenue, Urbana, IL 61801, USA

<sup>3</sup> Center for Biophysics and Computational Biology, University of Illinois at Urbana-Champaign, 1304 West Springfield Avenue, Urbana, IL 61801, USA

<sup>4</sup> Department of Animal and Dairy Science, University of Georgia, 425 River Road, Athens, GA 30602, USA



RNASeg genes Whole blood RNA <sup>434</sup>				F-B
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Placenta RNA Seg <sup>879</sup>				
Merged RNASeg 881	the state of the state	and a last	a with the former at	
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Alveolar macroph <sup>298</sup>				
10 tissues RNAse <sup>206</sup>	and the contract will the fill		untillate man	
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VEGA49 assembly Contigs		7:27200000-27320000 < CT737281.12		



# **ENCODE for farmed / companion animals**

- Species
  - single consortium / one per species / species groups
- Cells
  - transformed cells / primary cells / iPS cells
  - sharing
- Data management, publication
  - across groups
  - wider community
    - Data hubs model
  - Toronto Statement
- Coordination



## alan.archibald@roslin.ed.ac.uk

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# A User's Guide to the Encyclopedia of DNA Elements (ENCODE)

The ENCODE Project Consortium<sup>¶\*</sup>

### Abstract

The mission of the Encyclopedia of DNA Elements (ENCODE) Project is to enable the scientific and medical communities to interpret the human genome sequence and apply it to understand human biology and improve health. The ENCODE Consortium is integrating multiple technologies and approaches in a collective effort to discover and define the functional elements encoded in the human genome, including genes, transcripts, and transcriptional regulatory regions, together with their attendant chromatin states and DNA methylation patterns. In the process, standards to ensure high-quality data have been implemented, and novel algorithms have been developed to facilitate analysis. Data and derived results are made available through a freely accessible database. Here we provide an overview of the project and the resources it is generating and illustrate the application of ENCODE data to interpret the human genome.

## *PloS Biology* 2011 9: e1001046



### Data Standards

### **Guidelines for Experiments**

The ENCODE Consortium has adopted uniform guidelines for the most common ENCODE experiments. The guidelines have evolved over time, as technologies have changed. The current guidelines are informed by results gathered during the project. Previous versions of the standards are also posted for reference.

### **Platform Characterization**

ENCODE datasets are collected using a variety of platforms, such as ChIP-seq and RNA-seq. The consortium has undertaken several efforts to characterize these platforms to better understand the data being collected using them. Some of these efforts are described on this page.

### Quality Metrics<sup>New</sup>

The ENCODE consortium analyzes the quality of the data produced using a variety of metrics. To view the quality metrics for many ENCODE datasets, go to this page. The quality metrics will be updated on occasion to include more recent data. Note that antibody validation information can be found at ENCODE Antibodies. The Platform Characterization page, described above also examines an issue related to data quality.

Updated 04 June 2012

### ChIP-seq guidelines and practices of the ENCODE and modENCODE consortia

Stephen G. Landt, Georgi K. Marinov, Anshul Kundaje, et al.

Genome Res. 2012 22: 1813-1831

# **Next steps**

- White paper
- Develop data management strategy
- Review / promote ENCODE experimental protocols
- Develop / review cell line resources
- Develop communications strategy
- COST Action application (Sept 2013)
- US-EU ABWG workshop at PAG 2014





## Where next?

- Improving reference genomes\*
- Functional annotation (cf. ENCODE)\*
- Sequencing 1000's to millions of individuals
  - Genotyping-by-sequencing and imputation
- Genomic selection





# Acknowledgements

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- Ewan Birney
- Jen Harrow, Jane Loveland











# edinburgh genomics.



Coming soon!

Edinburgh Genomics will be a new facility formed from the merger of ARK Genomics and the GenePool, both world-class genomics facilites.

For further information, please e-mail us.

# http://genomics.ed.ac.uk

# edinburgh genomics.

- Merger of ARK-Genomics & Gene Pool
- Sequencing
  - Sanger: ABI3730
  - Illumina: 6x HiSeq2500, 3 MiSeq
- Bioinformatics
  - bioinformaticians
  - Edinburgh Parallel Computing Centre
    - secure multi terrabyte data store
    - secure compute Grid
- Genotyping, gene expression (ARK-Genomics)

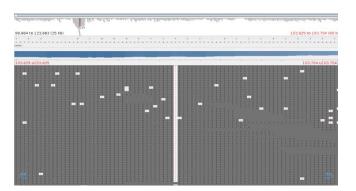
# edinburgh genomics.

## **DNA Sequencing**

Illumina Sequencing

- Up to 250 bp paired
- Novel genomes
- Resequencing
- RNA-Seq
- ChIP-Seq
- Epigenetics

Illumina HiSeq 2500 Illumina MiSeq Sanger 3730



## Genotyping

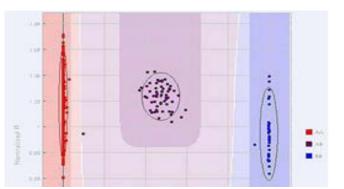
Illumina - from HD to

custom chips

- iScan, Inifinium
- BeadXpress, Goldengate
- BeadChip

### Affymetrix

- GeneTitan, Axiom
- Process 96 arrays / run



## Microarrays

Gene Expression

- Affymetrix
- Agilent
- Illumina
- Whole genome
- Exon-level
- microRNA

### CGH, ChIP-Chip, MeIP

• Nimblegen

