Modulation of the redox state in pigs differing in feed efficiency as revealed by a proteomic analysis

<u>F. Gondret¹</u>, S. Tacher¹ & H. Gilbert²

¹UMR Pegase, 35590 Saint Gilles, France ²UMR GenPhySe, 31326 Castanet-Tolosan, France

Florence.gondret@rennes.inra.fr



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RFI: ~ one third to half of the total variation in feed intake in growing pigs (*Knapp, 2009;* Young & Dekkers, 2012)



Variations in residual feed intake (RFI):

Feed consumption & behavior (number of visits, duration of meal intake, time spent eating, etc...)

Young et al., 2011 Meunier-Salaün et al., 2012

Digestion and utilization of nutrients

Barea et al., 2010

Organ physiology : thermoregulation basal metabolic rate energy efficiency of mitochondria muscle protein turnover rate immunity, etc.

Le Naou et al., 2012; Gabler et al., 2013; Grubbs et al., 2013





The liver is a complex and unique organ responsible for a breadth of functions that are crucial for sustaining life

We propose to use a proteomic differential approach to identify the modulation of **hepatic** metabolic processes participating **to feed efficiency**





Experimental design



RFI corresponded to a biologically imposed feed restriction

Results: Animal performance



A total of 50 protein spots in the liver had a differential abundance between pigs

Proteins were included in pathways related to AA metabolism, glycolysis/gluconeogenesis, ion binding, purine binding, and **oxido-reduction**

11 unique proteins belong to a top cluster related to cell oxido-reduction processes (enrichment score = 2.92)	
Response to oxidative stress	PRDX2, PRDX4, PRDX5, PRDX6, SOD1
Cell redox homeostasis	PRDX2, PRDX4, PRDX5, PRDX6, PDIA3, PDIA6, GSTO1
Oxidation reduction	PRDX2, PRDX4, PRDX5, PRDX6, SOD1, HAAO, DMGDH, ETFA

• Results: differential proteins in liver

Liver mitochondria and **Reactive Oxygen Species** (ROS) production

Tricarboxylic acids (TCA) cycle Fatty acid oxidation

Oxidation of reducing equivalents

Transfer of electrons along the respiratory chain



Liver mitochondria are a main site of ROS production



flavoprotein (ETFA) is considered as leading to ROS elevation

Levels in feed-restricted high RFI pigs were similar or intermediary to levels observed in low RFI pigs

Results: greater anti-oxidant enzymes in low RFI pigs

0.05

0

A low abundance of ETFA signs a low activity of the respiratory chain

A severe inhibition of the respiratory chain is considered as inhibiting lipid oxidation process in rats (*Pande, 1971*)



Lower fatty acid oxidation in the low RFI pigs, independently of feed intake

Results: lower lipid oxidation pathway in low RFI pigs

Balance in labile methyl groups (methionine, betaine, choline) is involved in maintaining normal lipid functions and health (*Mato et al., 2008*)



• Results: homocysteine/methionine/betaine equilibrium ?

Pigs selected for a low residual feed intake had eaten less during the growing period

Higher anti-oxidant capacities in the liver of the most efficient pigs, due to (?) greater ROS generation

Consequences on the redox state of the animals remains to be determined (other tissues)

Levels of anti-oxidants were lower with feed restriction.

Relationships between feed intake and oxidative stress/redox metabolism in pigs deserve further studies

caloric restriction in aging mice decreased hydrogen peroxide production from the mitochondria.

Conclusions

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