



# Using whole genome sequences to identify candidate mutations affecting Milk Fatty Acids in dairy cattle

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# Introduction

Complex traits are influenced by **many QTLs**, each explaining a small part of their variability

QTL full characterization is especially challenging and **only a few QTLs have been identified so far**, in spite of large efforts

New tools have become available:

- High throughput **genotyping** of large populations
- **Whole genome sequencing**

Is it now possible to re-address the question of QTL identification in a more efficient way ?

# Introduction



## Objectives

Identification of candidate **causal mutations**  
for milk fat composition

### *PhénoFinLait project*

8746 cows with milk fat  
composition & 50k genotypes

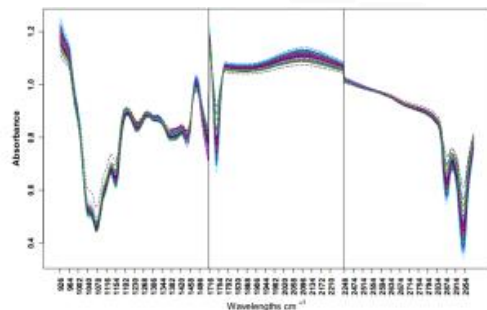
### *« 1000 bull genomes » project*

1147 bulls with whole genome  
sequences (RUN4)

Genome Wide Association Study (**GWAS**)  
at the **full sequence** level

# Material & methods: 23 Fatty acids estimated by MIR

## Mid-Infrared (MIR) spectra



Pre-correction of data  
for **non genetic** effects

Herd \* test-day

Month \* year of calving

Parity \* days in milk

### Total SAT

**C4:0**

C6:0

C8:0

C10:0

**C12:0**

C14:0

**C16:0**

C18:0

### Total UNSAT

Total MONO

**Total POLY**

**C18:1cis9**

C18:2cis9trans11

C18:1cis12

C18:2cis9cis12

C18:1t11t10

**C18:3n3**

TotC18:1

TotC18:3

TotC18:1cis

Omega 3

TotC18:1trans

Omega 6

# Material & methods: animals

~ 120,000 cows with phenotypes  
(~ 600,000 test-day milk samples)

8746 cows genotyped with the 50k Beadchip

**2882**

Montbéliardes

**MON**



**2816**

Normandes

**NOR**



**3048**

Holstein

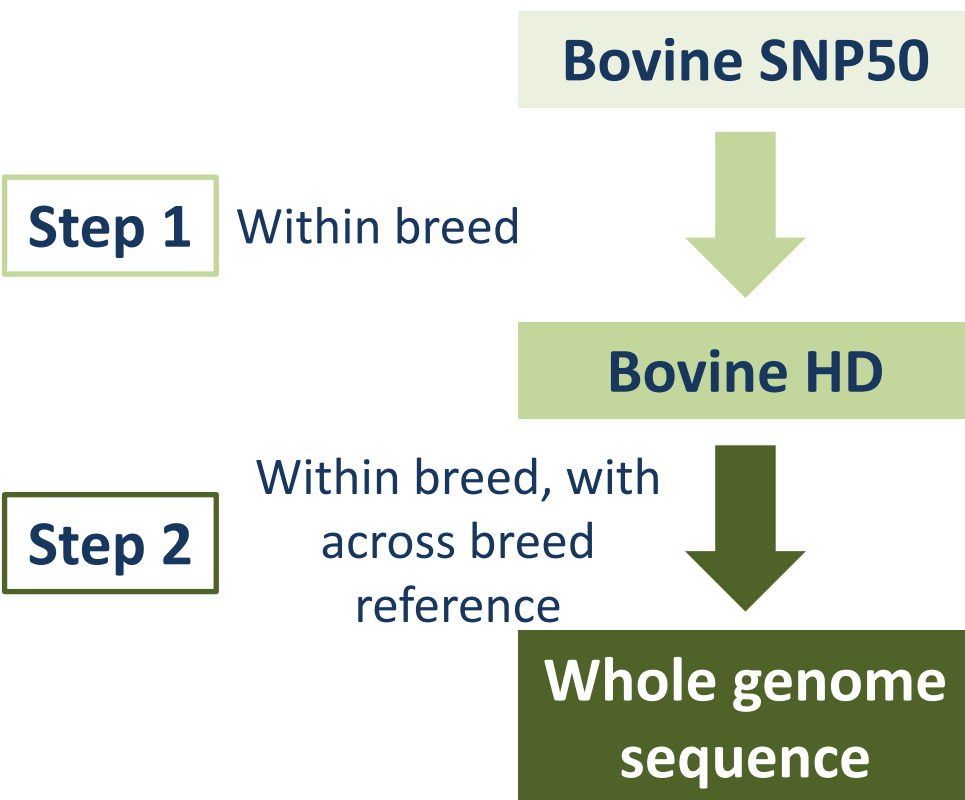
**HOL**



# Material & methods: genotypes & imputation

*Imputation in two steps with  
FImpute (Sargolzaei et al., 2014)*

*Reference populations*



**Within breed,  
HD genotyped bulls**  
522 MON  
546 NOR  
776 HOL

**1 Reference population**  
= 1147 bulls from  
« 1000 Bull Genomes »  
including  
28 MON + 24 NOR + 288 HOL

**27 millions** of sequence variants imputed for **8746** cows

# GWAS & Bayesian analyses



Within breed single marker **GWAS** with **GCTA** (Yang et al., 2011)  
**27 millions variants** , analyzed one at a time  
Polygenic effects of animals, GRM calculated from HD 631,000 SNP

Selection of the most  
interesting QTL regions



**Bayesian analyses (BayesC) with GS3** (Legarra et al., 2013)  
Within breed, Multimarker (up to 30,000 markers)  
Includes also a pedigree-based polygenic effect

## Bayesian analyses

Candidate variants were selected according to their **probability of inclusion** (based on 100,000 iterations, burn-in=20,000, thin = 50)

A **difficulty**: due to **very high LD**, inclusion probability of a region is distributed over many linked variants, and can be low for individual variants

Inclusion probabilities were **summed over 5kb windows** to detect the largest signals

Candidate variants were searched within the best windows

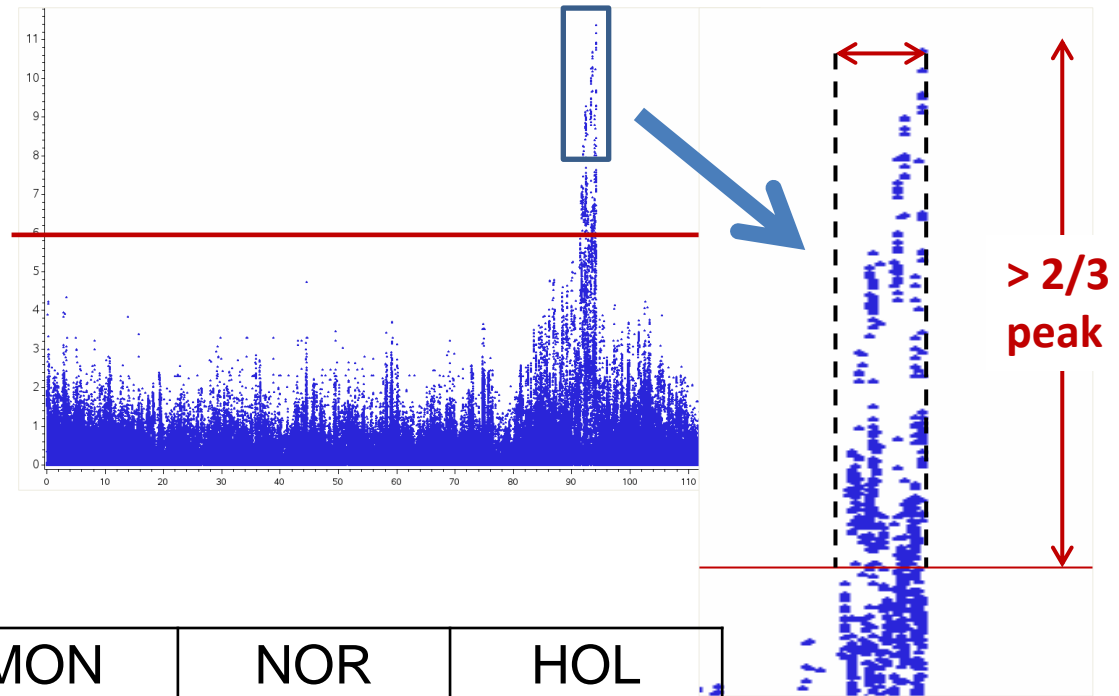
Complementary information : (1) **Across breed comparison**  
(2) **Variant Annotation** (1000 bull genomes)



# GWAS results

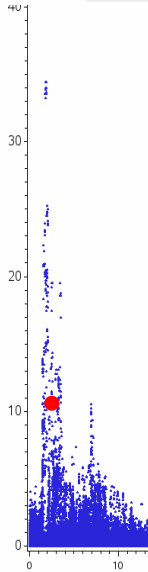
## Number of QTLs

- ✓  $-\log_{10}(p\_value) > 6$
- ✓ 1 QTL maximum in 2 Mb
- ✓ Drop-off value =  $\max(2, 2/3 \text{ peak})$



Trait	MON	NOR	HOL
C4:0	24	30	28
C12:0	24	18	31
SAT	23	11	26
MONO	22	15	20
$\omega 3$	25	21	44

# GWAS results: status of known mutations



**BTA14** DGAT1 HOL

C18:0

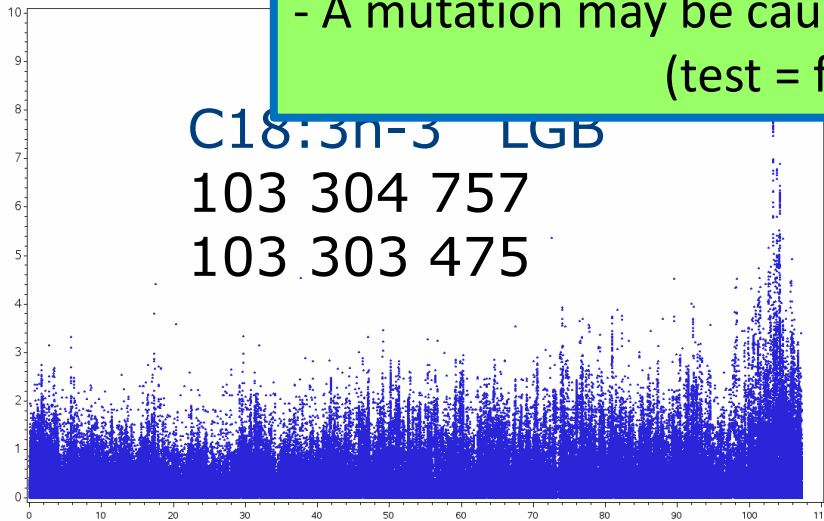
1 802 266

**BTA26** – SCD – MON

C12:0

21 144 708

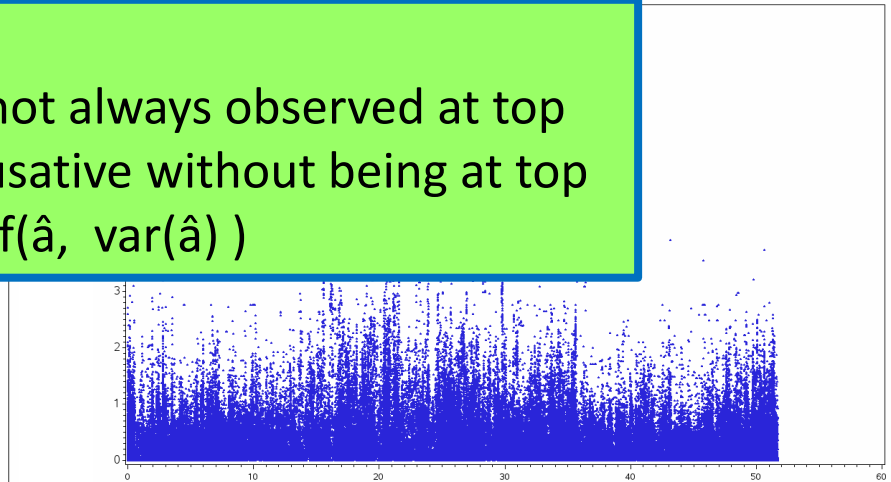
Difficult to conclude :  
- Known mutations are not always observed at top  
- A mutation may be causative without being at top  
(test =  $f(\hat{\alpha}, \text{var}(\hat{\alpha}))$ )



**BTA18** C18:3n-3 LGB

103 304 757

103 303 475



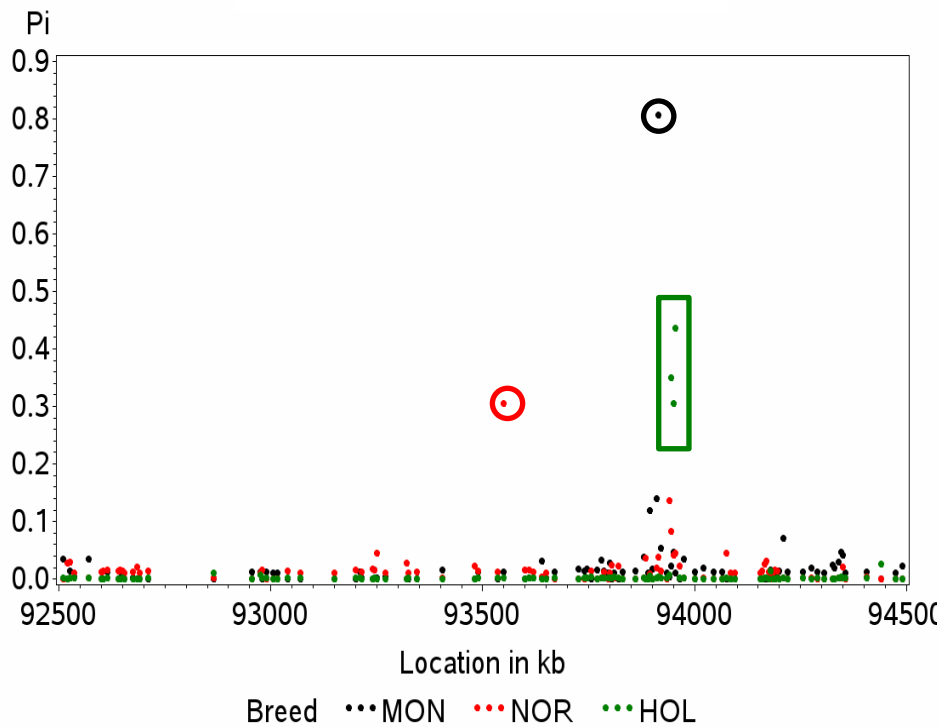
# Results: BayesC

The regions studied

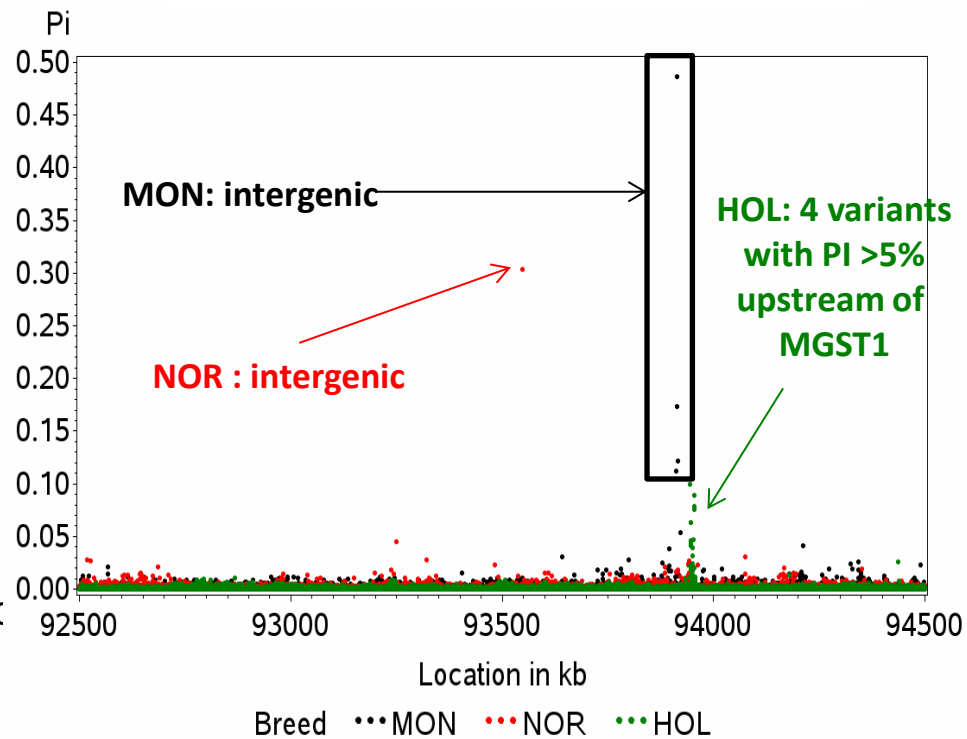
Chromosome	Région (Mb)	Trait	GWAS test $\text{Log}_{10}(1/p)$ max MON – NOR – HOL
5	92.5-94.5	SAT	14.2 – 13.2 – 24.4
14	1.3-3.8	SAT	34.4 – 79.9 – 169.8
17	52.5-55.0	C4:0	30.2 – 47.8 – 12.3
19	50.0-53.0	C12:0	28.2 – 15.2 – 38.8
27	36.0-36.5	C16:0	16.8 – 9.3 – 9.7

# Results: Chromosome 5, SAT

Sum over 5kb windows

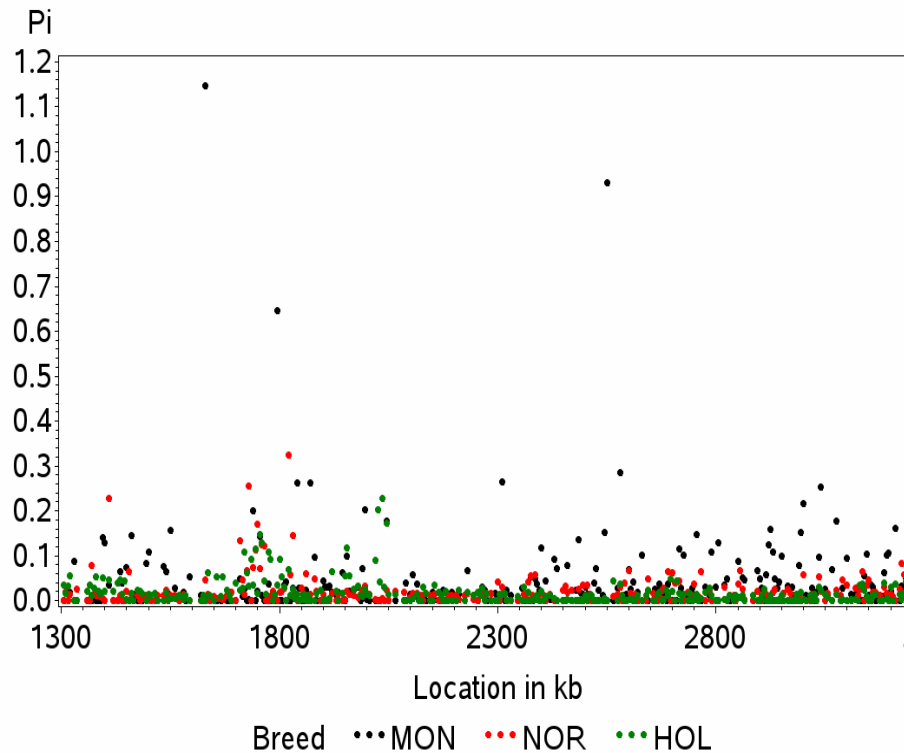


Inclusion probability for each SNP

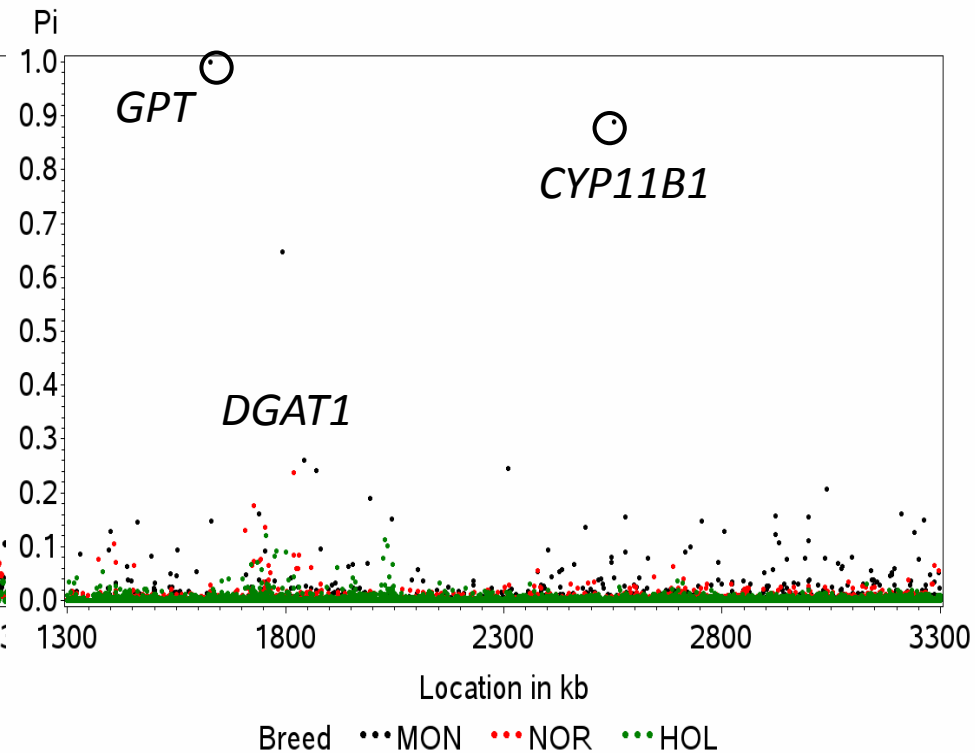


# Results: Chromosome 14, SAT

Sum over 5kb windows



Inclusion probability for each SNP

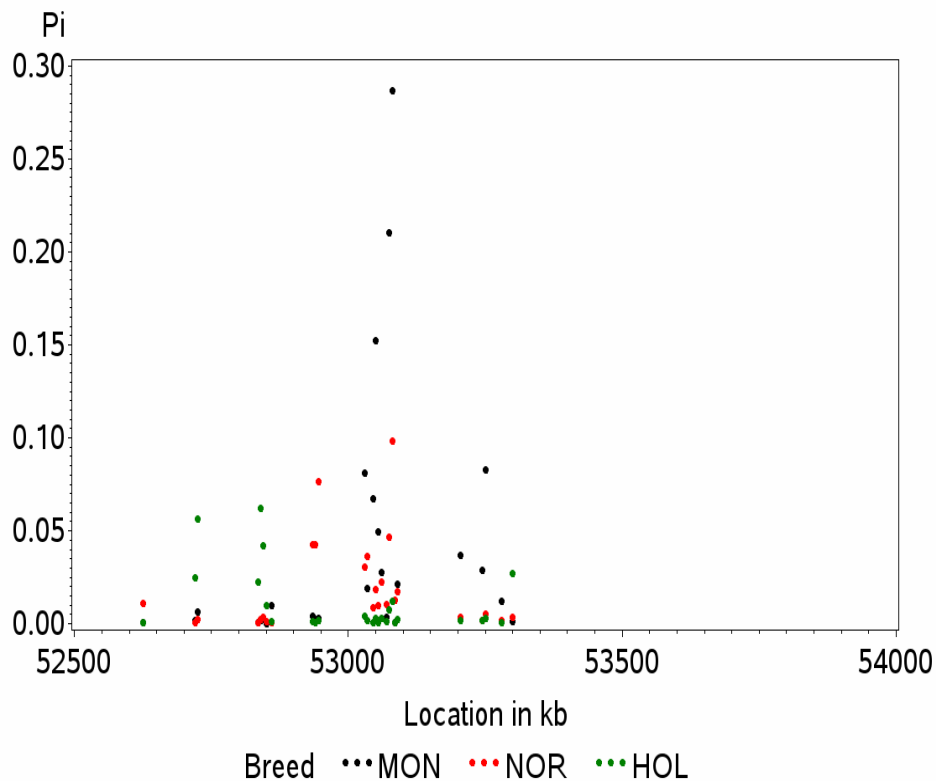


Other mutations than K232A suspected: QTL in populations fixed for K232A and in bulls homozygous for K232A

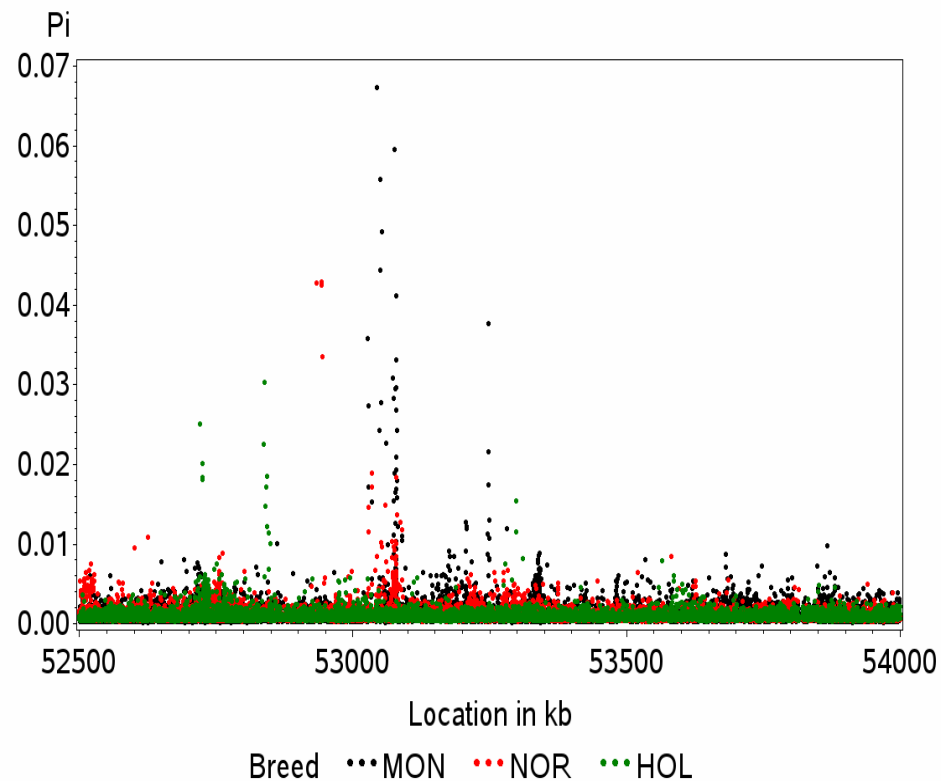
This region seems to be rich, with other mutations in DGAT1(4 ?) and in 2 other genes

# Results: Chromosome 17, C4:0

Sum over 5kb windows



Inclusion probability for each SNP

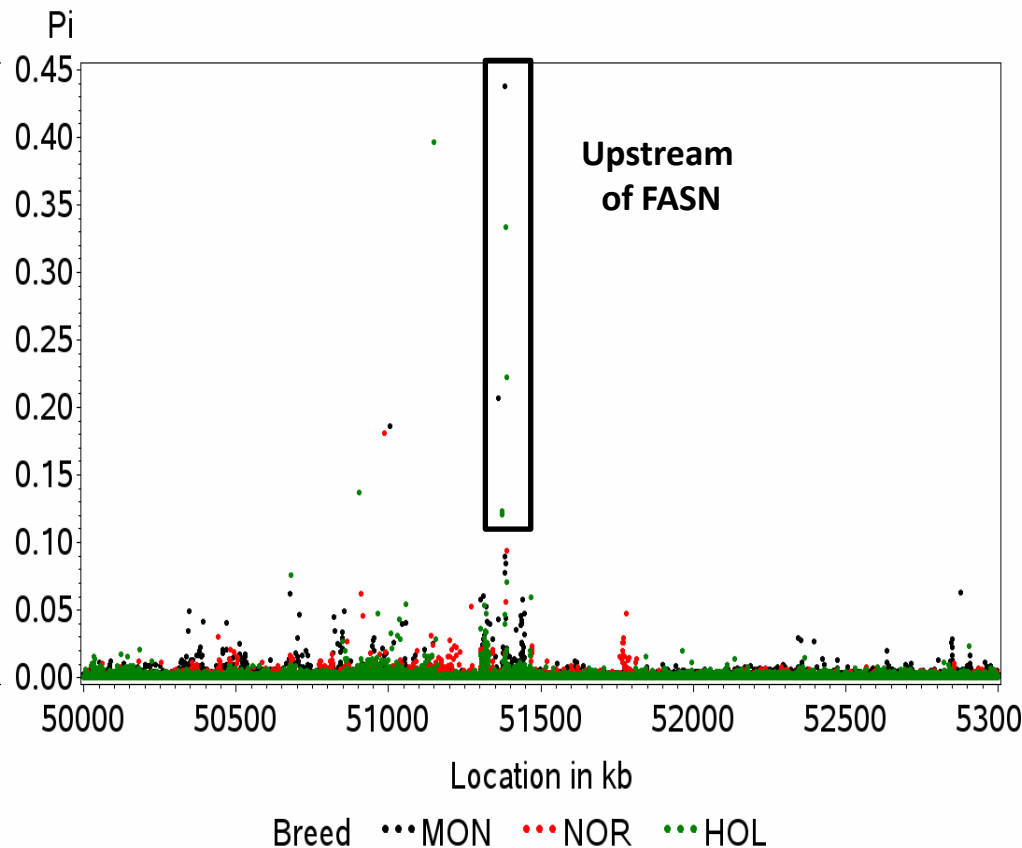
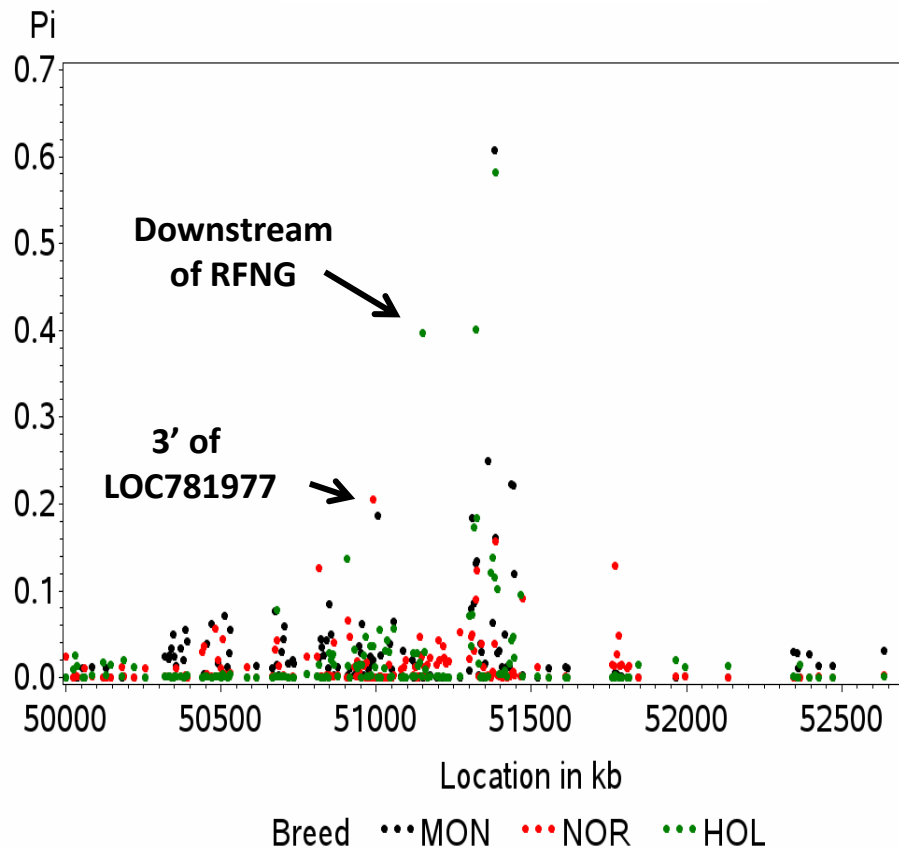


25 markers in very high LD, with similar probabilities, in the BRI3BP gene (all intronic)

# Results: Chromosome 19, C12:0

Sum over 5kb windows

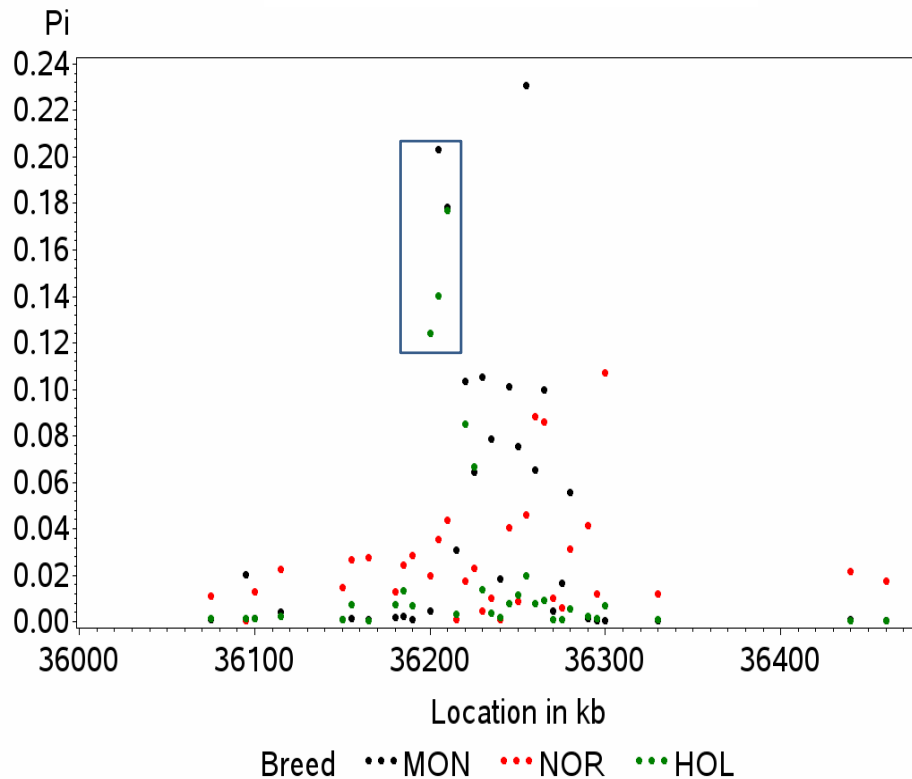
Inclusion probability for each SNP



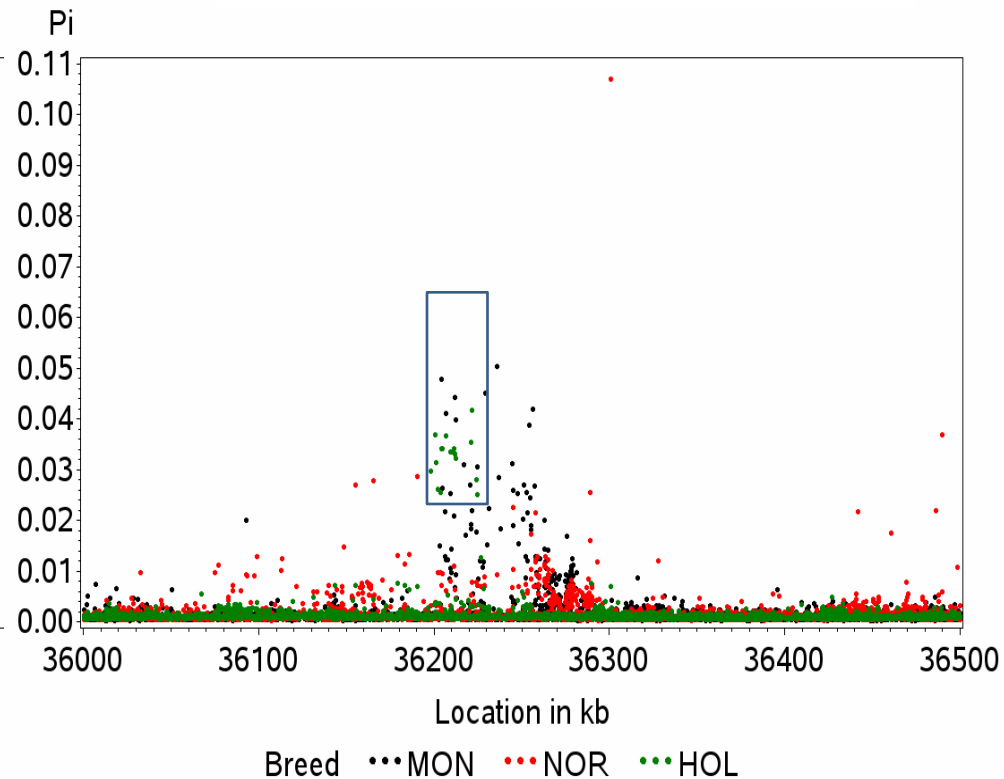
Several genes involved  
6 candidates in the upstream regulatory region of FASN

# Results: Chromosome 27, C16:0

Sum over 5kb windows



Inclusion probability for each SNP



12 variants very close to each other present PI between 2 and 5% in two breeds  
4 with the highest probabilities are upstream of AGPAT6 and are the best candidates



# Results: Summary of results

BTA	Bounds of peak (kb)	Trait	Candidate variants	Genes	Annotation of variants in genes
5	93 940-93 955	SAT	4	MGST1	Upstream
	1620-1625	SAT, POLY	1	GPT	3'UTR
14	1790-1870	SAT	4	DGAT1	Various
	2700-2720	POLY	4	CYP11B1	Upstream / Downstream
17	53 075-53 085	C4:0	22	BRI3BP	Intronic
19	51 360-51 385	C12:0	6	FASN	Upstream
27	36 205-36 220	C16:0	4	AGPAT6	Upstream

## Conclusion

BayesC, used to analyze targeted regions, is a good tool to select candidate mutations, in combination with functional annotation

Across breeds, when QTL co-localize, we observed that the same genes are involved

But across breed information is weaker than expected to target candidates. A majority of candidate mutations seem to be breed specific

Invitation: Talk 214 Session 20, Tuesday, 3:30 : Identification of causal variants for milk protein composition using sequence data in dairy cattle, by MP **Sanchez** et al

# Acknowledgements



« The 1000 bull genomes » consortium

