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Data integration and network construction with muscle metabolome and meat quality data in pigs

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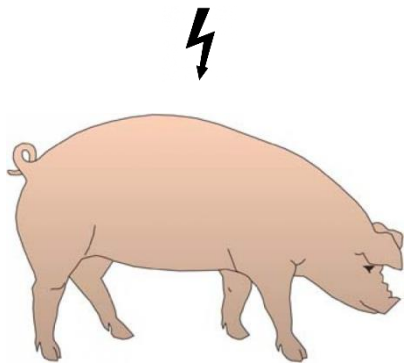
Introduction

- meaning of drip loss in pork

e.g. stress conditions
before slaughtering

high drip loss

reduction in meat quality¹
by impairment of...



- 1) technological meat characteristics
 - reduced acceptance of processing industries
- 2) sensory meat characteristics
 - reduced consumer acceptance

¹Fischer et al. 2007

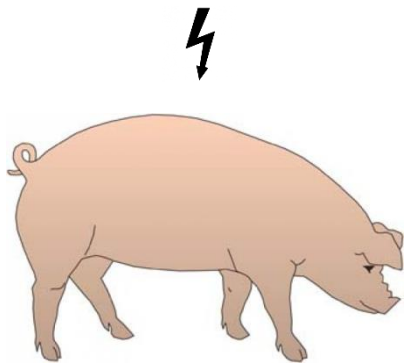
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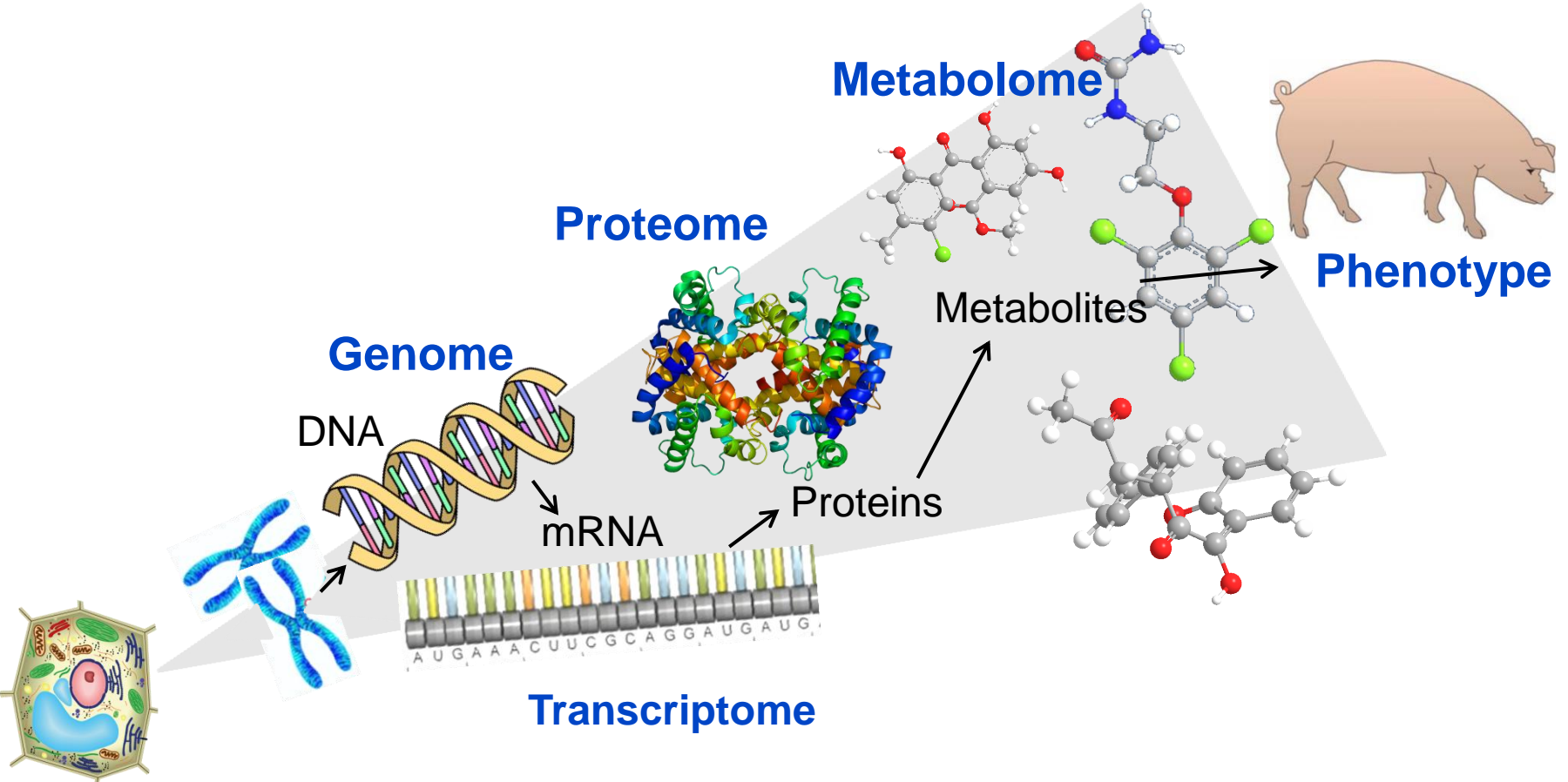
- 1) technological meat characteristics
 - reduced acceptance of processing industries
- 2) sensory meat characteristics
 - reduced consumer acceptance

➔ How to select for low drip loss despite low heritability!?

¹Fischer et al. 2007

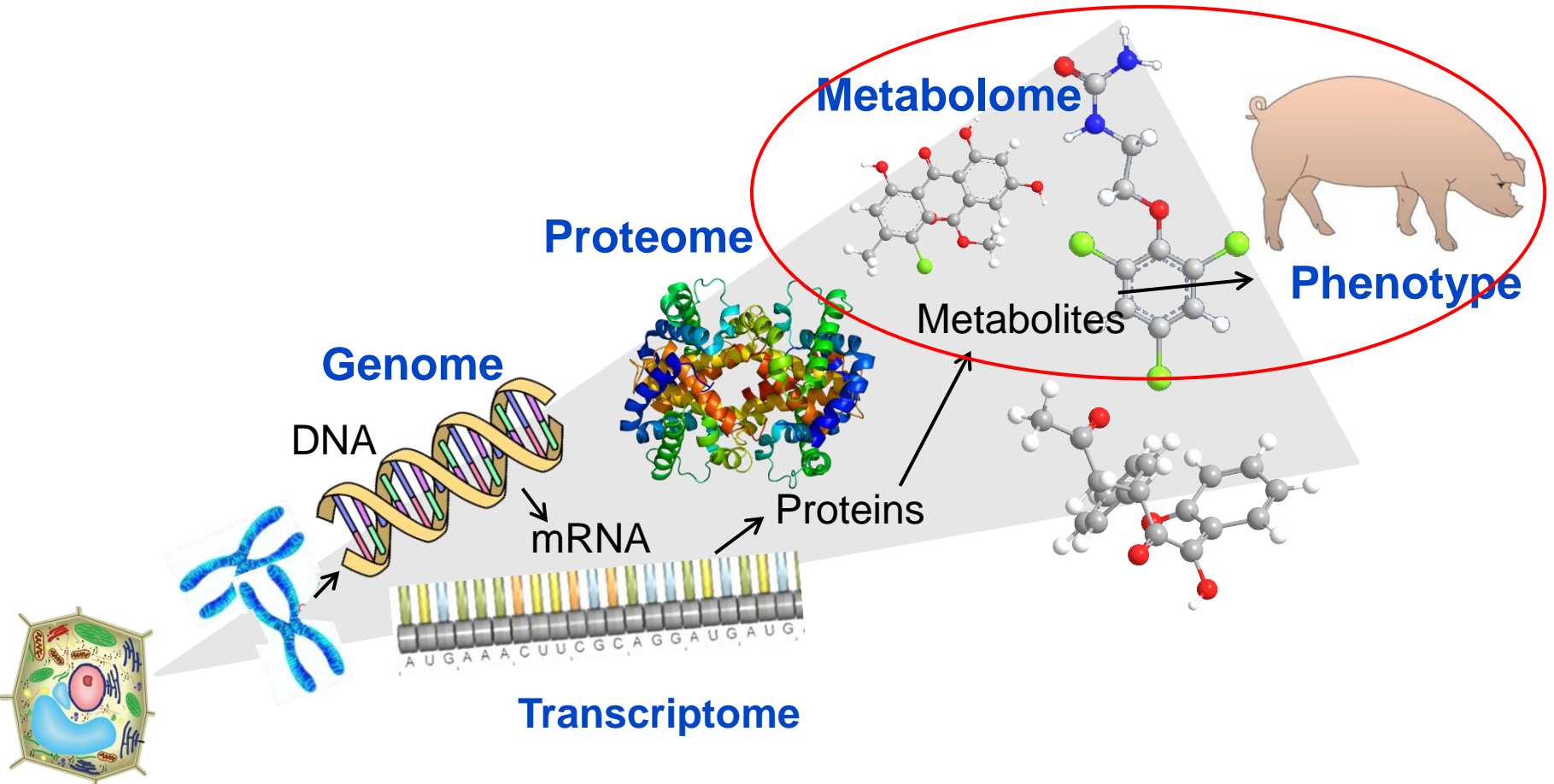
Introduction

- the way from genome to phenotype



Introduction

- the way from genome to phenotype

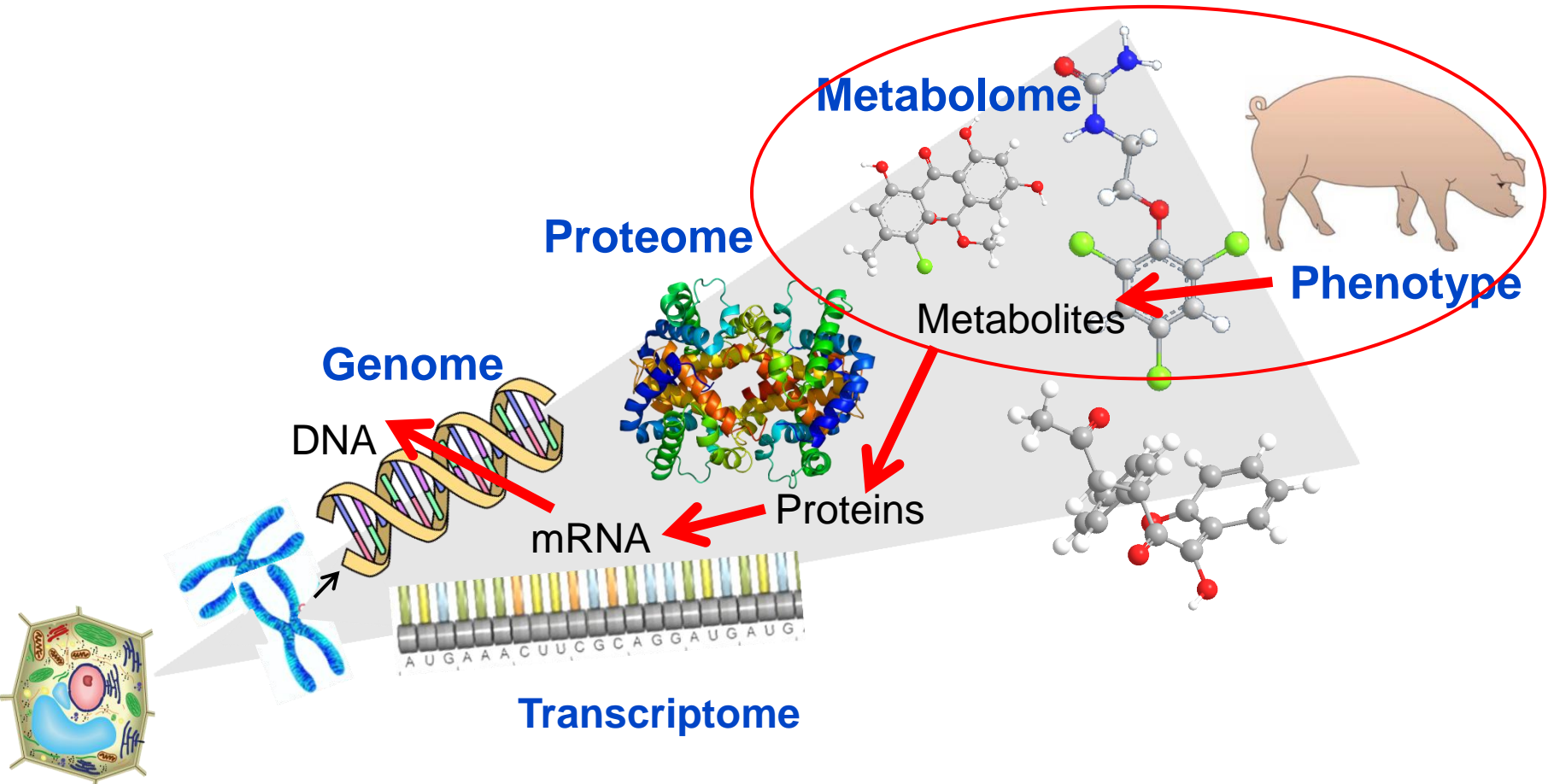


→ Metabolites as more reliable phenotype¹

¹Bino et al., 2004

Introduction

- the way from genome to phenotype



→ Metabolites as more reliable phenotype¹

→ Fill in the gap between phenotype and genome

¹Bino et al., 2004

Objective of the study

Investigation of the **relationship** between **metabolite profiles** and **drip loss**

- get new insights in the complex biochemical processes of drip loss
- test different statistical approaches to reveal the most important metabolites
- detect metabolites as potential biomarkers

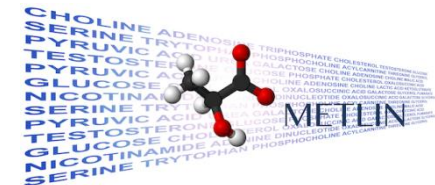
Animals and drip loss phenotyping

- 97 F₂ pigs of Duroc × Pietrain resource population
- Meat samples of *M. longissimus dorsi*
- Record of drip loss 24h pm
 - Bag-Method of *Honikel* (1986)
 - drip loss range: 0,4 – 5,3%





Quantification and annotation of metabolites

- by gas – and liquid chromatography with mass spectrometry (GC-MS, LC-MS)
- „untargeted“ metabolomics profiling
 - Detection of the whole metabolome
 - 1993 metabolites detected
- functional annotation
 - Human Metabolome Database, Lipid Maps, METLIN
 - 400 of 1993 metabolites annotated





- Pre-correction of phenotypes and metabolite profiles
 - generalized linear model (GLM) with slaughter weight and season

- Statistical approaches to analyse the drip – metabolite associations
 1. Correlation analysis
 2. Principal component analysis (PCA)
 3. Weighted network analysis (WNA)
 - using  WGCNA¹
 4. Random forest regression² (RFR)
 - using  party³

¹Langfelder & Horvath 2008, ²Breimann 2001, ³Strobl et al. 2007

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**handle the
„Large p, small n“-
problem (Overfitting)**

¹Langfelder & Horvath 2008, ²Breimann 2001, ³Strobl et al. 2007

2. PCA

- Condenses the metabolite profiles into representative, uncorrelated principle components (PCs)
- Metabolites are quantified by their corresponding loadings

3. Weighted network analysis (WNA)

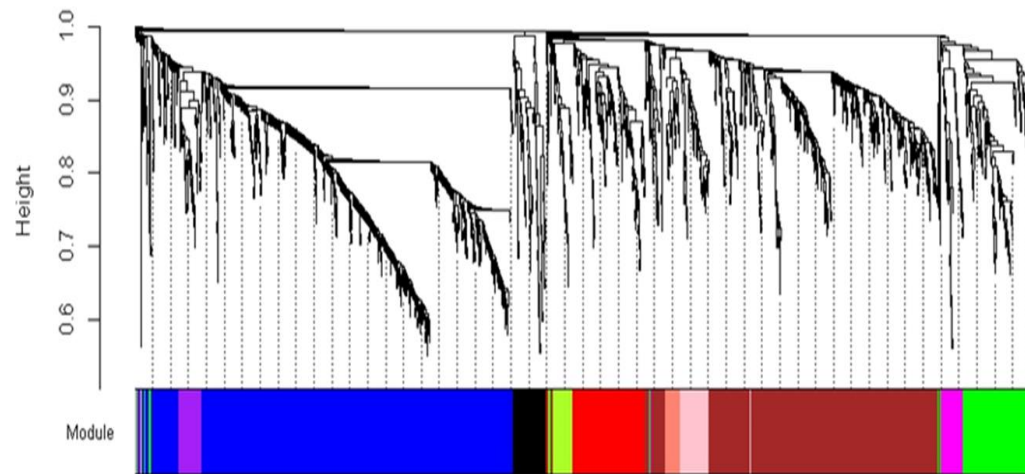
- Generates biological interpretable modules based on a hierarchical clustering dendrogram
- Metabolites are characterised by

➤ module membership (MM)

- Connectivity of the metabolites within a module

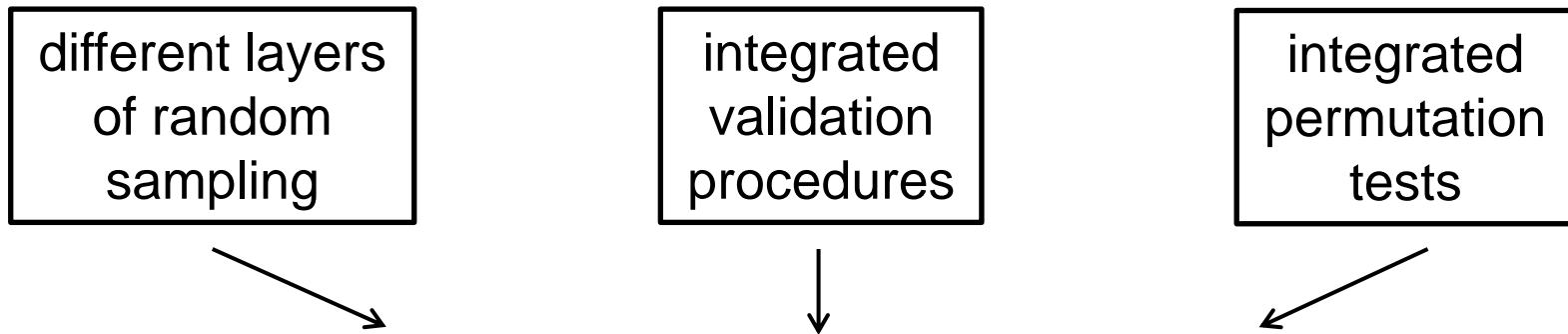
➤ metabolite significance (MS)

- Based on the trait ↔ metabolite correlation



4. Random Forest Regression (RFR)

- supervised learning tool using tree-based methods
- key characteristics of RFR:



➤ RFR is able to handle datasets with complex interaction structures and highly correlated variables

- RFR calculates *variable importance (VI)* values, that are based on prediction accuracy¹

¹Strobl et al. 2007

Results

– Identification of biomarkers

1. Correlation analysis

- 71 (5) metabolites positive (negative) correlated ($p \leq 0,05$)
- Range: 0.24 to 0.28 respectively -0.24 to -0.23

2. PCA

- First 3 PCs specify 46.9 % of metabolite expression variance
- Loadings are very weak (range: -0.1 to 0.1) → not significant¹
- PCA not used for biomarker identification
- But: PCA was used to reduce the data set for RFR

¹DiLeo et al., 2011

Results

– Identification of biomarkers

3. Weighted Network analysis (WNA)

- Clusters the metabolites into 10 modules
 - two modules significantly associated with drip loss

trait	module	cor.	p-value	number metabolites
drip loss	,purple'	+ 0.21	$p \leq 0.04$	52
	,green'	+ 0.21	$p \leq 0.04$	49

4. Random Forest Regression (RFR)

- 293 metabolites with significant ($p \leq 0,05$) *variable importance* (VI) for drip loss

Results – network construction

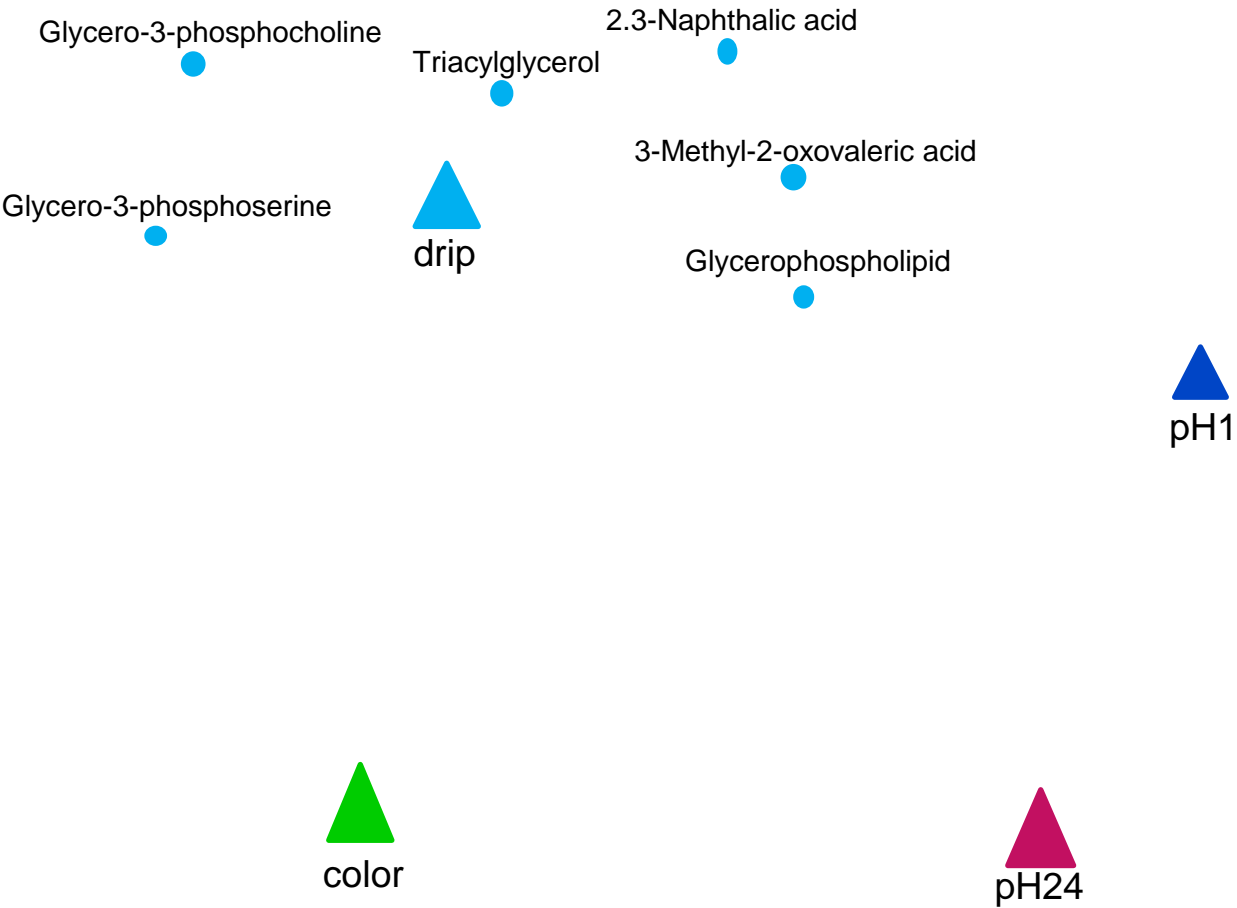

drip


pH1

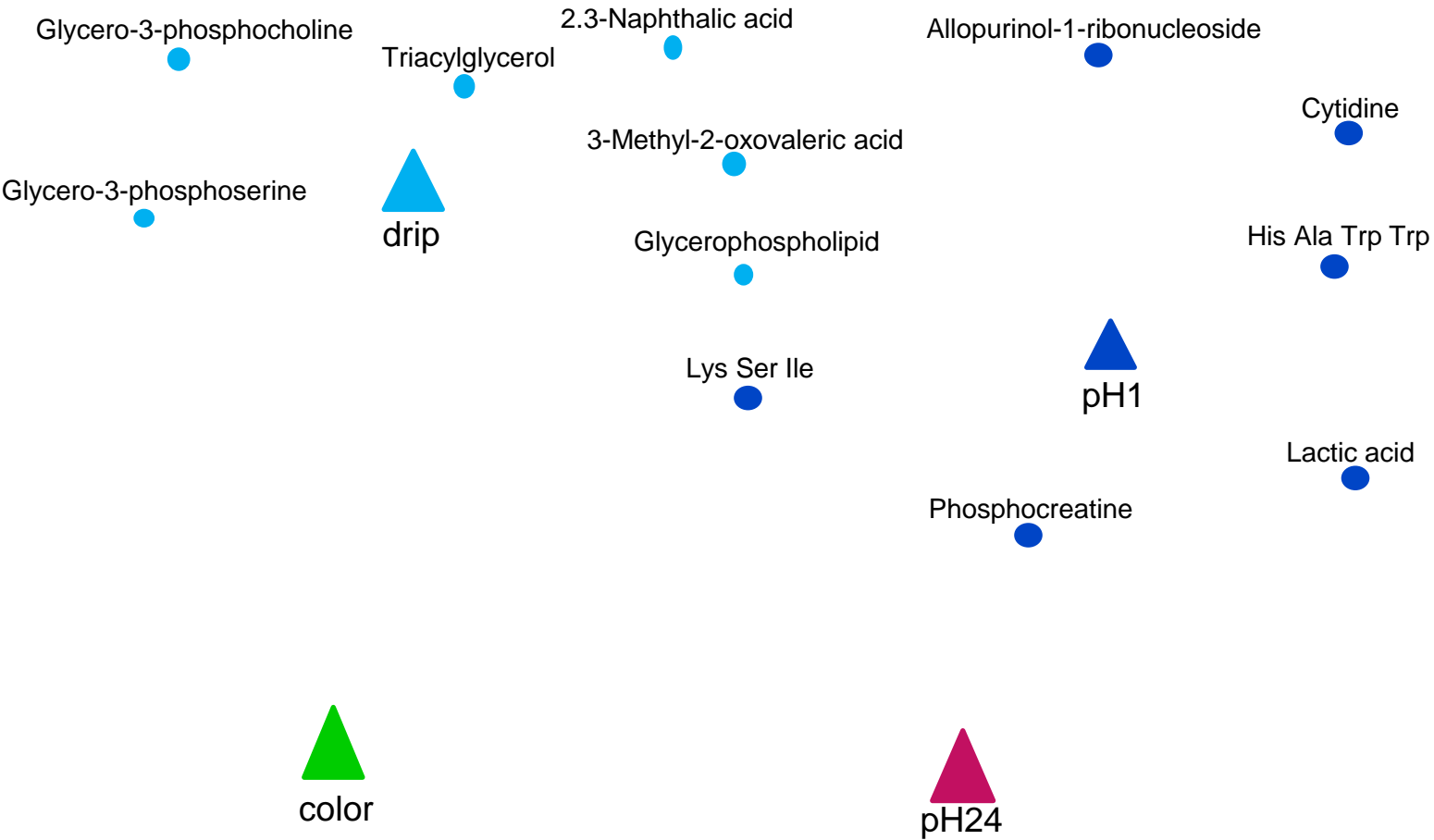

color


pH24

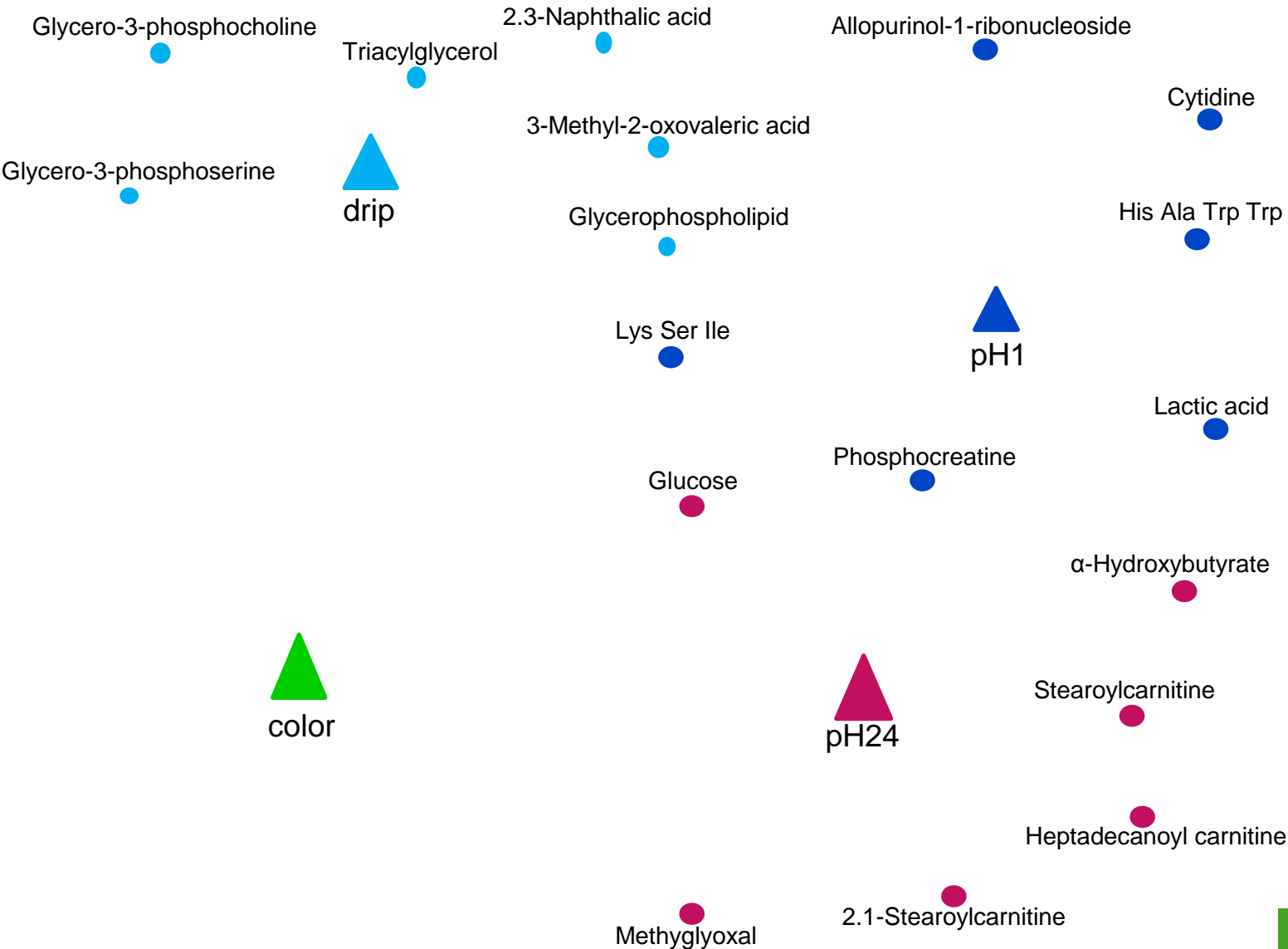
Results – network construction



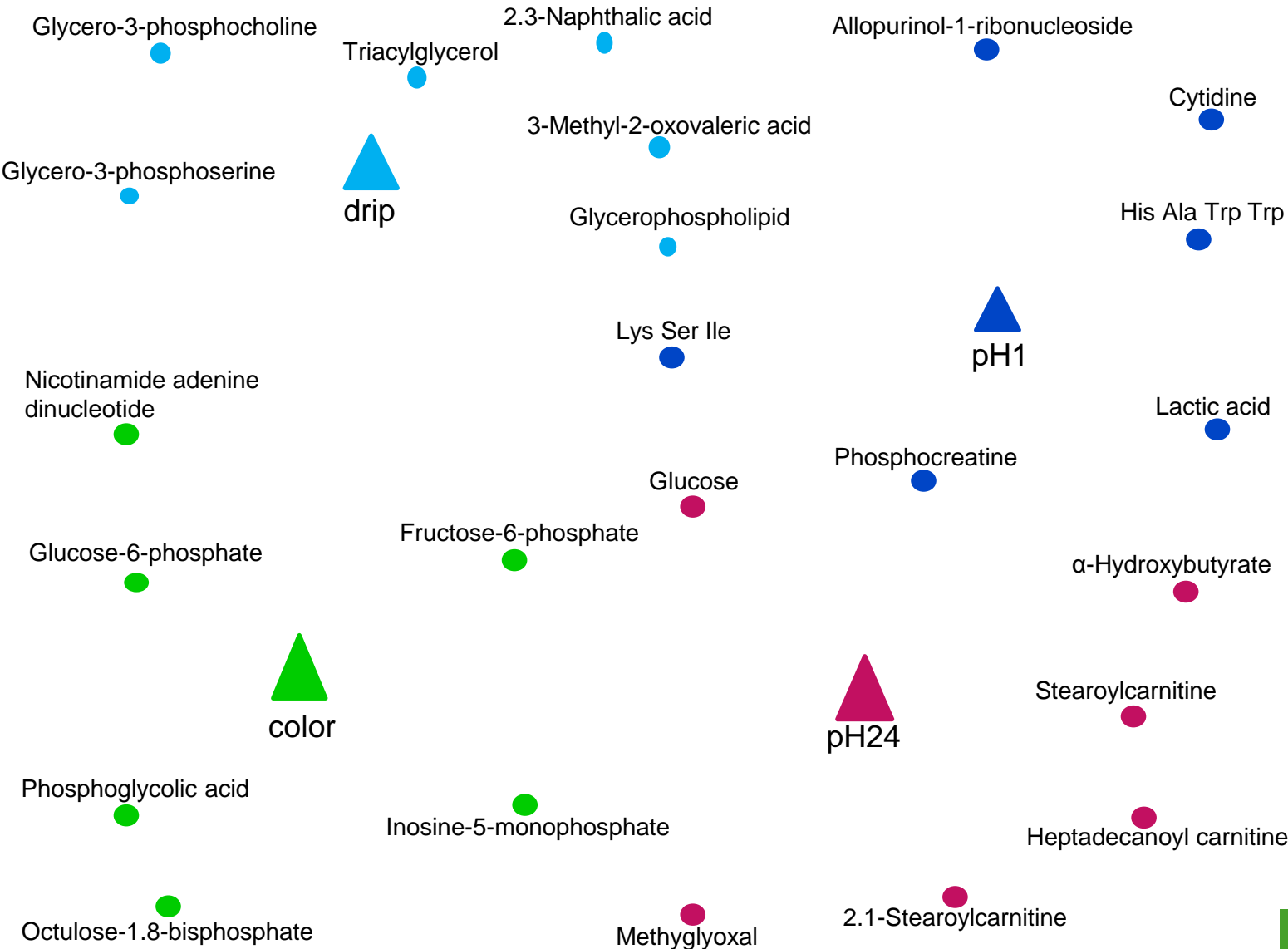
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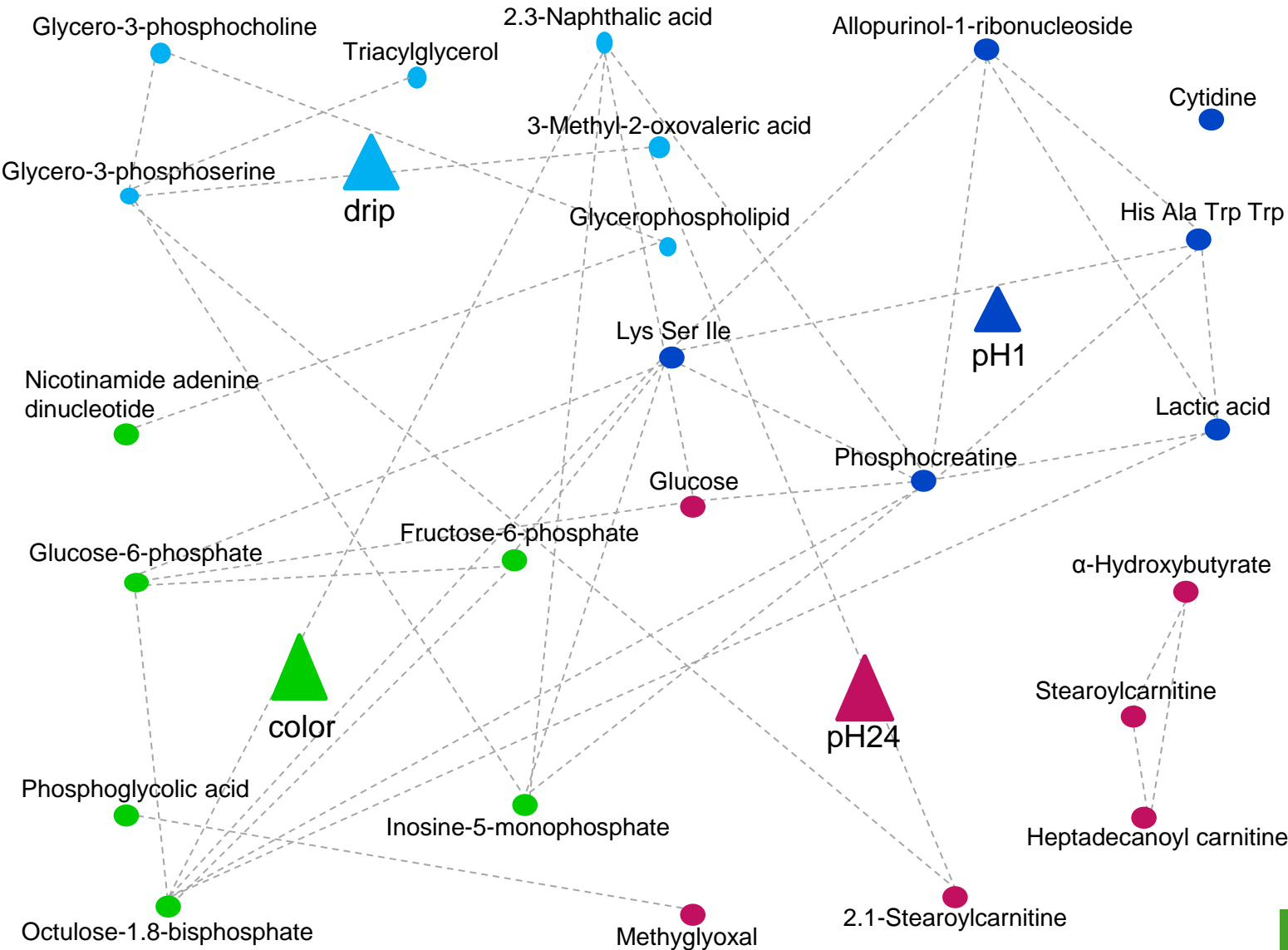
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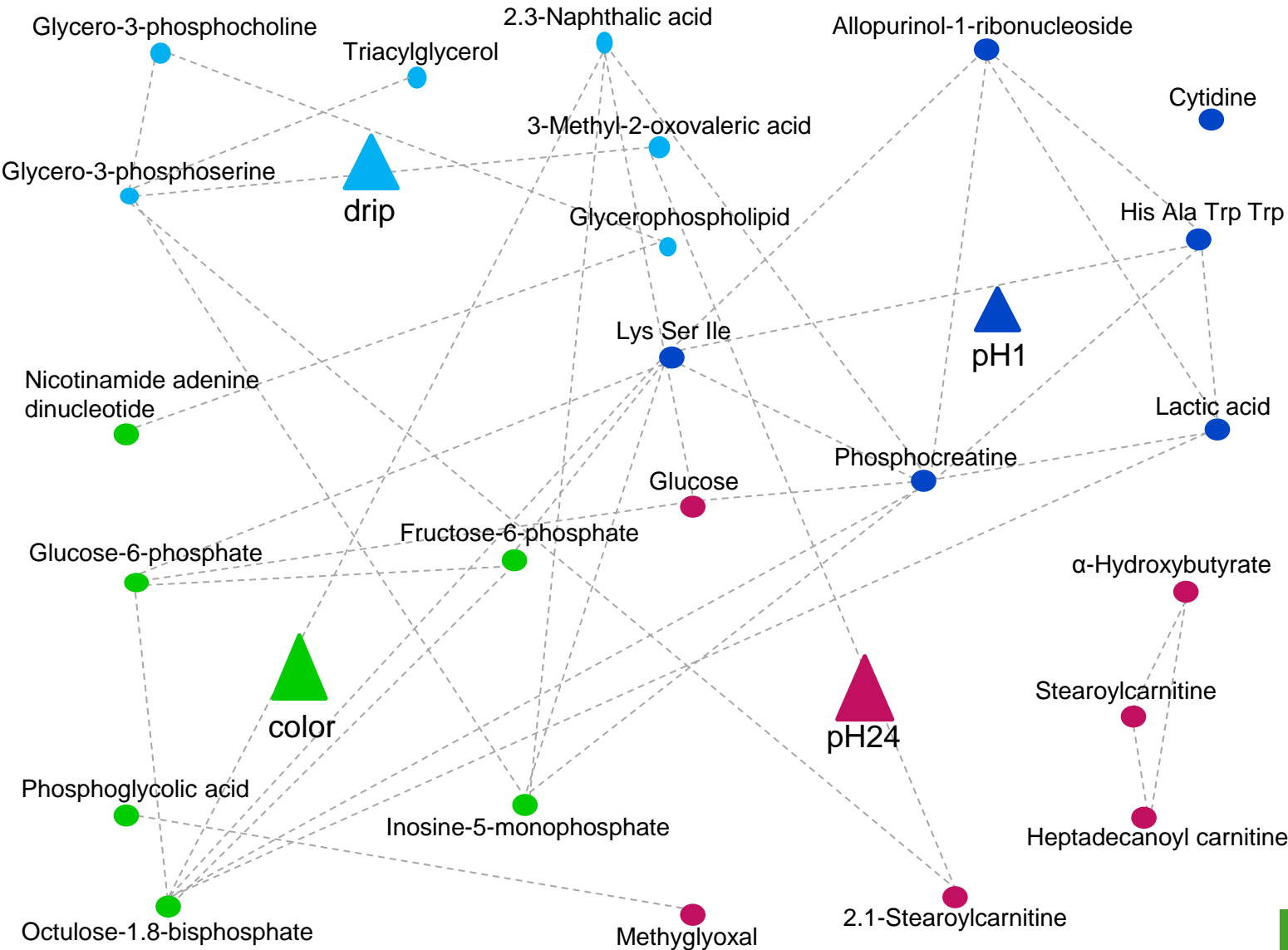


Results – network construction



Dotted lines:
connectivity
between
metabolites

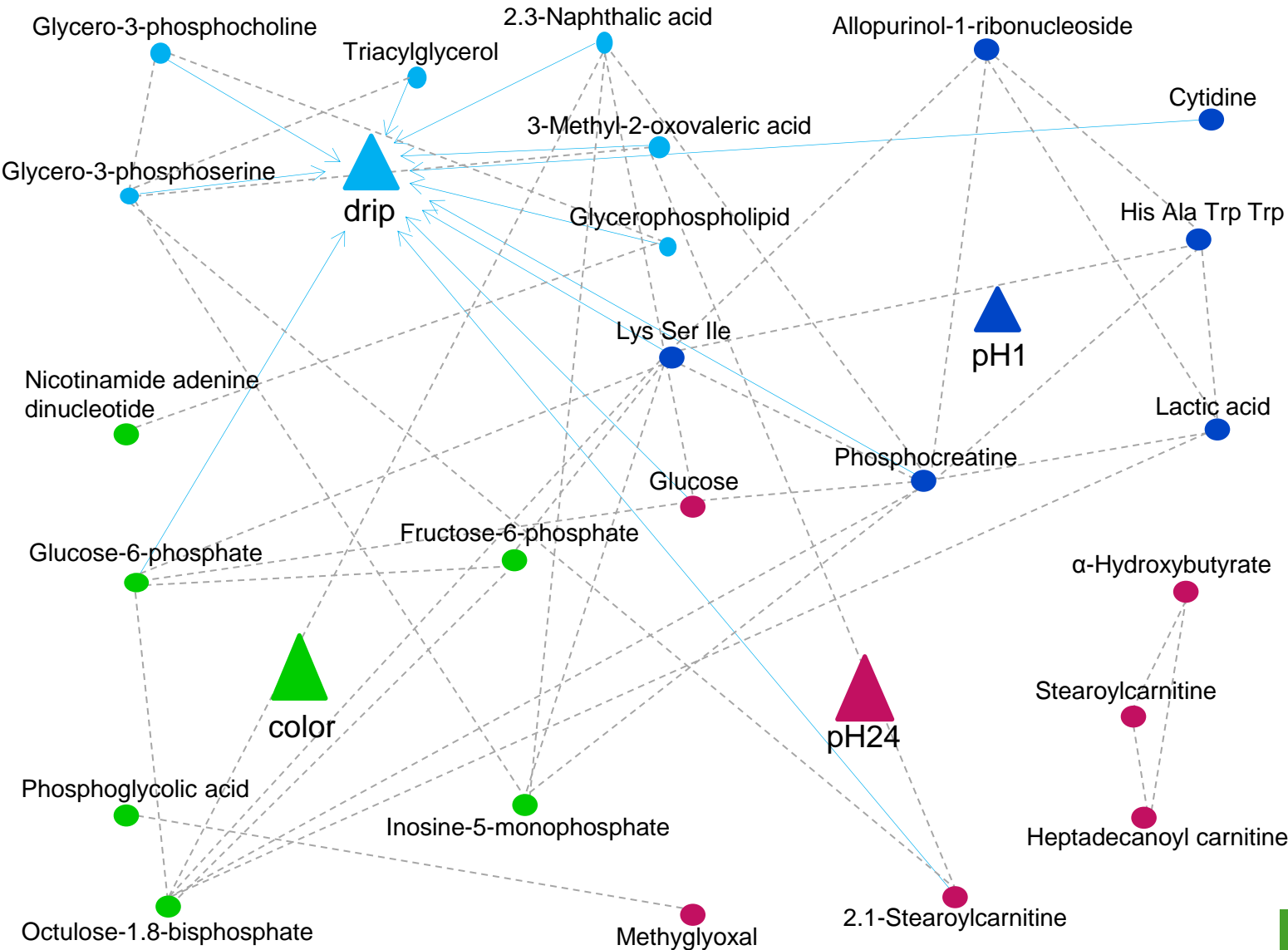
Results – network construction



Dotted lines:
connectivity
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Arrows:
directed connections
metabolite
→ trait

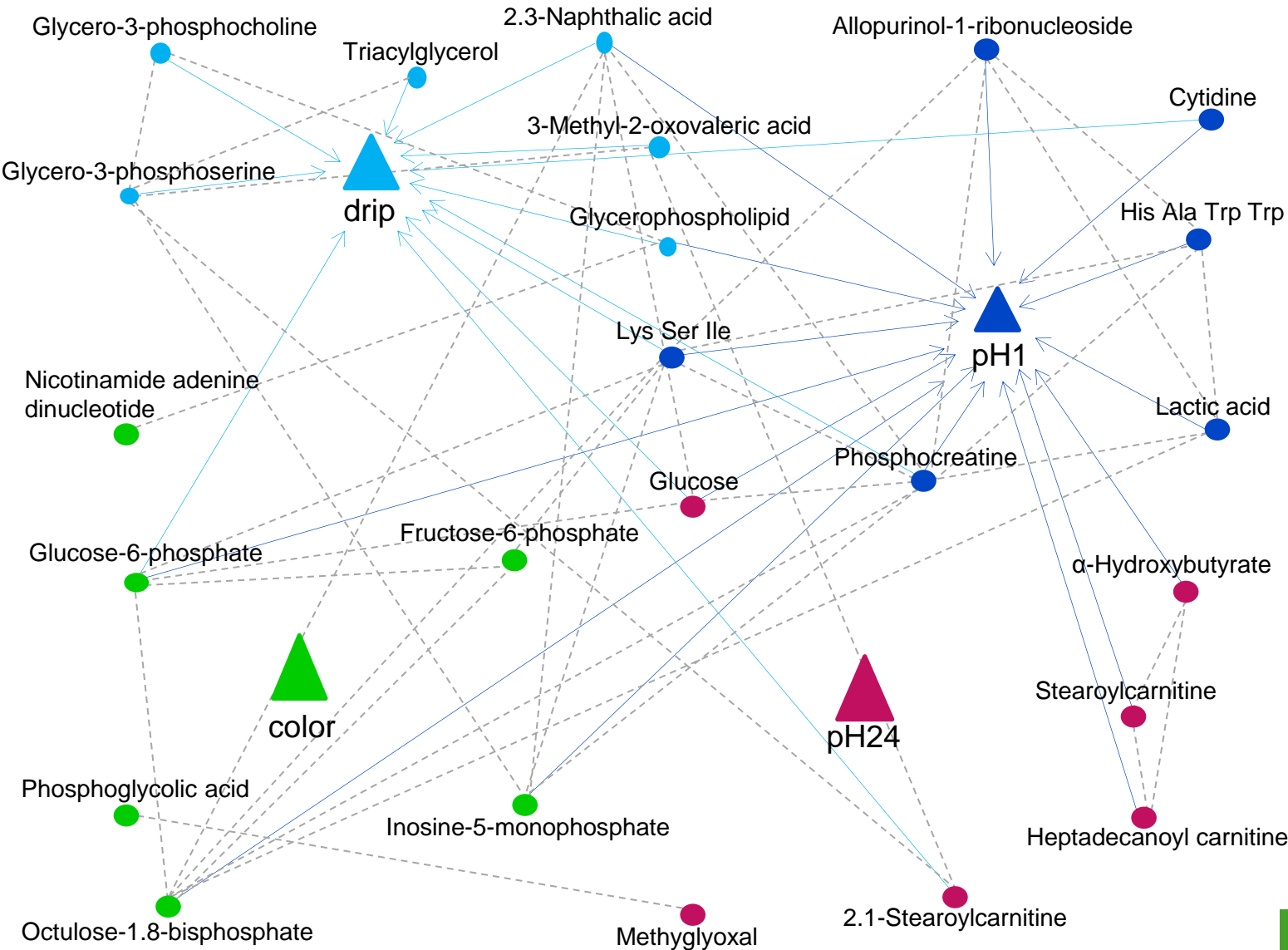
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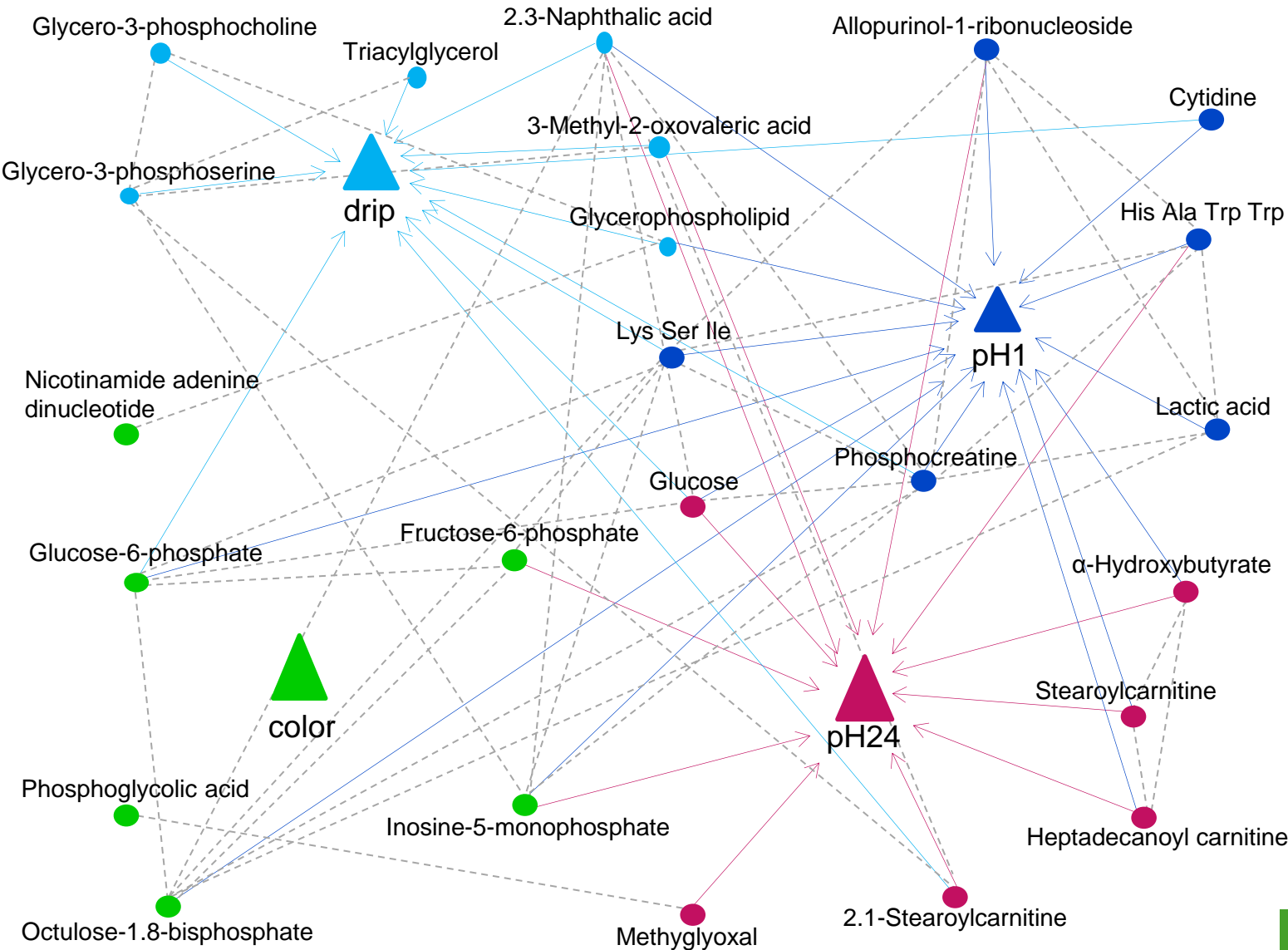
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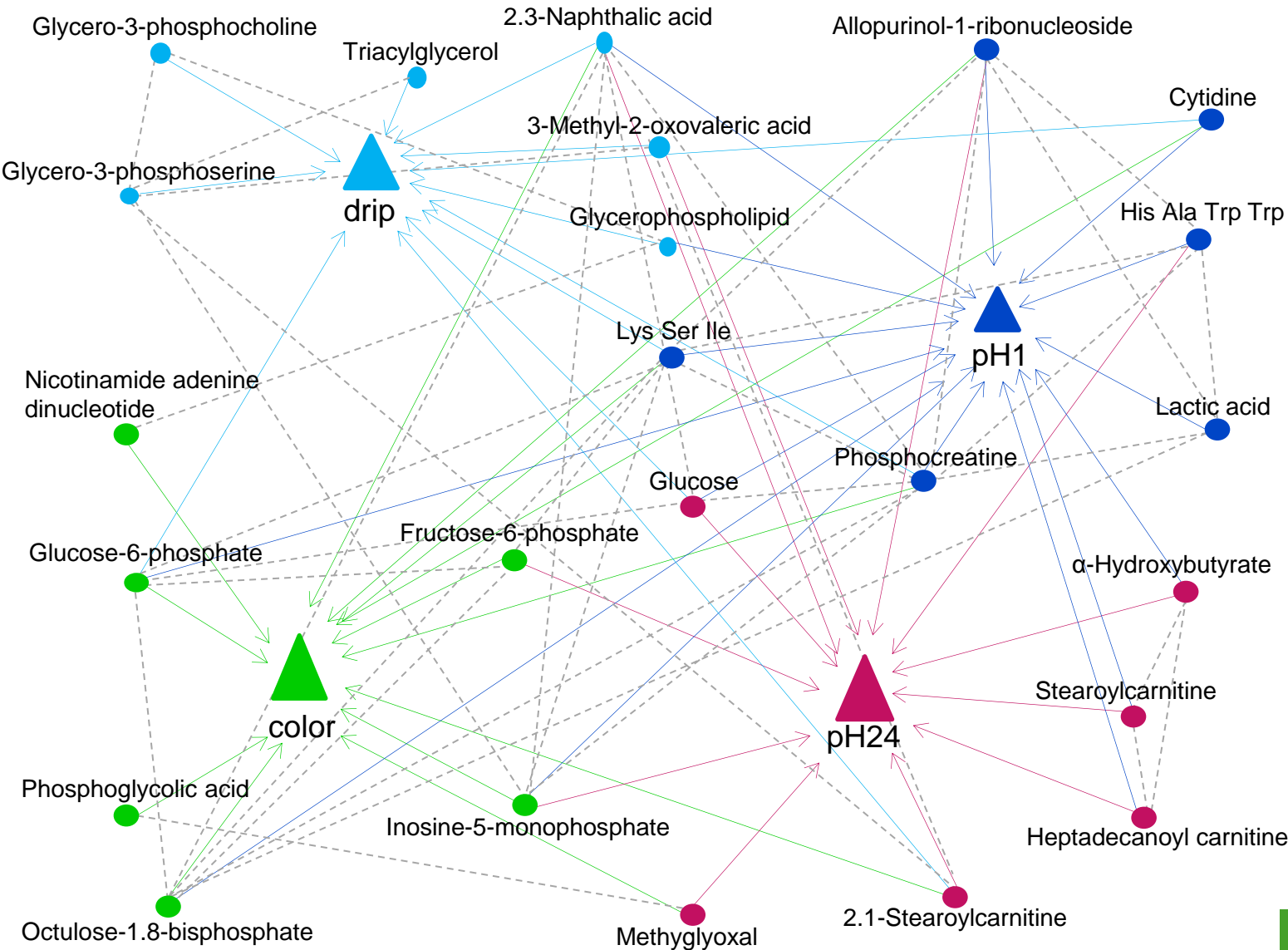
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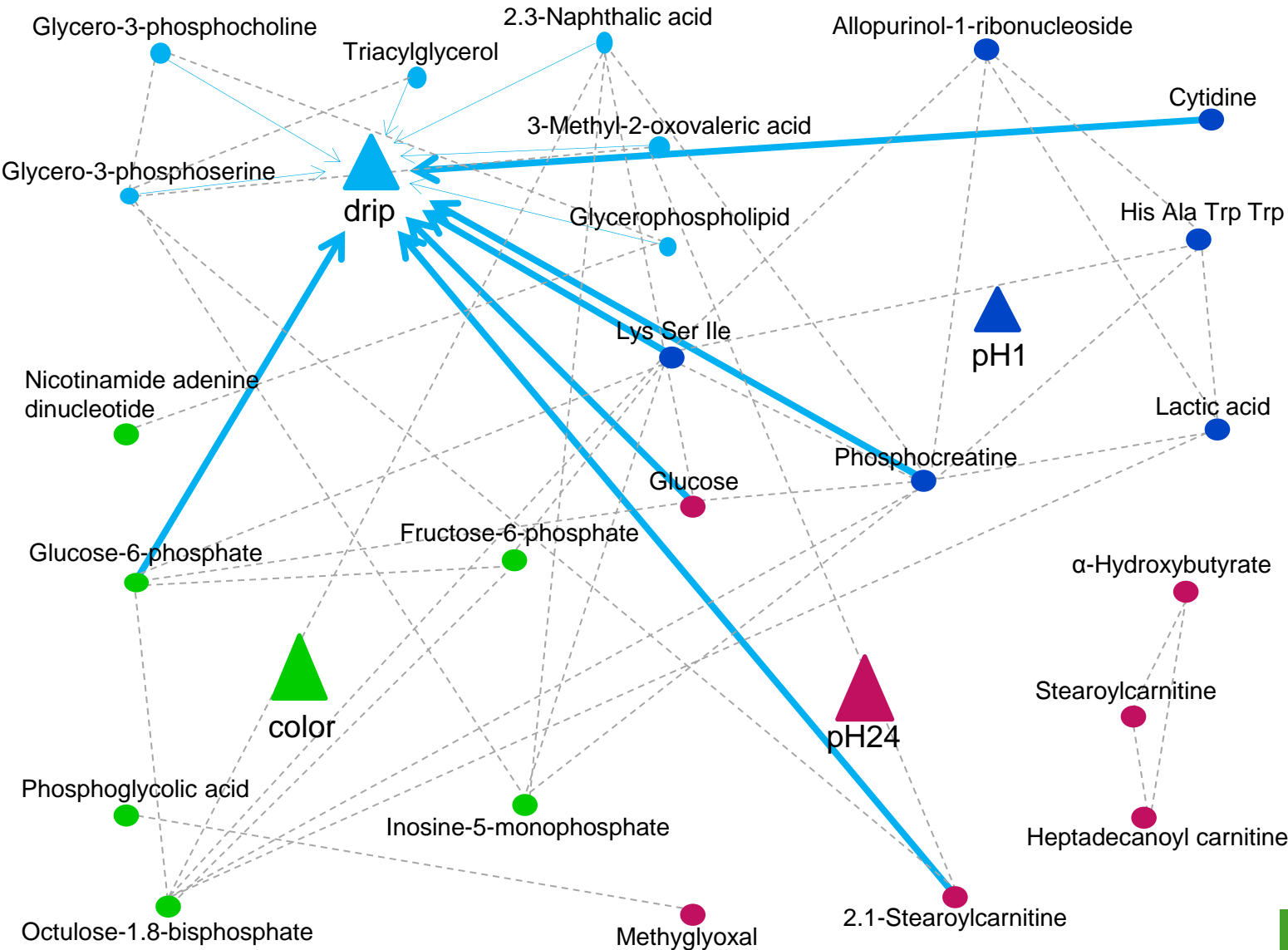
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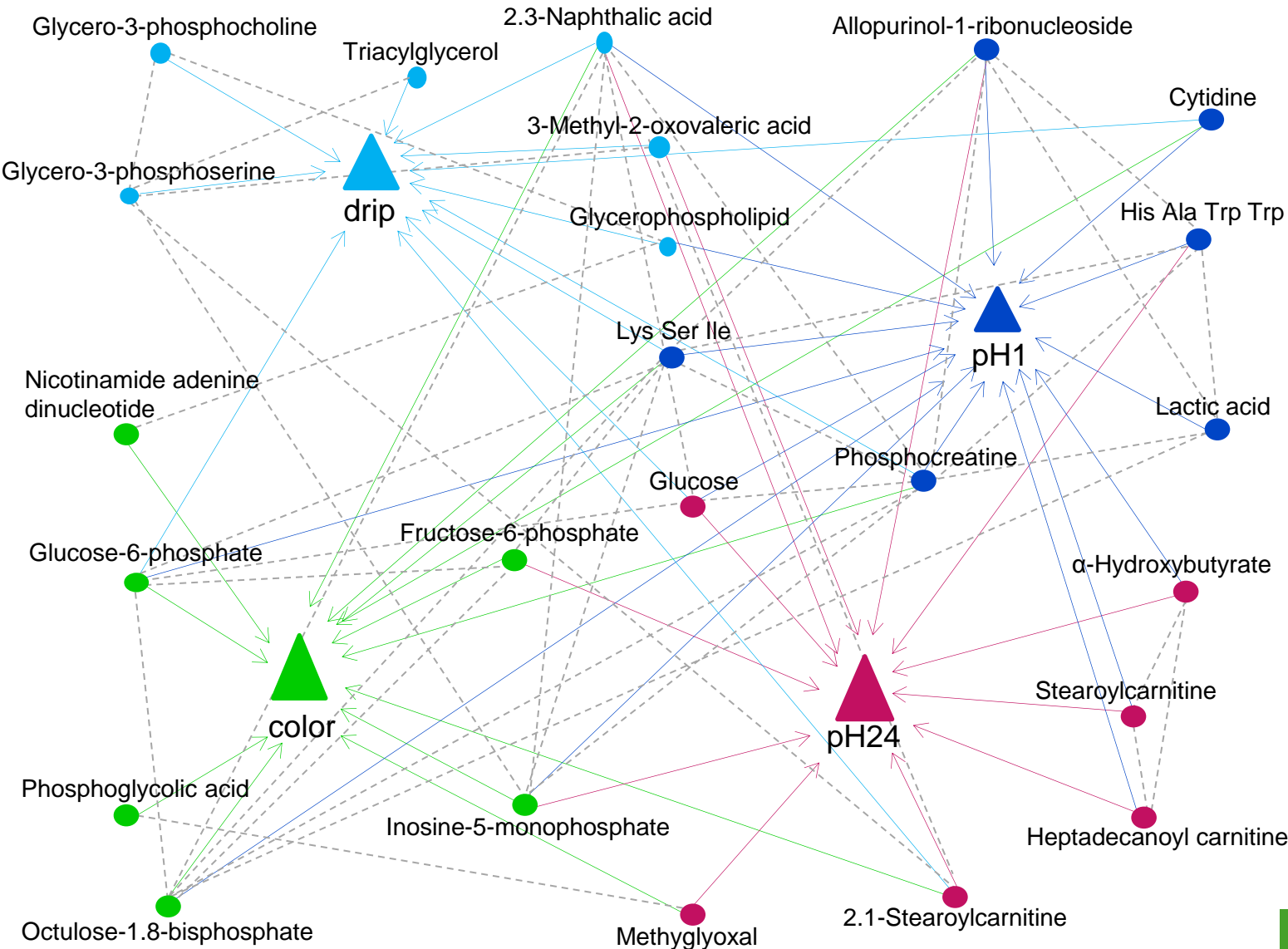
Results – network construction



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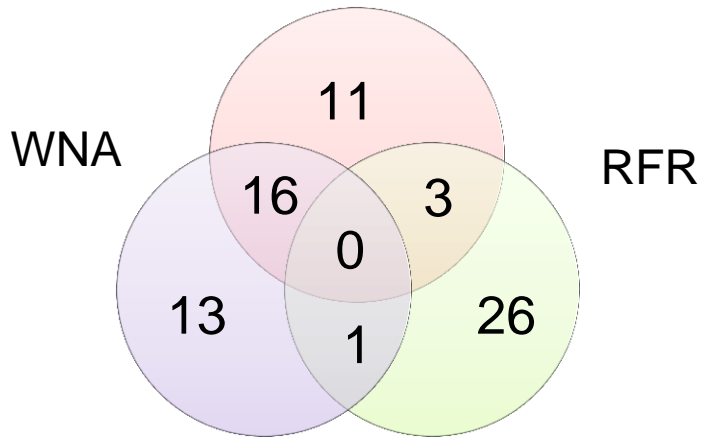
Dotted lines:
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metabolite
→ trait

Results - prediction accuracy of selected metabolites

- overlap in “Top 30” significant metabolites for drip loss

correlation analysis



- 20 metabolites identified by more than one method
- multiple R²: 32.73 %

- stepwise Regression of the 20 metabolites
- resulted in 5 important metabolites

biomarker for drip loss	regression coefficient
Glycerophosphocholine	45.82*
C24:1 Sphingomyelin	-50.53*
Ubiquinon (Prenol Lipid)	532.20*
C26:1 Sphingomyelin	1870.00**
not annotated	398.50***

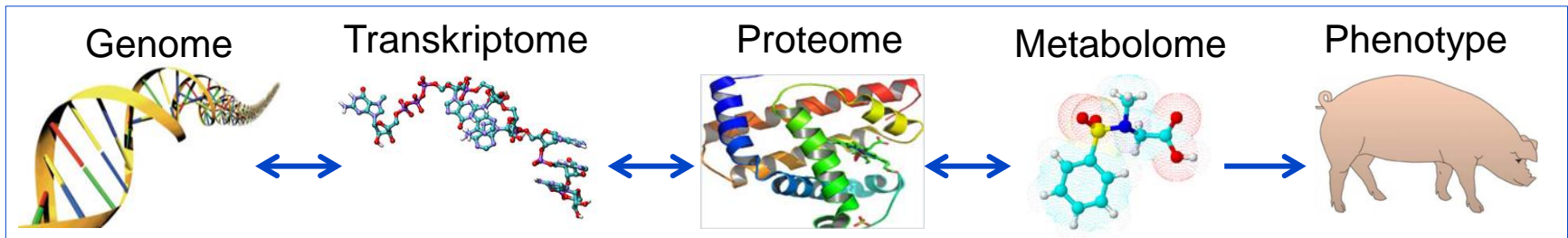
- multiple R²: 26.61 %

Conclusion and perspective

- Correlation analysis, WNA and RFR are suitable in identification of predictors
- Glycerosphospho- und Sphingolipids are the most promising biomarkers
- Selected set of metabolites has moderate prediction accuracy and also an effect on other meat quality traits



- Requirements of the development of reliable metabolite biomarkers
 - ↳ enhanced abilities of metabolite quantification and annotation
- Future perspective: usage of combined omics-profiles as more exact phenotype and in GWAS of identify ,real‘ powerful SNPs



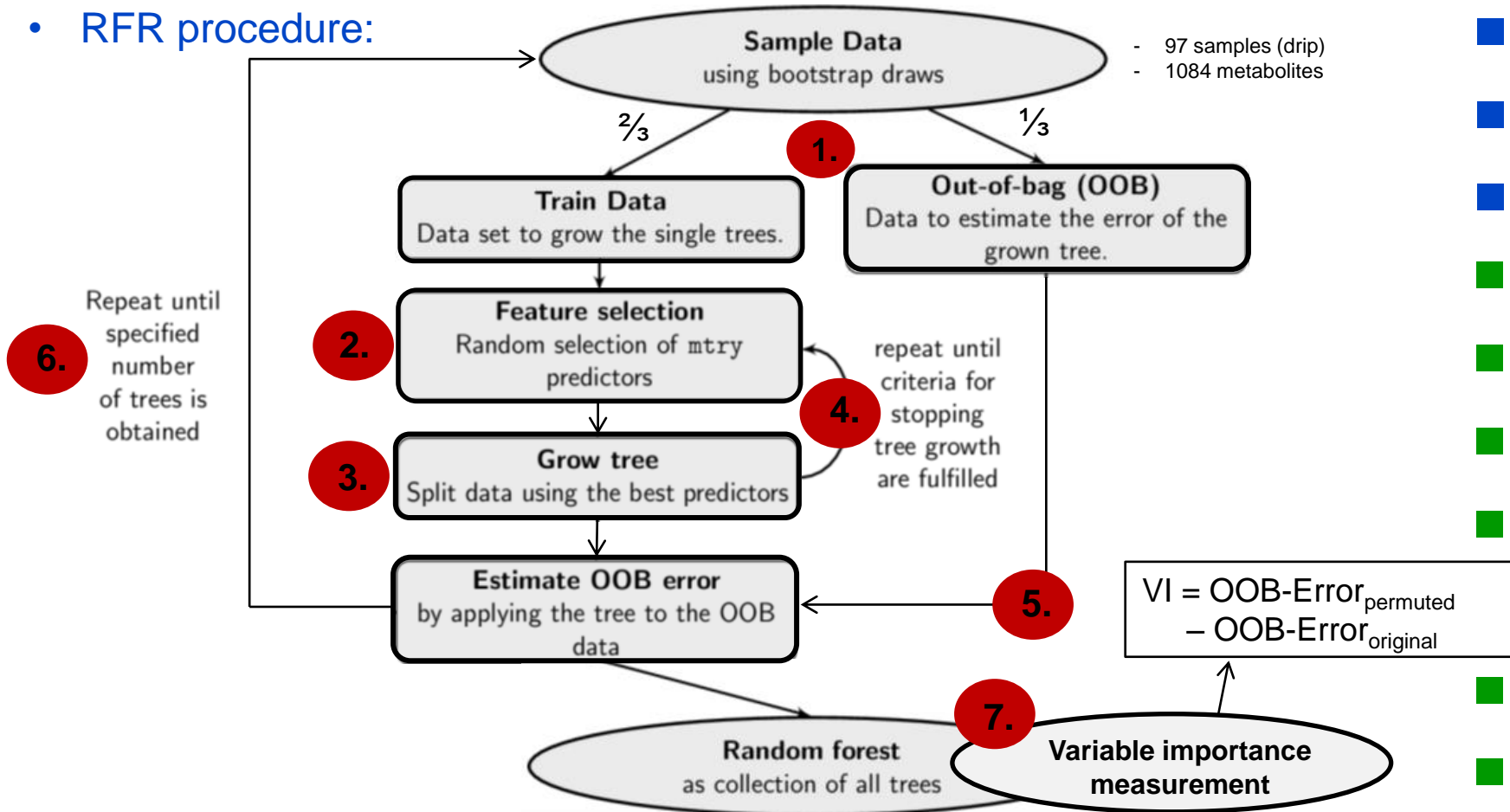


**Thank you for
your attention**

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Random Forest Regression (RFR)

- supervised learning tool using tree-based methods with integrated permutation tests
- RFR procedure:



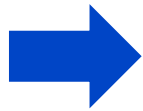
Results - important biomarkers

- Ranking of metabolites in „top 30“ of correlation analysis, WNA and RFR

drip loss	Cor	MS	VI	pH1	Cor	MS	VI
2.3-Naphthalic acid	23.	x	10.	His Ala Trp Trp	5.	4.	2.
Glycero-3-phosphocholine	8.	x	7.	Cytidine	25.	8.	12.
Glycero-3-phosphoserine	x	28.	23.	Allopurinol-1-ribonucleoside	x	9.	25.
Glycerophospholipid	22.	14.	x	Lactic acid	24.	x	10.
Triacylglycerol	19.	12.	x	Lys Ser Ile	19.	x	6.
3-Methyl-2-oxovaleric acid	21.	13.	x	Phosphocreatine	26.	x	21.

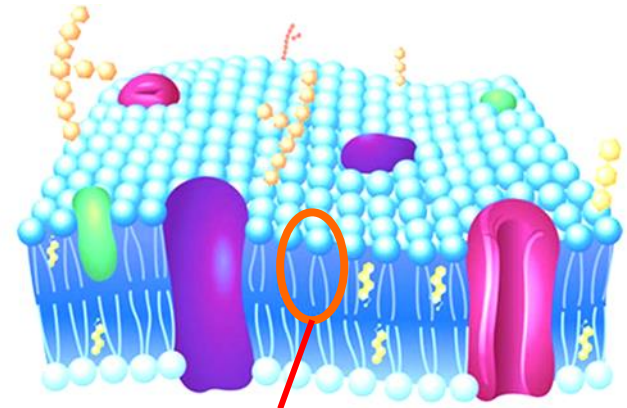
pH24	Cor	MS	VI	color	Cor	MS	VI
α -Hydroxybutyrate	1.	1.	x	Octulose-1.8-bisphosphate	7.	1.	x
Heptadecanoyl carnitine	2.	2.	x	Fructose-6-phosphate	27.	9.	x
Stearoylcarnitine	3.	4.	x	Glucose-6-phosphate	23.	7.	x
Gle-cholesterol	x	x	2.	Inosine-5-monophosphate	28.	10.	x
Methyglyoxal	x	x	9.	Phosphoglycolic acid	11.	12.	x
Glucose	x	x	11.	Nicotinamide adenine dinucleotide	4.	x	2.

Results – potential biomarkers

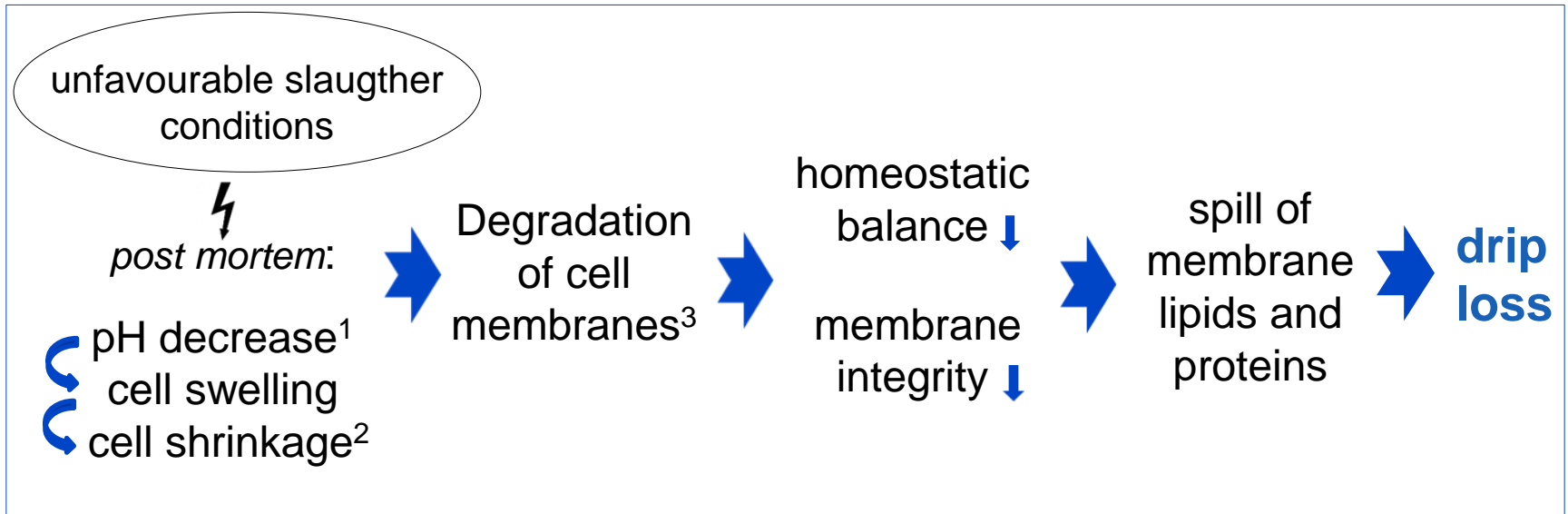


consistent results of statistical approaches:

Glycerosphospholipids (GPLs),
Sphingolipids (SLs)



membrane lipid



¹Ortenblad et al., 2003; ²Bertram et al., 2004; ³Lambert et al., 2001