

# Effect of iron source on the alleviation of <sup>KJ21</sup> nutritional anaemia in common sole (*Solea solea*)

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## Diapositiva 1

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**KJ21**

**abstract number 22866**

Kals, Jeroen; 18/08/2016

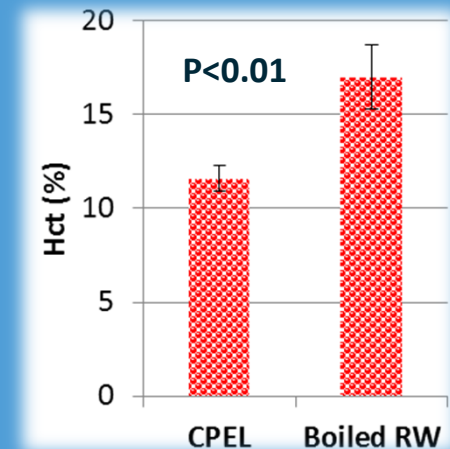
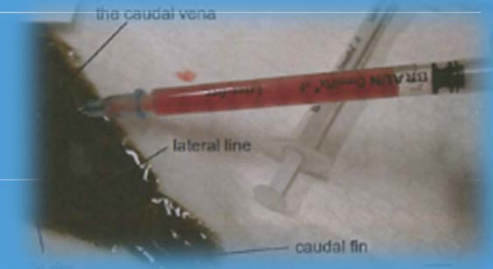
# Effect of iron source on the alleviation of nutritional anaemia in common sole

- Introduction
- Hypotheses
- Material & methods
- Results
- Discussion
- Conclusions



# Introduction

- Sole fed commercial pellets (CPEL) suffer from a **nutritional** anaemia
- An iron (**Fe**) deficiency is a common cause
- Intake of Fe can be eliminated
- Intake **Fe** in sole fed boiled ragworm  $\approx$  sole fed CPEL, but **Hct** sign. higher
- Yet, **bioavailability** of iron cannot be excluded



# Introduction



- Bioavailability iron depends on **form** (e.g. heme vs. non heme)
- Absorption **heme** vs. **non heme** differs as heme
  - has its own pathway
  - independent of pH
  - immune for antagonists (e.g.  $\text{Ca}^{2+}$ ) KJ22
- Fe in CPEL is mainly non heme from premix or damaged heme
- **Alkaline** character of sole's intestine might **hamper** absorption of non heme iron, leading to an **iron** deficiency?



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**KJ22** non heme requires reduction of  $\text{Fe}^{3+}$  to  $\text{Fe}^{2+}$ ,  
and hence, an acidic environment  
Kals, Jeroen; 19/08/2016

# Hypotheses



1. **nutritional** anaemia in sole fed CPEL is due to an iron deficiency
  2. assumed Fe deficiency is due to **poor** absorption of Fe
  3. increase in absorption due to a higher bioavailability of heme or iron chelates will **alleviate** anaemia in sole
  4. haematocrit (**Hct**) and haemoglobin (**Hb**) are expected to follow iron absorption patterns
- We also estimated absorption of Cu, Co, Cr, Mn, Mo & Zn to evaluate **interaction** between iron source and other minerals

# Material and Methods



- Sole raised on CPEL and anaemic at start
- Dietary treatments:
  - Fe sulphate
  - Fe methionate
  - Fe proteinate
  - Haemoglobin
- Feeding: restricted, equal feeding levels for all diets
- Tank experimental unit (n=3, 10 fish.tank<sup>-1</sup>)
- Duration 23 days, sampling at start & day 23





# Material and Methods

## Formulations

Dietary Fe

sources 64%  
basal diet 36%

Marker: Yttrium

Processing:

Cold extrusion was used to keep iron sources in their native state

Diet Code	A	B	C	D
Iron source	<i>Iron sulphate</i>	<i>Heme</i>	<i>Iron proteinate</i>	<i>Iron methionate</i>
<b>Basal ingredients (%)</b>	57.2	57.2	57.2	57.2
<b>Test ingredients (%):</b>				
Caseine	16.38	9.00	16.35	16.35
Pea protein conc.	13.00	12.70	13.00	13.00
Corn gluten	12.50	13.00	12.50	12.50
L-threonine	0.35	0.40	0.35	0.35
DL-methionin	0.16	0.30	0.16	0.12
L-iso leucin	0.00	0.30	0.00	0.00
Corn Starch	0.27	0.86	0.28	0.34
Yttrium oxide	0.02	0.02	0.02	0.02
<b>Iron sources</b>				
<i>Bovine hemoglobin</i>	--	6.200	--	--
<i>Iron sulfate hydrate (20%)</i>	0.101	--	--	--
<i>Iron proteinate</i>	--	--	0.127	--
<i>Iron methionate</i>	--	--	--	0.106
<b>Check</b>	100.00	100.00	100.00	100.00

# Material and Methods

## Formulations

- isonitrogenous
- isoenergetic
- equal in

\*AA comp. **KJ12**

\*total iron **KJ9**

\*calcium

\*taurine **KJ11**

\*vitamin B12

content.

Iron source		<i>Iron sulphate</i>	<i>Heme</i>	<i>Iron proteinate</i>	<i>Iron methionate</i>
DM	(g.kg <sup>-1</sup> )	909	913	920	917
ASH	(g.kg <sup>-1</sup> .dm)	94	92	94	94
CP	“”	661	655	669	661
EE	“”	130	122	130	127
CF	“”	6.6	6.6	5.4	5.5
NFE	“”	17.9	37.9	22.2	30.4
Check	----	909	913	920	917
GE	(MJ.kg <sup>-1</sup> )	21.2	21.1	21.5	21.3
CP/GE	----	31.1	31.0	31.1	31.0

Vit B12	(ug.kg <sup>-1</sup> .dm)	1254	1150	959	1075
Fe	(mg.kg <sup>-1</sup> .dm)	355	323	299	349
Ca	(g.kg <sup>-1</sup> .dm)	8	8	8	8
Na	“”	19	19	19	19
K	“”	6	6	6	6
Mg	“”	2	2	2	2
Cu	(mg.kg <sup>-1</sup> .dm)	12	12	12	12
Co	“”	2	2	2	2
Cr	“”	5	5	4	5
Mn	“”	36	35	38	38
Mo	“”	1	1	1	1
Zn	“”	119	115	121	120

<sup>1</sup>DM=Dry matter, ASH=Ash, CP=Crude Protein, EE=Ether extract, CF=Crude Fibre, NFE=Nitrogen Free Extract, GE=Gross Energy



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- KJ9** iron content was formulated to be equal to the iron content in ragworm (352 mg.kg-1.dm).  
Kals, Jeroen; 18/08/2016
- KJ11** Taurine can affect haematological parameters  
Kals, Jeroen; 18/08/2016
- KJ12** Individual amino acids were added to compensate for the amino acid content of the iron sources, especially of the heme iron.  
Kals, Jeroen; 18/08/2016

# Analyses



- **Hct**: centrifuging blood (5 min, 5000g)
- **Hb**: colorimetric (van Kampen & Zijlstra 1961)
- Minerals:
  - ICP-AES (e.g. Cu, Fe, Mn, Zn) **KJ25**
  - ICP-MS (e.g. Co & Mo) **KJ26**
- Calculation apparent absorption coefficient (**AAC**)

$$AAC_{\text{mineral}(x)} = 100 - \left(100 * \left(\frac{Yttrium_{\text{diet}}}{Yttrium_{\text{faeces}}}\right) * \left(\frac{\text{mineral}(x)_{\text{faeces}}}{\text{mineral}(x)_{\text{diet}}}\right)\right)$$

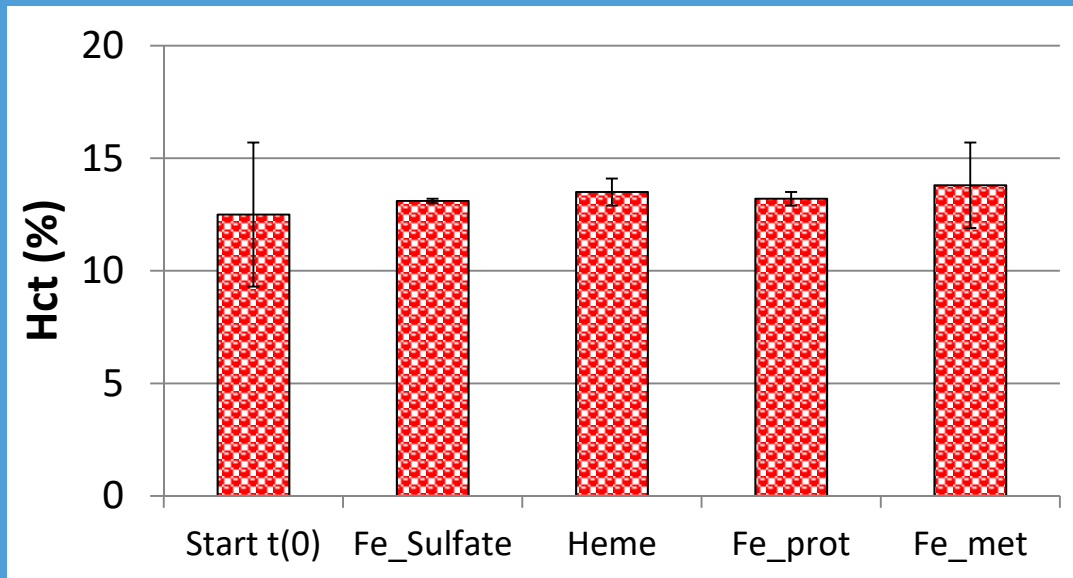
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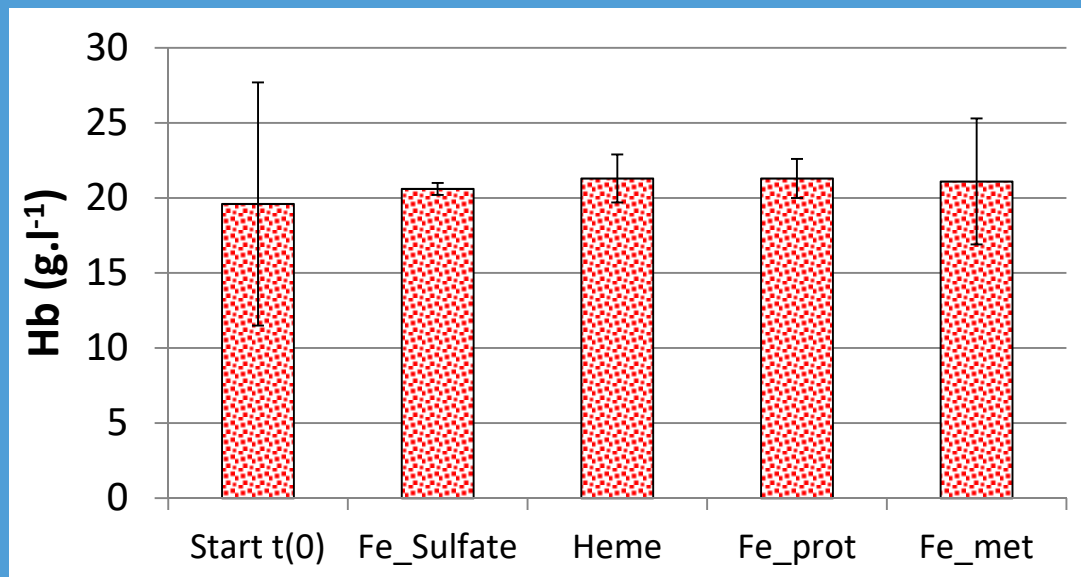
**KJ25** inductively coupled plasma-atomic emission spectrometry  
Kals, Jeroen; 18/08/2016

**KJ26** Inductively Coupled Plasma Mass Spectrometry  
Kals, Jeroen; 18/08/2016

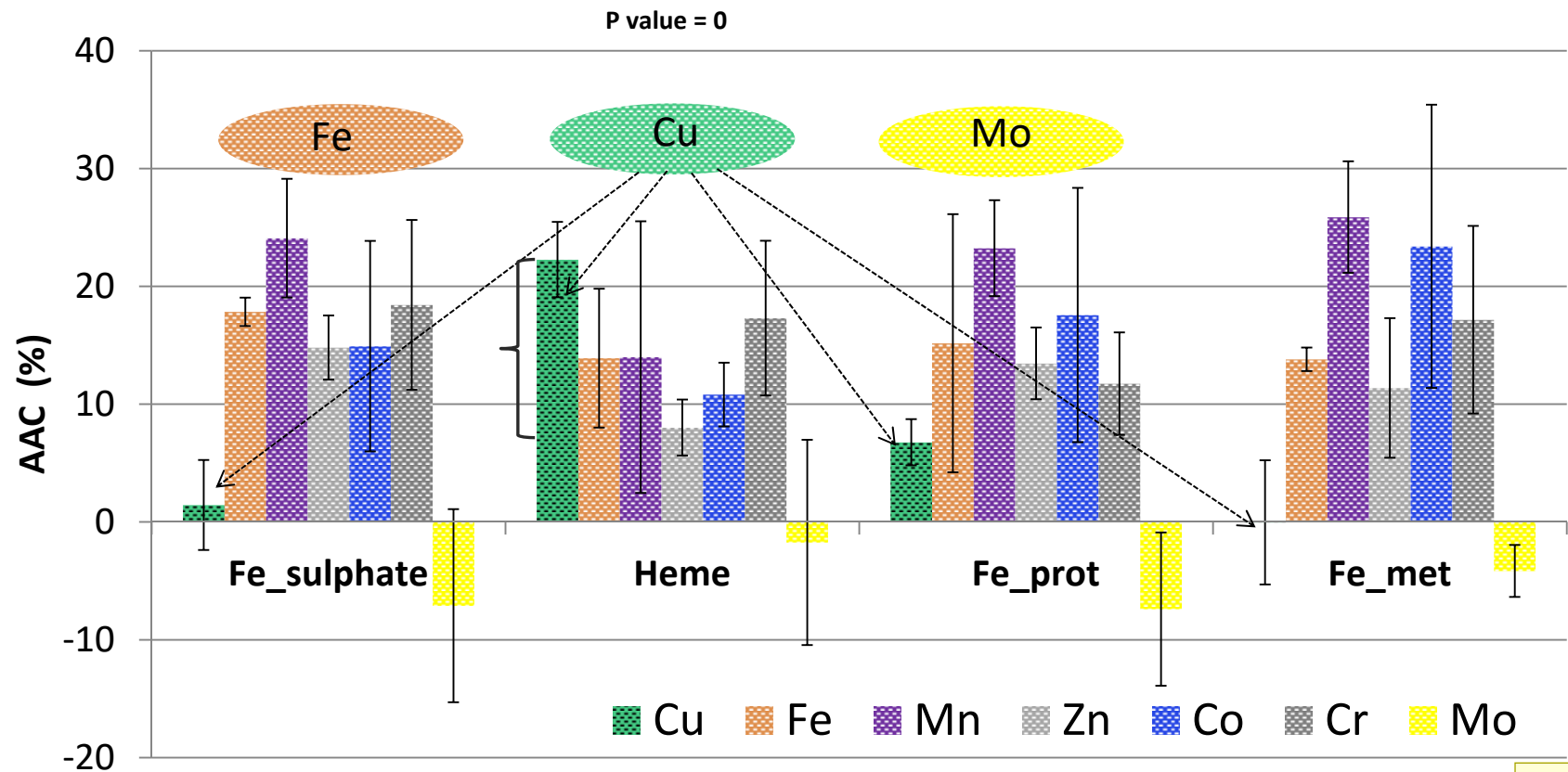
# Results Hct and Hb



- Hct & Hb levels **not** different from t(0)
- Hct & Hb levels **not** affected by source



# Results AAC's



KJ6  
KJ16

- AAC Fe, Mn, Zn, Co, Cr, Mo, except Cu not affected by source

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**KJ6**

Yet, the iron absorption was high for all sources.

The AAC of Cu was 15-22% higher in sole fed the diet with heme.

Kals, Jeroen; 18/08/2016

**KJ16**

The negative ADC's of Mo are most likely a result from the relatively high presence of Mo in seawater and the fact that marine fish must drink to keep their water balance in order

Kals, Jeroen; 27/07/2016



# Discussion



- Despite the **high** absorption of iron, fish stayed anaemic **independent** of iron source
  - > Implies nutritional anemia is not an iron deficiency anaemia
- The **alkaline** character of the sole's intestine would **hamper** the ability to absorb non heme iron
  - > Iron absorption in relation to **iron source** seems not a limiting factor

# Discussion



## ■ Hypotheses

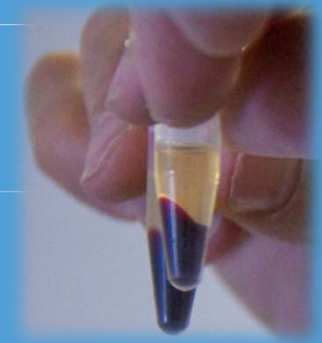
- 1) **nutritional** anaemia in sole fed CPEL is due to an iron deficiency anaemia,
- 2) NA is caused by **inadequate** absorption of iron,
- 3) **Rise** of iron absorption due to a higher bioavailability of heme and/or iron chelates alleviates anaemia in sole  
**all** have to be **rejected!**
- 4) Hence, hypothesis 4: Hct & Hb levels, follow iron absorption patterns could not be tested

# Discussion



- AAC of **Cu** was high using heme
- none heme Fe & Cu, need DMT1
- Heme is not claiming DMT1 capacity
  - > Pos. effect of heme on Cu can be explained by a reduced competitive binding of Cu and non heme Fe claiming DMT1
- Cu is **crucial** for iron uptake
- AAC of Cu is high in sole fed heme, yet **Hct** & **Hb** did not respond
  - > nutritional anaemia is not a Cu deficiency anaemia

# Conclusions



- Iron **absorption** is high & **independent** of iron source
- Heme iron **increases** absorption of **copper**
- High absorption of Fe & Cu in sole fed heme did **not** affect **Hct** and **Hb**, suggesting the nutritional anaemia in sole is
  - > not an iron
  - > nor a Cu deficiency anaemia

# References

- Kals J., Blonk R.J.W., Mheen H.W. van der, Schrama J.W. & Verreth J.A.J. (2015<sup>a</sup>). Feeding ragworm (*Nereis virens* Sars) increases haematocrit and haemoglobin levels in common sole (*Solea solea* L.). *Aquaculture Research*, DOI: 10.1111/are.12767.
- -Kals J., Blonk R.J.W., Palstra A.P., Sobotta T.K., Mongile F., Schneider O., Planas J.V., Schrama J.W. & Verreth J.A.J. (2015<sup>b</sup>) Feeding ragworm (*Nereis virens* Sars) to common sole (*Solea, solea* L.) alleviates nutritional anaemia and stimulates growth. *Aquaculture Research*. <http://dx.doi.org/10.1111/are.12919>.
- Kals J., Blonk R.J.W., Mheen H.W. van der, Schrama J.W. & Verreth J.A.J. (2015<sup>c</sup>). Effect of different iron sources on the alleviation of nutritional anaemia in common sole (*Solea, solea*). *Aquaculture*. 451: 266-270. <http://dx.doi.org/10.1016/j.aquaculture.2015.08.036>
- van Kampen E.J. & Zijlstra W.G. (1961). Standardization of hemoglobinometry II. The hemiglobincyanide method. *Clinica Chimica Acta* 6, 538–544.

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# Thank you!

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Thanks to everybody  
who helped me with  
the presented work,  
yet especially,

\*Co-authors

\*Animal caretakers

Questions?

