

The perspectives of genetically modified livestock in agriculture and biomedicine

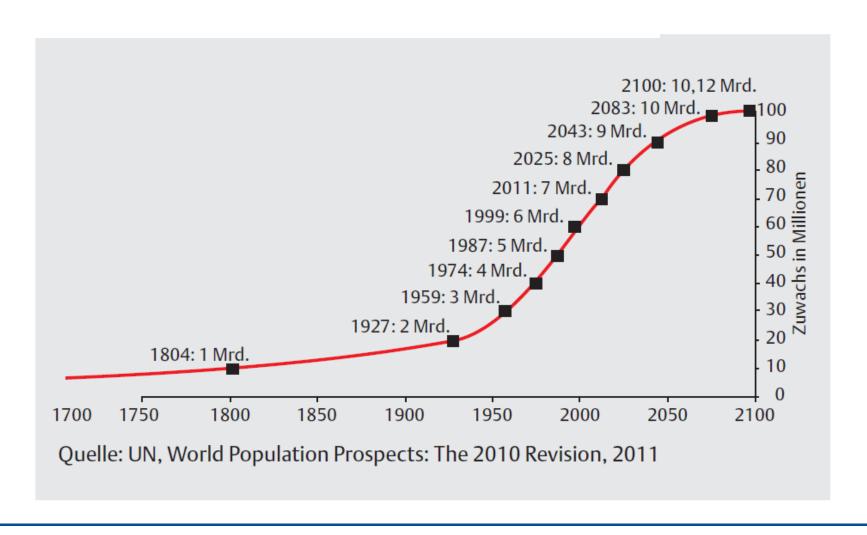
- Agricultural perspectives
- Biomedical perspectives

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Projected increase in human global population



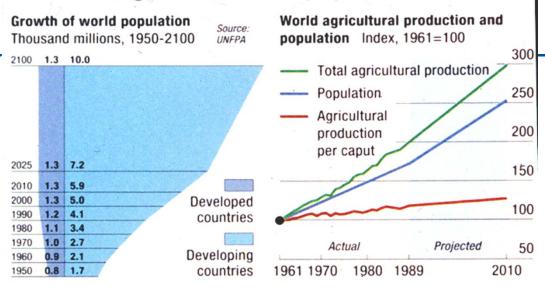


Selected constraints of agricultural production

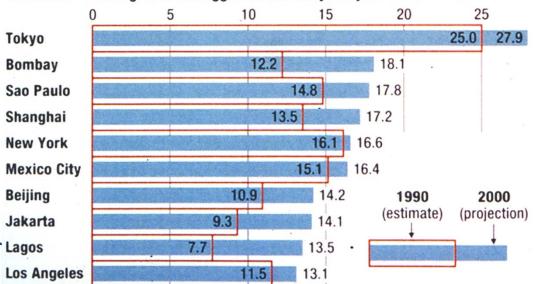
- ~5% of global land is usable for agriculture
- Increase in affluency if several parts of the world is associated with changes in diet towards more valuable animal proteins
- Environmental constraints of livestock production (~1/3 of climate relevant emission comes from agriculture)
- FAO: 1.3 kg CO_{2eq}/milk (Northern America, Europe);
 7.5 CO_{2eq}/milk (Africa)
- Consequences: Food production needs to be doubled or tripled
- **Need**: higher productivity without detrimental side effects (sustainable intensification).



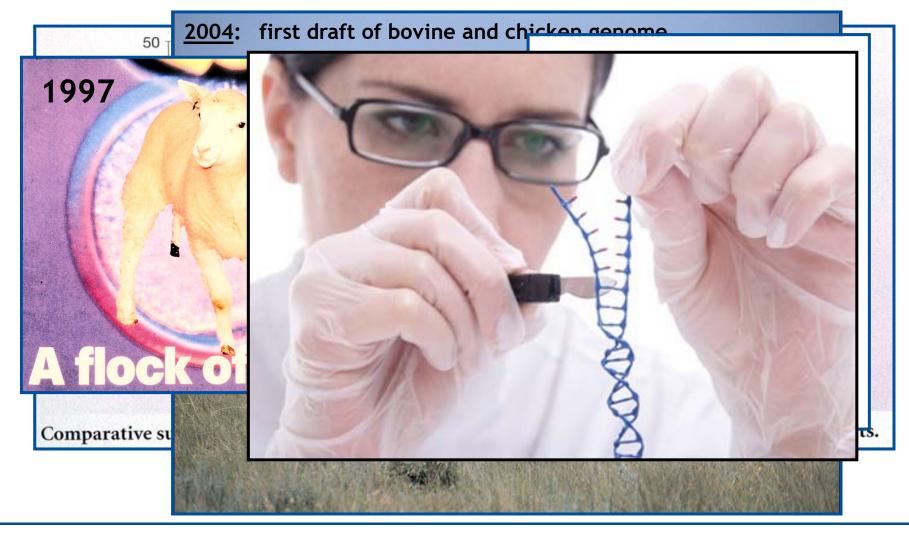
Agriculture and population







A new era in biology: Genome sequencing, somatic cloning and embryonic stem cells





Agricultural perspectives of genetically modified farm animals

- Growth and development (myostatin, GH, GHrec, IGF)
- Wool production
- Lactation (amount, composition)
- Hornless cattle (Polled locus)
- Disease resistance (Mx-gene, IgA, BSE, TB, PRRS, etc)
- Reproduction
- Environmental improvements (Phytase)
- Dietetic improvements
- Skewing the gender



Improvements in economically important parameters of growth hormone transgenic swine

Constructs

Weight gain

Feed efficiency

Backfat thickness

"Side effects " GH)

mMTI-bGH*

+ 23

+ 18

7.5 mm (from 21 mm)

+++

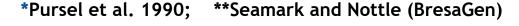
hMTI-pGH(cDNA)**

10 - 20%

10 - 15%

significantly reduced

- (30 - 40ng/ml

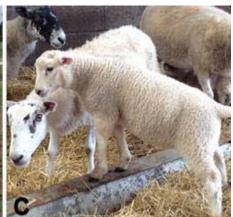




Cattle and sheep with TALEN induced knockout of the myostatin gene







Nelore WT	GTGATGAACACTCCACAGAATCTCGATGCTGTCGTTACCCTCTAACTGTGGATTTTGA	
Bull 1 Allele 1 Bull 1 Allele 2 Bull 1 Allele 3	GTGATGAACACTCCACAGAATCTCGATGCTGTCGTTACCCTCTAACTGTGGATTTTGA GTGATGAACACTCCACAGAATCTCGATGCTGTTACCCTCTAACTGTGGATTTTGA GTGATGAACACTCCACAGAATCTCGATGC-GTCGTTACCCTCTAACTGTGGATTTTGA	WT ΔR283 Δ1
Heifer Allele 1 Heifer Allele 2	${\tt GTGATGAACACTCCACAGAATCTCGATGCTGTCGTTACCCTCTAACTGTGGATTTTGAGTGATGAACACTCCACAGAATCTCGATGCTGTCGTTACCCTCTAACTGTGGATTTTGAGTGATTTTGAGTGATTACCCTCTAACTGTGGATTTTGAGTGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATGA$	WT WT
Bull 2 Allele 1 Bull 2 Allele 2	${\tt GTGATGAACACTCCACAGAATCTCGATGCTGTCGTTACCCTCTAACTGTGGATTTTGAGTGATGAACACTCCACAGAATCTCGATGTCGTTACCCTCTAACTGTGGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGATTTTGAGTGAGTGATTTTGAGTGATTGAGTGAGTGATTTTGAGTGAGTGATGA$	WT ΔC281
Bull 3 Allele 1 Bull 3 Allele 2	GTGATGAACACTCCACAGAATCTCGATGCTGTCGTTACCCTCTAACTGTGGATTTTGA GTGATGAACACTCCACAGAATCTCGAAGGACAG	WT Δ219 +7
Sheep WT	${\tt GTGATGA\underline{G}CACTCCACAGAATCT}{\tt CGATGCTGTCGTTACCCTCTAACTGTGGATTTTGA}$	
Sheep Allele 1 Sheep Allele 2	$\label{eq:gtgatga} \textbf{GTGATGA}\underline{\textbf{G}}\textbf{CACTCCACAGAATCTCGATGCTGTCGTTACCCTCTAACTGTGGATTTTGA}\\ \textbf{GTGATGA}\underline{\textbf{G}}\textbf{CACTCCACAGAATCTCGATGCTGTTACCCTCTAACTGTGGATTTTGA}$	WT ΔR283



Myostatin knockout pigs after employing TALEN



Produced in Korea 2015



Gene-edited minipigs as pets



BGI announced its plan to sell the micropigs as pets at a summit in Shenzhen, China.



Transgenic animals with improved fibre production

Introduced modification	Application	Species	Reference
Ovine insulin-like growth factor 1	6.2% more fleece	Sheep	Damak et al. 1996
Ovine growth hormone	Improved wool production	Sheep	Adams et al. 2002
Ovine keratine intermediate filament	Improved wool processing and wearing properties	Sheep	Bawden et al. 1998
Bacterial serine transacety- lase and O-acetylserine sulfhydrylase	Improved wool production	Sheep	Ward 2000



Genetically modified animals with improved milk production

Introduced modification	Species	Reference	
Bovine a-lactalbumin	Increase milk yield and piglet survival	Pig	Wheeler et al. 2001
Bovine b-and k-casein	Improved milk composition	Cattle	Brophy et al. 2003
siRNA-ß-lactoglobulin	reduced allergenicity	Cattle	Jabed et al. 2012
ß-lac-GFP-neo- L ycostaphin	improved udder health	Cattle	Wall et al. 2005
hlysozyme in ß-cas locus	improved udder health	Cattle	Liu et al. 2014



Production of ß-lactoglobulin free milk in siRNA-ß-lac transgenic cows

Targeted microRNA expression in dairy cattle directs production of β -lactoglobulin-free, high-casein milk

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Edited by R. Michael Roberts, University of Missouri, Columbia, MO, and approved August 28, 2012 (received for review June 22, 2012)

Milk from dairy cows contains the protein β-lactoglobulin (BLG), which is not present in human milk. As it is a major milk allergen, we wished to decrease BLG levels in milk by RNAi. In vitro screening of 10 microRNAs (miRNAs), either individually or in tandem combinations, identified several that achieved as much as a 98% knockdown of BLG. One tandem construct was expressed in the mammary gland of an ovine BLG-expressing mouse model, resulting in 96% knockdown of ovine BLG in milk. Following this in vivo validation, we produced a transgenic calf, engineered to express these tandem miRNAs. Analysis of hormonally induced milk from this calf demonstrated absence of BLG and a concurrent increase of all casein milk proteins. The findings demonstrate miRNA–mediated depletion of an allergenic milk protein in cattle and validate targeted miRNA expression as an effective strategy to alter milk composition and other livestock traits.

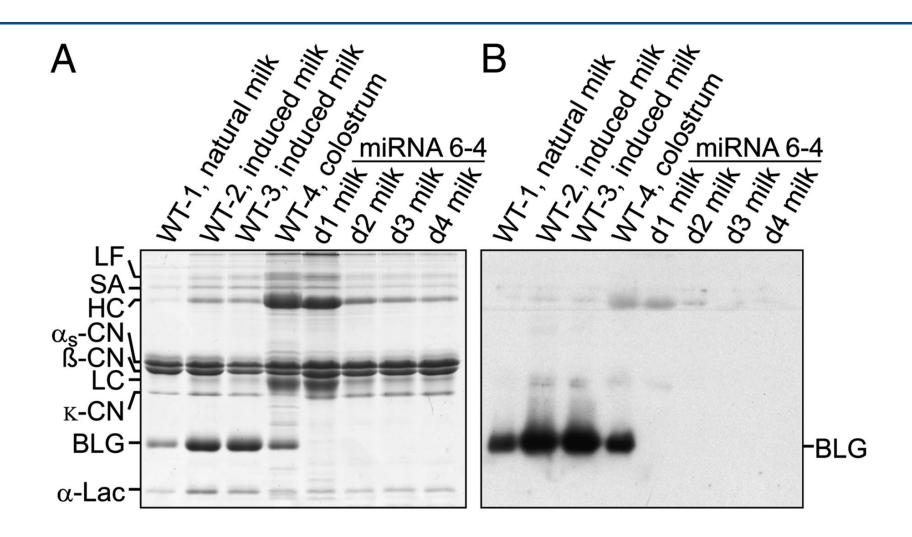
nuclear transfer | transgenic cattle

allergenicity of cows' milk (9). Moreover, RNAi could allow finetuning of BLG expression, which may be advantageous if some BLG is required for normal milk physiology. Artificial RNAi molecules that enable the knockdown of target transcripts, either by mRNA degradation or a block of translation, have been used in different forms such as siRNAs, shRNAs, or miRNAs (12, 13).

A recent in vitro study demonstrated the effectiveness of several shRNAs and miRNAs directed against the porcine variant of BLG (14). We used artificial miRNAs based on the murine miRNA-155 to knock down BLG. miRNAs can be driven by Pol II promoters, which enable spatiotemporally restricted expression and greatly limit off-target effects that may arise from constitutive miRNA expression. Indeed, constitutive expression of BLG-specific RNAi constructs negatively affects primary cell growth, indicating that abundant interfering RNAs aimed at BLG may be toxic (14). When the same RNAi constructs were controlled by a lactation-specific promoter, they showed no ad-



miRNA-mediated depletion of BLG in bovine milk



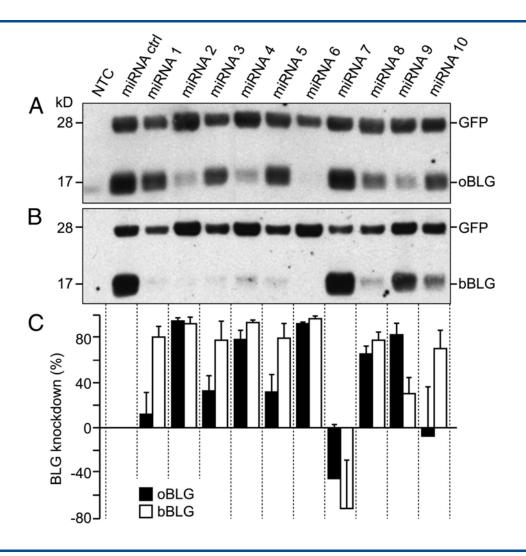


Targeted microRNA expression in dairy cattle directs production of B-lactoglobulin-free, high-casein milk

Cow	Milk Tota	l Casein,	mg/g	Wh	ey, mg/g	
		·	a-Lac		BLG-b	Total
miRNA 6-4	Induced, day 1	98.2	3.9 3.5	0.0	0.0 0.0	3.9 3.5
miRNA 6-4 miRNA 6-4	Induced, day 2 Induced, day 3	96.8 106.6	4.3	0.0	0.0	4.3
miRNA 6-4	Induced, day 4	128.6	5.3	0.0	0.0	5.3
WT-1	Natural, day 69	39.6	1.5	5.7	0.6	7.8
WT-2	Induced, day 5	38.8	1.5	7.6	0.7	9.8
WT-3	Induced, day 5	32.5	1.5	7.3	0.9	9.4
WT-4	Colostrum, day 1	48.1	1.7	10.1	4.0	15.7
SEM-*	-	1.27	0.09	0.12	0.11	0.12



In vitro knockdown of BLG in COS-7 cells.





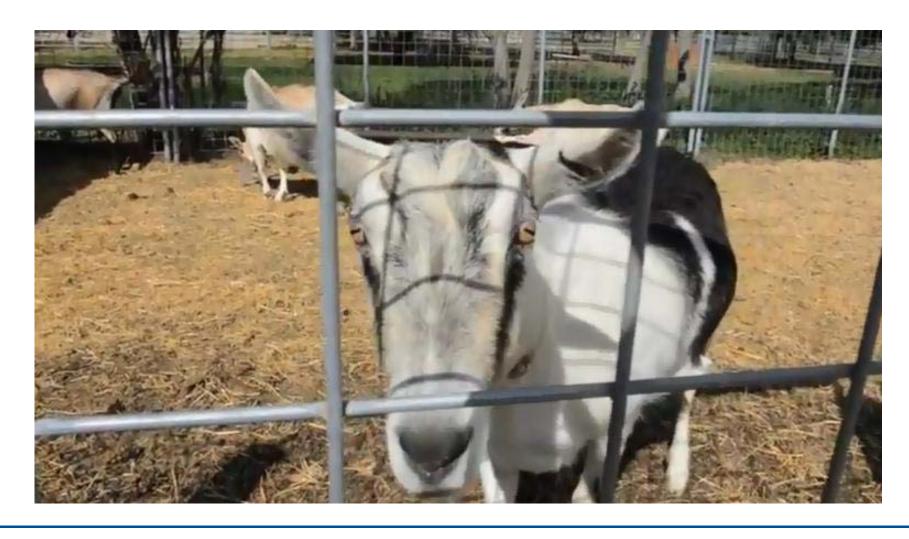
Production of cattle with elevated concentration of B- and k-Casein in milk

Table 2. Milk composition of transgenic and nontransgenic cows								
Cow	Cell line	Age at induction, months	Protein ^b , %	Casein (CN) ^a , mg/ml	β-CNª, mg/ml	κ-CNª, mg/ml	β-CN:CN	κ-CN:CN
A Cow	s induced	in July 2001						
TG2	TG2	8	5.6	44.5	15.8	11.2	0.36	0.26
TG3-1	TG3	7	6.9	54.9	19.9	10.7	0.36	0.20
TG3-2	TG3	7	5.2	42.0	17.6	11.6	0.42	0.29
TG3-3	TG3	8	5.9	47.5	20.9	12.0	0.44	0.26
TG3-4	TG3	7	5.9	47.2	18.4	10.1	0.39	0.22
TG3-5	TG3	7	5.3	43.0	14.1	8.4	0.33	0.20
Mean TG3			5.8	46.9	18.2	10.6	0.39	0.24
CC-1	NA	10	4.6	36.2	14.3	5.1	0.40	0.15
CC-2	NA	10	5.0	39.6	14.8	5.8	0.37	0.15
Mean			4.8	37.9	14.6	5.5	0.39	0.15
B Cow	s induced	in Decembe	r 2001					
TG3-6	TG3	9	4.5	33.8	14.0	10.7	0.41	0.32
TG3-7	TG3	9	6.4	34.8	17.8	13.0	0.51	0.37
TG3-8	TG3	9	4.5	33.8	17.0	14.1	0.50	0.42
TG5	TG5	9	3.8	25.6	12.2	5.7	0.48	0.22
TG7	TG7	7	4.5	25.3	13.9	5.6	0.55	0.22
Mean TG3			5.1	34.1	16.3	12.6	0.48	0.36
BFF-1	BFF	7	3.8	26.4	14.4	5.0	0.55	0.19
BFF-2	BFF	7	5.1	24.0	14.9	5.0	0.62	0.21
BFF-3	BFF	7	4.5	25.5	14.5	5.0	0.57	0.20
Mean			4.5	25.3	14.6	5.0	0.58	0.20

^aBased on skim milk. ^bBased on whole milk. Milk samples were collected after hormonal induction in July 2001 (A) and December 2001 (B). Milk from cows induced in July 2001 was analyzed for total protein and casein (CN) using infrared spectroscopy and milk from cows induced in December 2001 was analyzed by total combustion (protein) and HPLC (CN). β-and κ-casein concentrations were determined nephelometrically.

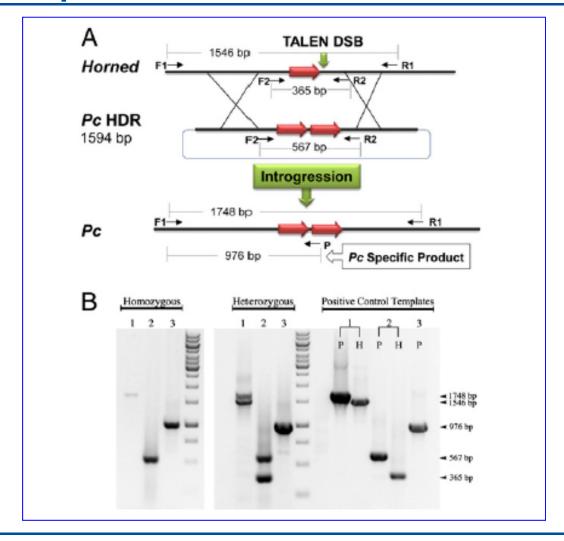


Milk from hLZ transgenic goats helps children with diarrhea





TALEN induced mutation of the Polled locus to produce cattle without horns





Gene-editing of Polled locus: Spotiguy, born 2015, with two of his clones



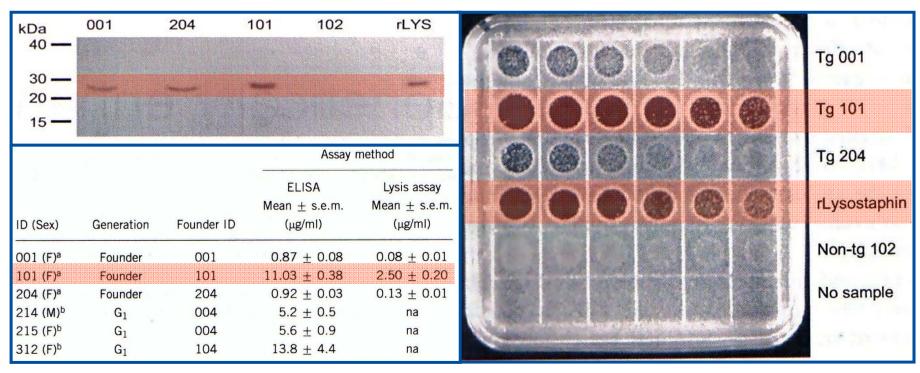


Approaches towards animals transgenic for enhanced disease resistance

- General:
- Mx 1 protein (pigs)
- Immunoglobulin A (pigs)
- Visna virus envelope (sheep)
- Transmissible Gastroenteritis virus (TGEV; mammary gland specific mouse model)
- Knockout of specific genes (f.ex. Prion; PRRS, PERVs)
- siRNA mediated knockdown of pathogenic virus expression
- Mammary gland:
- a-Lactalbumin (pigs)
- Lysozyme (goats, cattle): antimicrobial effects
- Lactoferrin (cattle): bacteriostatic, bacteriocidic, iron provider
- Lycostaphin-transgenesis: St. aureus resistant cows



Transgenic cattle with mit Lysostaphin induced resistance of the mammary gland against infections with St. aureus



resistance against *St. aureus* infusions: Tg: 18/21(85.3%) (Infection only after very high doses) vs. WT 13/47 (31.7%)



Mastitis resistant cows by transgenic expression of human lysozyme expression from the B-casein locus

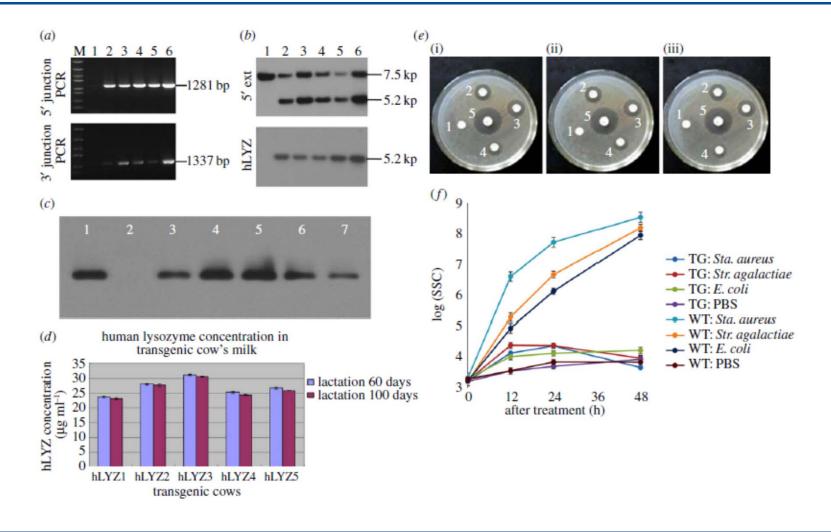
Table 4. Infection rate of three types of bacterium infused into mammary glands of five transgenic and five non-transgenic lactating cows. During each challenge experiment, each gland was infused with one of the three types of bacterium and the fourth gland was infused with PBS. TG, transgenic cows; WT, non-transgenic cows.

		mammary glands infected ^a	number of bacteria (×10³ CFU ml ⁻¹)			
group	mammary glands treated		0 h	12 h	24 h	48 h
TG	5 (Sta. aureus)	0	0	0	0	0
TG	5 (Str. agalactiae)	0	0	0	0	0
TG	5 (E. coli)	0	0	0	0	0
TG	5 (PBS)	0	0	0	0	0
WT	5 (Sta. aureus)	5	0	1.9 ± 0.4	3.2 ± 0.7	4.8 ± 0.5
WT	5 (Str. agalactiae)	4	0	1.4 ± 0.3	5.9 ± 0.8	5.7 ± 0.7
WT	5 (E. coli)	5	0	1.6 ± 0.2	4.5 ± 0.6	4.1 ± 0.8
WT	5 (PBS)	0	0	0	0	0

alnfection was defined as bacterium growth in two consecutive milk samples collected 12-24 h apart.



Mastitis resistant cows by transgenic expression of human lysozyme expression from the B-casein locus





Mastitis resistant cows by transgenic expression of human lysozyme expression from the B-casein locus

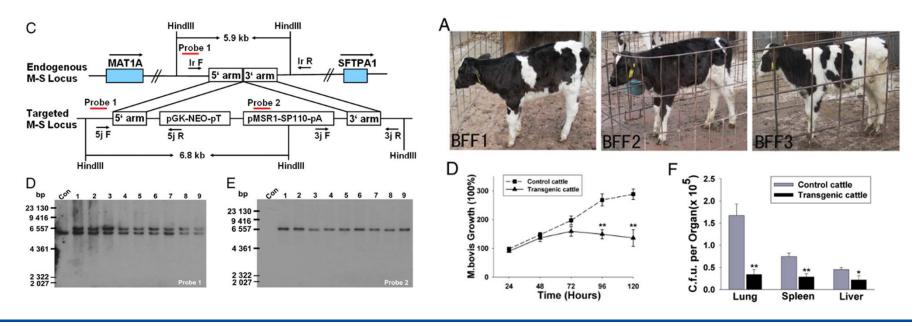
Table 3. Raw components of transgenic milk compared with conventional milk. No significant differences were detected between transgenic and non-transgenic groups (p > 0.05).

components (g 100 ml ^{—1})	transgenic (n = 5)	non-transgenic (n = 5)
fat	4.38 ± 0.39	4.47 <u>+</u> 0.35
protein	3.62 ± 0.28	3.53 ± 0.24
lactose	4.69 ± 0.21	4.81 <u>+</u> 0.38
solids	13.89 ± 0.77	13.55 ± 0.69



Increased Resistance against Tuberculosis via gene editing (TALEN) and transgenic technology

- The mouse *SP110* gene can control *M.bovis* growth in macrophages and induce apoptosis in infected cells.
- Transfer of the mouse *SP110* gene into the genome of Holstein-Friesian (Macrophage Scavenger Receptor (*MSR1*)-locus) by TALENs led to an increased resistance against *M.bovis* infection by macrophage-specific expression of *SP110*.





Cattle resistant against mycobacterium tuberculosis infection, produced via gene editing and transgenic technologies

Table 2. Gross pathology of transgenic cattle challenged with *M. bovis* by endobronchial instillation

Animal	No. of lobes infected*	Lung score	No. of lymph nodes infected [†]	Lymph node score	Total pathology score	Mean [‡]
Transgenic 1	2	4	3	4	8	6.5
Transgenic 2	1	2	2	3	5	
Transgenic 3	0	0	0	0	0	
Control 1	5	21	6	14	35	32.0
Control 2	4	15	8	18	33	
Control 3	4	14	6	14	28	

^{*}Lung lobes (left apical, left cardiac, left diaphragmatic, right apical, right cardiac, right diaphragmatic, and right accessory lobes) were examined for lesions using a gross pathology scoring system.



[†]Lymph nodes (mandibular, parotid, medial retropharyngeal, mediastinal, tracheobronchial, hepatic, mesenteric, and prescapular lymph nodes) were examined for lesions using a gross pathology scoring system.

[‡]Median values per group (n = 3). Only animals with lesions were taken into account.

Cattle with resistance to BSE after knockout of the prion locus

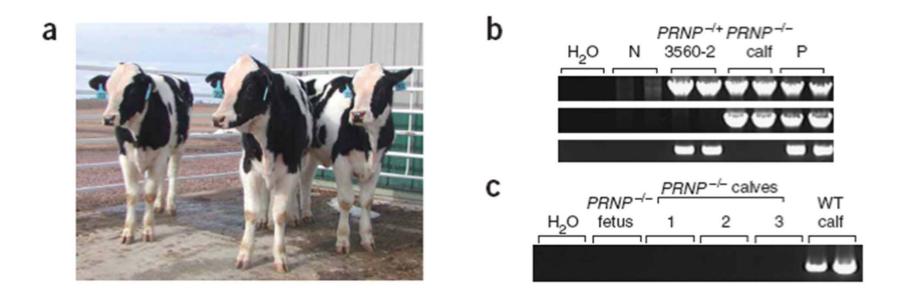


Figure 1 Generation of PRNP^{-/-} cattle. (a) PRNP^{-/-} cattle at 13 months of age. (b) Verification of PRNP^{-/-} genotype in the ear biopsy fibroblasts by genomic PCR. P, positive control⁶; N, negative

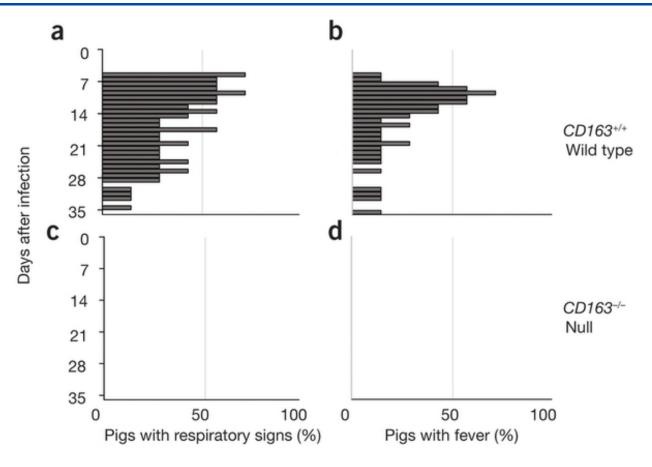


CRISPR/Cas-derived CD 163 knockout piglets were kept together with WT-piglets. All animals were infected with PRRS virus. Five days later, the WT-piglets showed typical PRRSV symptoms, while CD 163 KO piglets remained completely healthy.

(PRRS: Porcine reproductive and respiratory syndrome virus, *CD163* is a macrophage differentiation antigen belonging to the scavenger receptor cysteine-rich (SRCR) family of membrane proteins)

Whitworth et al., Nature Biotechnology 34, 20-22 (2016) Foto: University of Missouri

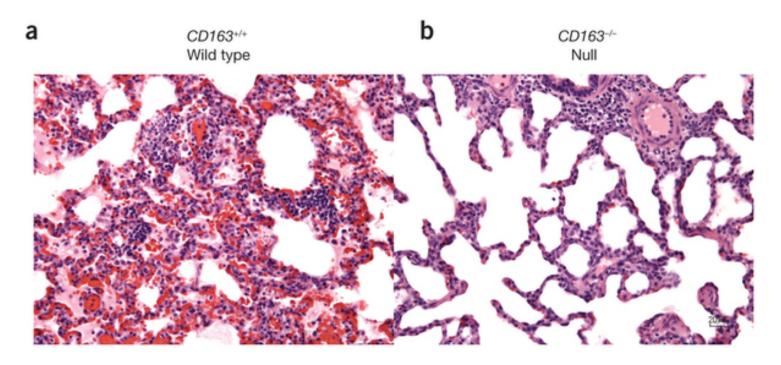




PRRS: Porcine reproductive and respiratory syndrome virus



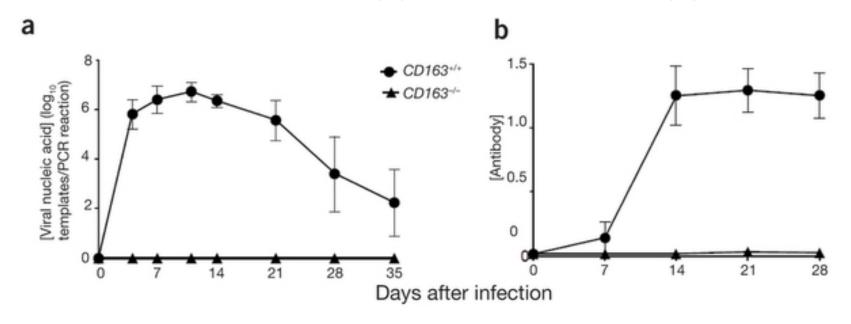
Microscopic image of the lung of CD163+/+ and CD163-/- pigs



PRRS: Porcine reproductive and respiratory syndrome virus



PRRS specific DNA (a) und antibodies (b)



PRRS: Porcine reproductive and respiratory syndrome virus



Gene-editing record smashed in pigs

Researchers modify more than 60 genes (PERV) in effort to enable organ transplants into humans.

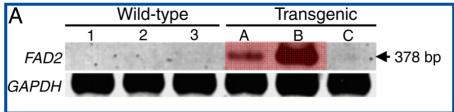


The gene-edited pigs will be raised in isolation from pathogens.

Also ~20 genes altered related to immunology and relevant for Xenotransplantation



Spinach desaturase expression in transgenic pigs alters fatty acid profile in skeletal muscle



Desaturase expression in transgenic pigs leads to meat with more poly-unsaturated fatty acids.

Mol % of total fatty acids	Wild-type	Transgenic
14:0	4.5 ± 1.2	3.5 ± 2.0
14:1	0	0
16:0	38.2 ± 1.3*	26.6 ± 0.4^{b}
16:1	4.4 ± 0.7	6.1 ± 0.5
18:0	21.4 ± 1.8	22.0 ± 0.3
18:1	28.0 ± 0.14	18.8 ± 1.36
18:2n–6	1.9 ± 0.4*	20.3 ± 2.1 ^b
18:3n-3	1.6 ± 0.6	2.7 ± 1.4
20:0	0	0
20:2	0	0
20:4n-6	0	0
20:5n-3	0	0
22:0	0	0
22:1	0	0
22:5n-3	0	0
22:6n-3	0	0
Sum of saturated plus ∆9 unsaturated	96.5 ± 0.3*	77.1 ± 3.5 ^b
Sum of n=6 polyunsaturated	1.9 ± 0.4	20.3 ± 2.16
Sum of n=3 polyunsaturated	1.6 ± 0.6	2.7 ± 1.4
n–6/saturated plus ∆9 unsaturated	0.02 ± 0.04	0.27 ± 0.46
n-6/n-3	2.5 ± 1.7	5.6 ± 0.6



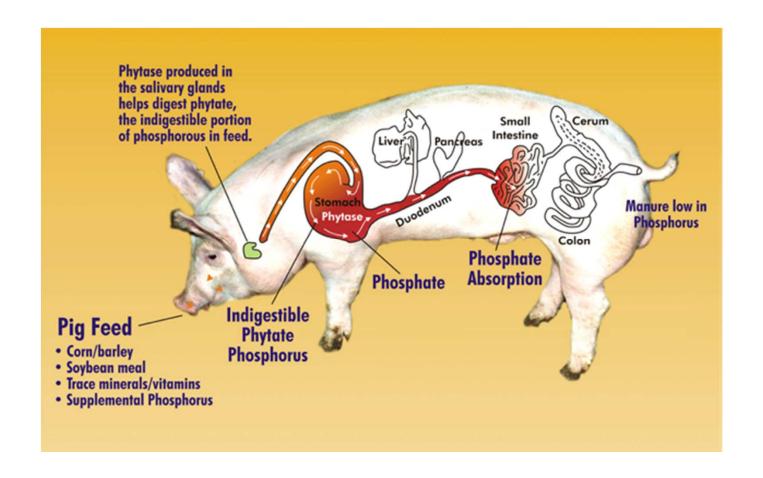
n-3 and n-6 fatty acids concentration and n-6/n-3 ratios in tail samples from *hfat-1* transgenic and wild-type piglets

Fatty acids in tails	Transgenic piglets (n = 8)	Wild-type piglets (n = 8)
ALA (18:3 <i>n</i> -3, %)	0.94 ± 0.10	0.63 ± 0.04
EPA (20:5 <i>n</i> -3, %)	4.21 ± 0.60	0.26 ± 0.07
DPA (22:5 <i>n</i> -3, %)	1.69 ± 0.19	0.35 ± 0.05
DHA (22:6 <i>n</i> -3, %)	1.75 ± 0.23	0.95 ± 0.21
Total <i>n</i> -3 FA (%)	8.59 ± 0.84	2.18 ± 0.25
Total <i>n</i> -6 FA (%)	14.28 ± 1.31	18.46 ± 1.41
<i>n</i> -6/ <i>n</i> -3 ratio	1.69 ± 0.30	8.52 ± 0.62

Lai et al. 2006, Nature Biotechnology

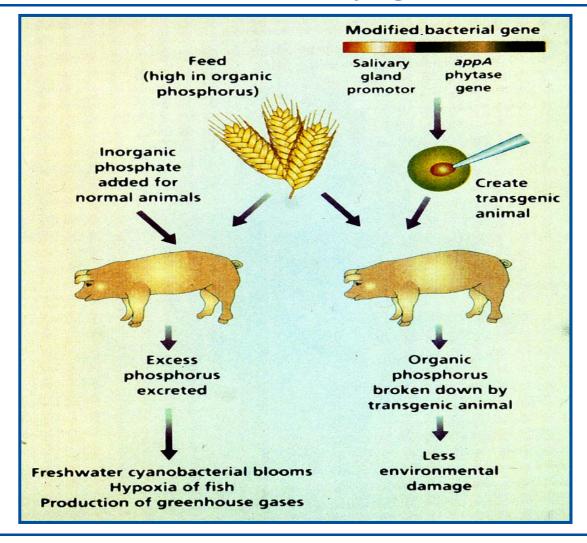


Transgenic pigs with expression of Phytase in salivary gland





Transgenic swine expressing Phytase in the salivary gland





Expression of Phytase in the salivary gland of transgenic pigs

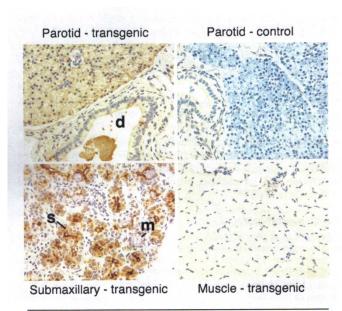


Table 1. True phosphorus digestibility (%) of transgenic phytase pig line WA using soybean meal as the sole source of phosphorus

Pigs	Non-transgenic	Transgenic
Weanling	48.5 ± 5.4°	87.9 ± 3.4 ⁵
vvearining	(n = 16)	(n = 14)
Growing-finishing	51.9 ± 10.3ª	98.8 ± 3.4 ^b
	(n = 16)	(n = 14)

^{a,b}Means in the same row with different superscript letters differ (P < 0.01). True digestibility is the percentage of total phosphorus digested and absorbed from the diets corrected for endogenous phosphorus released from the gastrointestinal tract. Data represent mean \pm s.e.m., as determined by a regression analysis technique 40,42 .

Expression of phytase in the salivary gland of transgenic pigs

Reduction of anorganic phosphorus feeding via improved metabolism

Reduction of phosphorus excretion by up to 75%



Reduction of costs

Environment protection



Transgenic animals with agriculturally important traits

Lactational performance

Transgenic cattle: lactoferrin, lysozyme, caseins; but problems in some of the mouse models with milk production, <u>Mariensee</u>: Lactase transgenic mice with significant effects on lactose levels

Transgenic pigs: bovine α-lactalbumin: elevated lactose levels better piglet performance.

Dietetic improvements

Transgenic pigs >unsaturated fatty acids by introduction of spinach desaturase gene

- Wool shearing
- Reproductive performance

(Estrogen receptor gene, inhibin reduction?)



Biomedical perspectives of genetically modified farm animals

Biomedical perspectives

Gene Pharming (rec. Proteins, mAbs)
Human blood substitute
Xenotransplantation
Inhibitors of chemical weapons

Basic research

Epigenetic reprogramming Models for human diseases



Approved GM vertebrates

- GloFisch, genetically engineered zebrafish, no regulation necessary (FDA statement 2003)
- Atryn (antithrombin III), produced in the mammary gland of transgenic goats, approved by EMA (2006) and FDA (2009)
- Ruconest (C1 esterase inhibitor), produced in the mammary gland of transgenic rabbits, approved by EMA (2010)
- AquAdvantage salmon, added growth hormon from Pacific Chinook salmon, all year long expression, faster growth, approved by FDA (Nov. 2015)







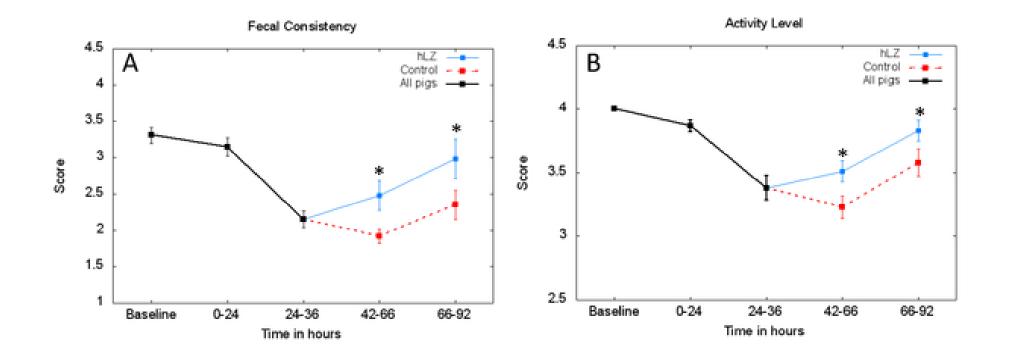
Transgenic chickens are the latest animals engineered to produce 'farmaceutical' drugs.

US government approves transgenic chicken





Pigs fed hLZ- milk have improved fecal consistency and activity scores

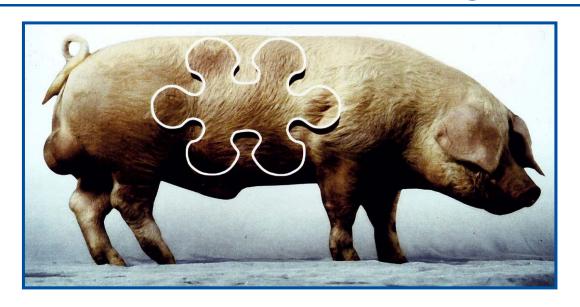


Towards the ultimate donor pig

CHOICE CUTS Researchers are looking to KIDNEY LUNG source an increasing variety A factory farm is being A kidney with six genetic of living tissues, including designed to produce modifications supported a solid organs, from pigs. Many baboon's life for 4 months. 1,000 pig lungs per year. are attempting to genetically engineer the animals to reduce the risk of rejection and infection in humans. CORNEA Pig corneas were PANCREAS approved for marketing in Phase III clinical China in April. trials of insulinproducing islet cells are under way. HEART A genetically modified LIVER pig heart implanted in a baboon's abdomen Livers could be engineered to survived for 2.5 years. produce their own antibodies against primate immune cells.



The domestic pig as a potential donor for human organs

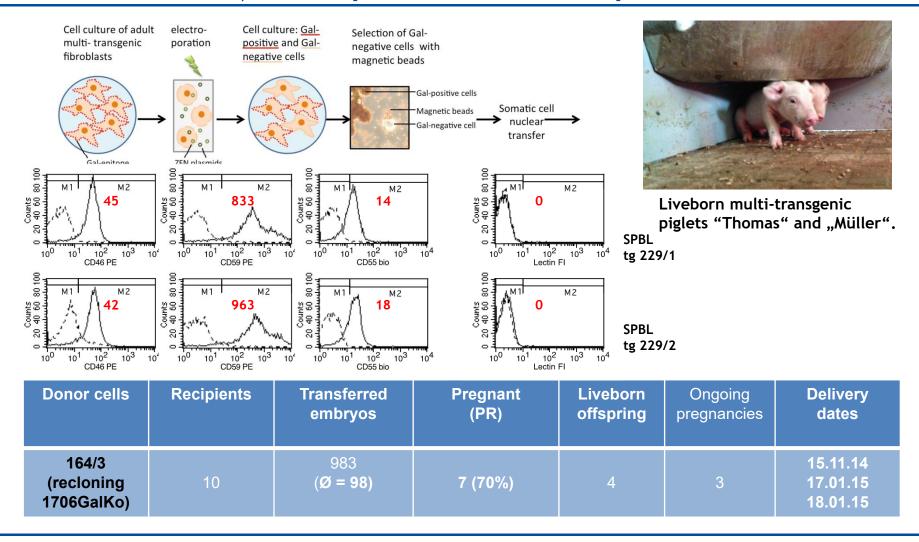


I like pigs; dogs look up to us, cats look down to us, pigs treat us as equal. (Winston Churchill)

- Domesticated species
- High fertility, great abundance, rapid growth
- Genetics, anatomy, physiology not too different from human
- Strict hygienic conditions possible
- Previous success with porcine insulin, heart valves, skin patches
- Genetic modifications possible



Multi-transgenic pigs (GGTA1-KO/hCD46/hCD55/hCD59/hA20/hHO-1) for improved xenotransplantation results





Successful use of TALEN in livestock

A			1650	1651	1652	1653	1654	1655	1656	1657	Wt		1661	1662	1663	1664	1665	1666	Wt		
	ΔΔ	Ξ,	100	-	_	-	-	_		-	-	Ξ	=	=	-	=		=	-	Ξ	4
	D	-										-				•				Marie	◀
		DAZL: BamHI APC: HindIII																			
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	APC	2																			
	Wt- HDR	TCAT TCAC	GGZ	AAGA	AGT	ATCA	GCCF												CAC		
	A1-	TCAT	rgg/	AAGA	AGT	ATCA	GCCF													HDR	
		TCA?			AGI	AICA	GCCF	TICA	1100		- CCC	AGGA	AAGA	CAGA	AAII	CIGO	GICE	AACC	AC	13	
	A1- A2-	TCAT	rgg/	AAGA AAGA	AGT														CAC	HDR Wt	
		TCA				ATCA	GCCF	TTC	ATCC	CTCC	CAG <mark>T</mark>	GAAG	CTT	ACAG	AAAT	TCTG	GGT(CAAC	CAC	HDR	
	A2-	TCAT	rGG/	AAGA	AGT	ATCA	GCCA	TTC	TCC	CTCC	GA::	: AGA	CAG	AAAC	TCTG	GGTC	CAACO	CAC		Δ3	





Application perspectives for genetically modified farm animals

Agricultural perspectives

Growth and development (myostatin, GH, GHrec, IGF)

Wool production

Lactation (amount, composition)

Hornless cattle (Polled locus)

Disease resistance (Mx-gene, IgA, BSE, TB, PRRS, etc)

Reproduction

Environmental improvements (

Dietetic improvements

Biomedical perspectives

Gene Pharming (rec. Proteins, mAbs)

Human blood substitute

Xenotransplantation

Inhibitors of chemical weapons

Basic research

Epigenetic reprogramming

Models for human diseases



Evolution of farm animal breeding

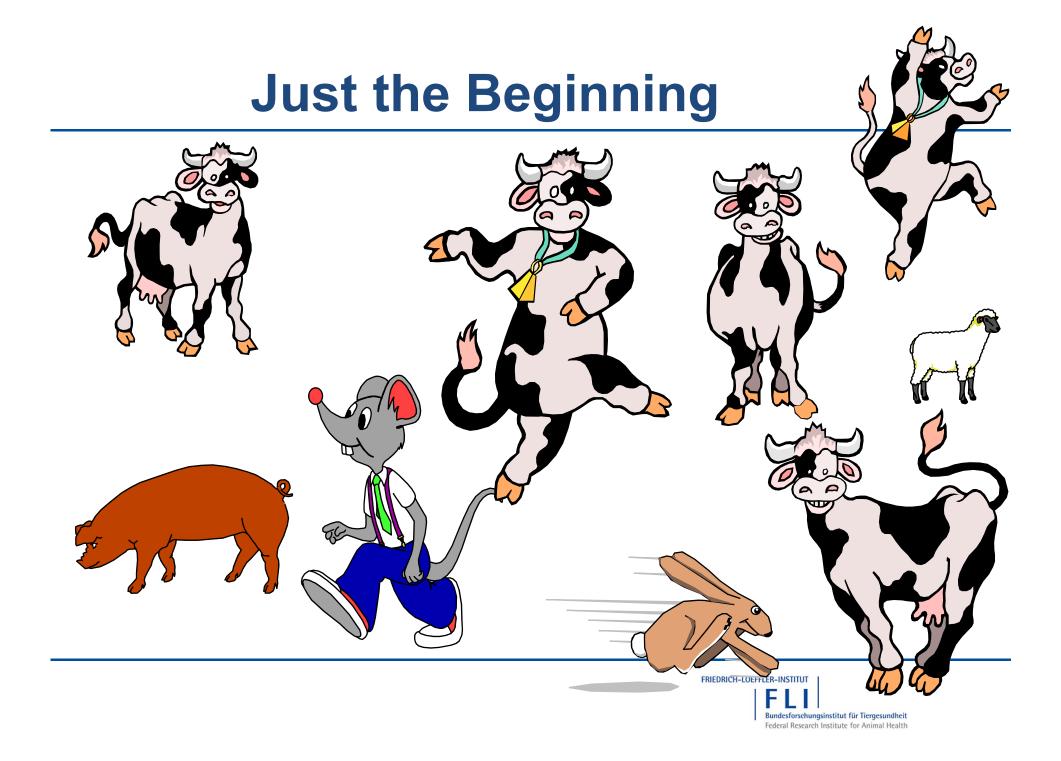
- Domestication
- Proliferation of "useful" populations
- Selection according to the Exterieur
- Selection according to specific traits
- Systematic breeding based on population genetics and statistics
- Reproductive technologies (AI, ET, IVP, SCNT, etc.)
- Molecular genetics and genome based breeding concepts (SNPs, GBV, etc.)
- Now entering the era of *Precision breeding*



The future: Targeted and diversified dairy production

- Full fat normal milk
- Defatted milk (knock-out of key lipid enzymes)
- Curd production (enhanced casein expression)
- Cheese production (enhanced casein expression)
- Coffee whitener and Creme liquor (ß-casein)
- Hypo-allergenic milk (reduced or omitted ß-lactoglobulin)
- Lactose free or -reduced milk (α lactalbumin knock-out, additional lactase expression)
- Infant milk (enhanced lactoferrin expression)
- Improved udder health (lysozyme, etc.)
- Pharmaceutical proteins





Thank you for your attention.









Aquabounty salmon: The first approved genetically engineered animal product



AquAdvantage Atlantic salmon (at back) grow to twice the size of an normal Atlantic salmon (Salmo salar) over the same time.



The practical use of this new genomic information

- Better targeted breeding programmes Genomic breeding values; Direct sequencing
- Transcriptomics/Proteomics/Phenomics
- Production of genetically modified (transgenic) animals
- New knowledge on genetic diversity
- Descent studies
- Comparative genomics



Summary and Conclusions

- The genomes of farm animals have been sequenced and annotated; informative gene maps are available that can be used for breeding purposes (GBV).
- Novel molecular tools, incl. DNA-nucleases such as ZFNs, TALEN, CRISPR/Cas are compatible with precise genetic modifications (gene editing), that can be induced easily and with high efficiency.
- The use of the new genomic information and gene editing tools allow the development of novel breeding strategies, both for agricultural and biomedical purposes.
- Gene editors are also beneficial in human medicine.
- A complex and complicated legal framework is in place for commercial use of transgenic animals. The application of gene editing is not (yet) legally regulated and could thus be immediately employed in future oriented animal breeding systems.



Novel perspectives for animal breeding in agriculture and biomedicine

