



Power and precision of mapping genes in simulated F2 crosses using whole genome sequence data.

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F2 data in the past



Many porcine F2 crosses were established for gene mapping experiments

Divergent lineages as founder breeds for F2 designs

- Distantly related (ASIA x EU, e.g. Rückert & Bennewitz 2010)
- Closely related (EU x EU, e.g. Boysen et al. 2010)

Available datasets

- Exact phenotypes for a lot of traits
- Mostly genotyped with microsatellite markers (low mapping resolution)
- Linkage mapping: no historical meioses can be considered
- \rightarrow many QTL could be found, however, the confidence intervals were large





Can we use F2 data in the era of GENOMICS to precisely map genes?



Current simulation study: What's it about?



Investigation of power and precision in GWAS using simulated sequence data in

- F2 designs with closely related founder breeds (i.e. EU x EU),
- F2 designs with distantly related founder breeds (i.e. ASIA x EU),
- and the pooled data of both designs.

We compare:

- F2 designs derived from closely and distantly related founder breeds
- Impact of pooling F2 data
- Purebred population vs. F2 crosses
- Small vs. large number of founder animals in F2 designs

Phylogeny of the founder breeds (drift model)



F2 crossing scheme





F2 crossing scheme





Individuals to be evaluated







- Causative SNPs (QTN) were simulated in the pool of all individuals to be evaluated
 - \rightarrow Random but known positions
- Variance components are set so that $h^2 \approx 0.5$ is valid for the populations

Single marker regression using Genome-wide Complex Trait Analysis (Yang et al. 2014)

Mixed linear model:

$y_i = \mu * b_j * SNP_{ij} + g_i + e_i$

 y_i phenotype of individual *i*

μ overall mean

- b_j regression coefficient of marker j
- SNP_{ij} gene content of SNP *j* of individual *i*
- g_i random polygenetic effect with $g \sim N(0, G\sigma_g^2)$ and G being the GRM
- e_i residual of individual i
- QTL mapping on chromosome 1 (2)
- Chromosome 2 (1) to model population structure (GRM)
 → SNPs to be tested are excluded from the GRM (MLMe)
- p-values were adjusted using Bonferroni correction, α = 0.01 (genome wide)

Results: Averaged values across all 50 replicates (10 simulations á 5 traits)



Parameter	QTN Power		QTL F	QTL Power		QTN Precision	
Significance level	α = 0.01		α = 0.01		α = 0.01		
	mean	sd	mean	sd	mean	sd	
EU2	0,21	0,08	0,29	0,11	0,90	0,10	
EU1 x EU2 (small F0)	0,26	0,12	0,46	0,17	0,87	0,11	
EU1 x EU2 (large F0)	0,22	0,09	0,32	0,10	0,89	0,11	
ASIA x EU2 (small F0)	0,39	0,20	0,69	0,22	0,69	0,13	
ASIA x EU2 (large F0)	0,36	0,18	0,65	0,23	0,70	0,14	
Joint analysis (small F0)	0,44	0,16	0,82	0,16	0,71	0,11	
Joint analysis (large F0)	0,41	0,14	0,76	0,18	0,75	0,13	

Power increase in F2 data compared to purebred populations: Highest power in pooled data



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Precision: F2 designs with closely related founders almost reach the precision of purebred populations



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Pooling data increases the precision in F2 designs derived from distantly related founder breeds



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Where is the benefit to use (pooled) F2 sequence data in GWAS?



<u>EUxEU</u>

■ Precise mapping results → short LD blocks

<u>ASIAxEU</u>

- High mapping power \rightarrow increased gene frequencies
- Low precision \rightarrow long LD blocks

Pooling data

- Increase in power \rightarrow enlarged sample size
- Increase in precision compared to single analysis of ASIAxEU → reduced LD block length

This is in agreement with Toosi et al. (2009) and Bennewitz and Wellmann (2014).





Can we use F2 data in the era of GENOMICS to precisely map genes?



Something to reflect when taking a shower...



 Applying NGS techniques on F2 data leads to suitable datasets for GWAS at reasonable costs.

 F2 data at a maximum marker density provides a powerful and (cost) efficient possibility to (fine) map genes when the founder breeds are closely related or can be pooled.

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Thank you!

thymine

This study was supported by a grant from the German Research Foundation (Deutsche Forschungsgemeinschaft, DFG).