

Machine learning transcriptome analysis to identify genes associated with feed efficiency in pig

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Molecular mechanisms underlying **feed efficiency** are still unknown

Machine-learning applied into a resampling strategy can provide a good assessment of the generalizability of the results

Objective

To identify genes associated with feed efficiency using transcriptomic (RNA-Seq) data from pigs phenotypically extreme for residual feed intake

$$RFI_{ijk} = FI_{ijk} - \left[S_j + \beta_{MBW_j} \times MBW_i + \beta_{ADG_j} \times ADG_i + \beta_{BFG_j} \times BFG_i \right]$$

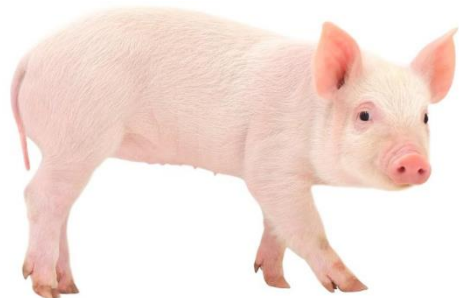
β_{MBW_j} , β_{ADG_j} and β_{BFG_j} partial regressions coefficients (within sex j).

FI: feed intake

MBW: metabolic body weight

ADG: average daily gain

BFG: backfat gain



25 Kg of body weight

IVO-automatic feeders
Body weight and backfat measured at
1, 3, 6, 9, 12 and 15 weeks

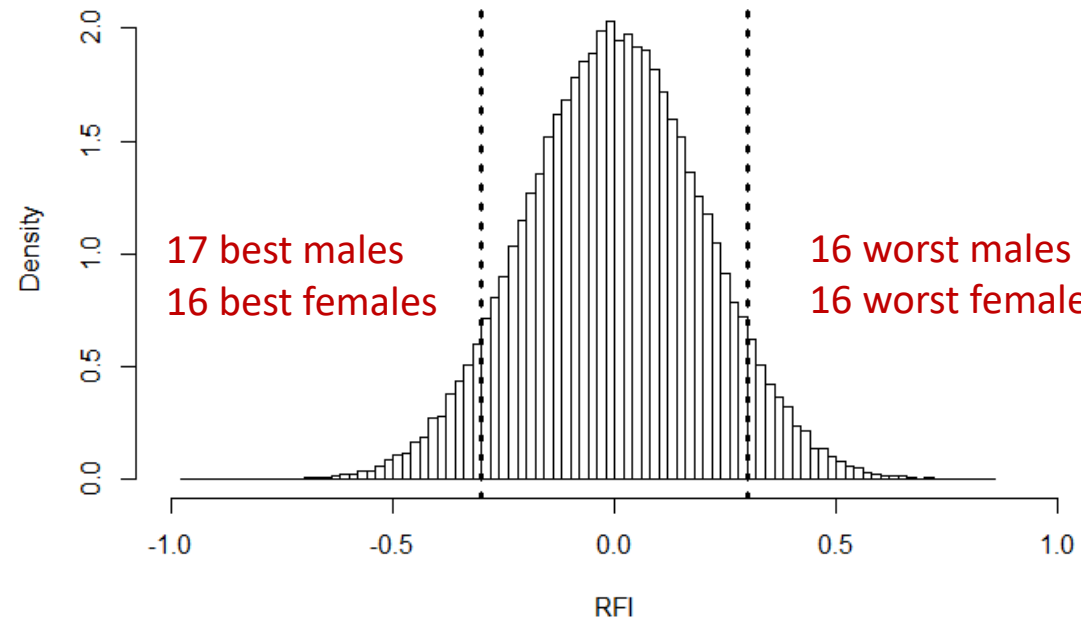


105 Kg of body weight

Low *RFI*



123 males and 121 females

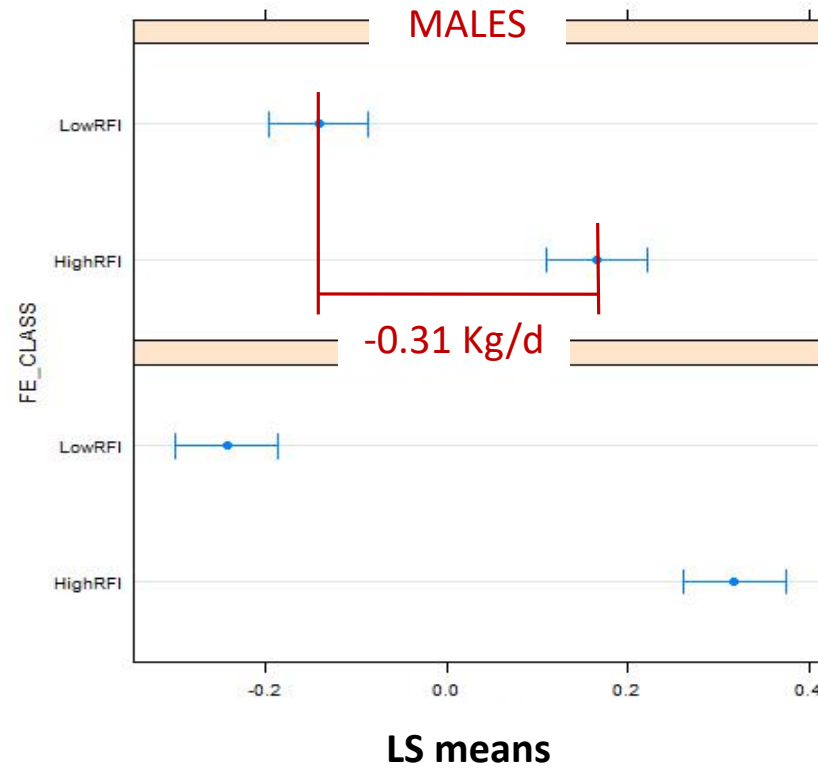


High *RFI*



Differences at the phenotypic level

Low *RFI*

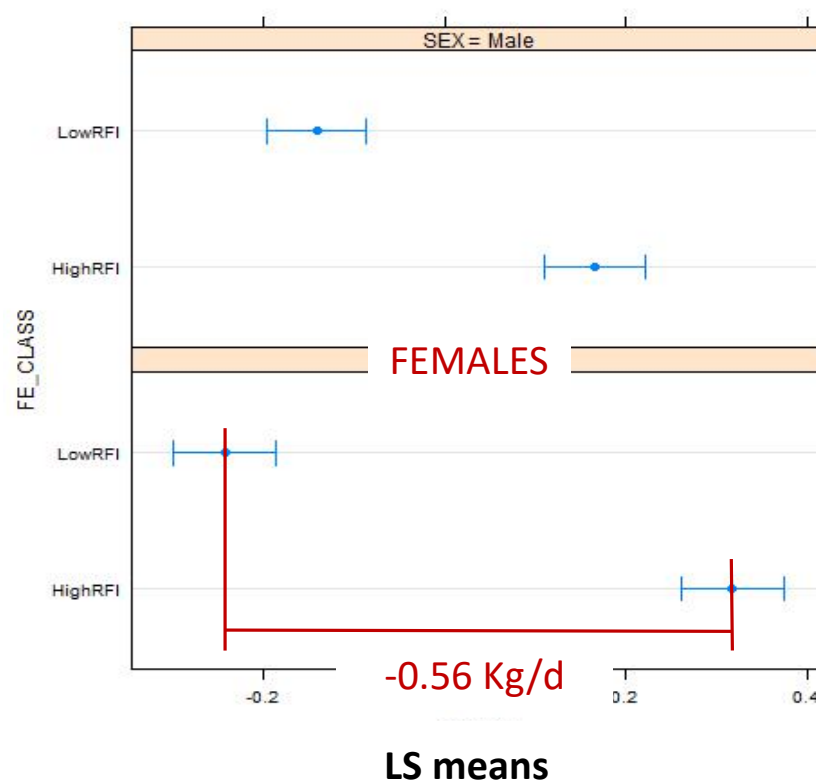


High *RFI*



Differences at the phenotypic level

Low *RFI*



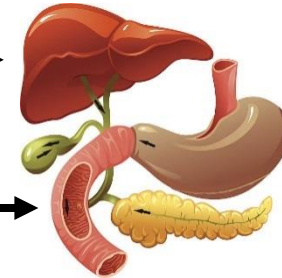
High *RFI*



65 animals



65 samples from liver



65 samples from duodenum



RNA-Seq with an Illumina HiSeq2000

Differential expression analyses: Machine learning algorithms

Functional categorization of DE genes: IPA

Differential expression analysis



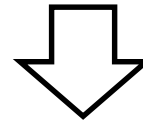
1st step: Gene ranking based on permutation accuracy importance score

Classification using Random Forest algorithm based on conditional inference

Data (X , y)

X is the **65 x 13990 predictor matrix of RNA-Seq data**

y is the **class vector: Low or High RFI**



2nd step: Classification using Machine learning algorithms

Data (X , y)

X is the **65 x p predictor matrix of RNA-Seq data**

Subsets of p predictors $p = 50, 75, 100, 125, 150, 200, 250, 300, 350, 400$

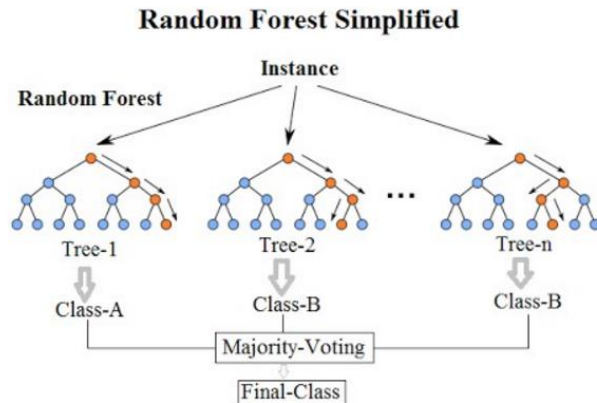
y is the **class vector: Low or High RFI**

Machine learning algorithms

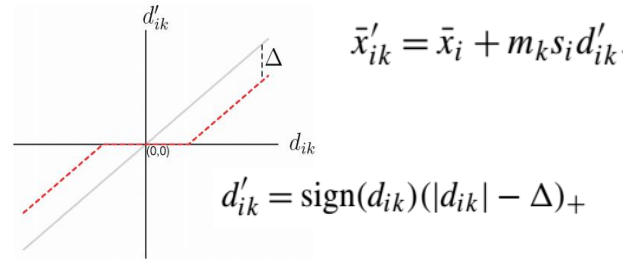


mlr R package

Random Forest (randomForest)



Nearest Shrunken Centroids (PAMR)

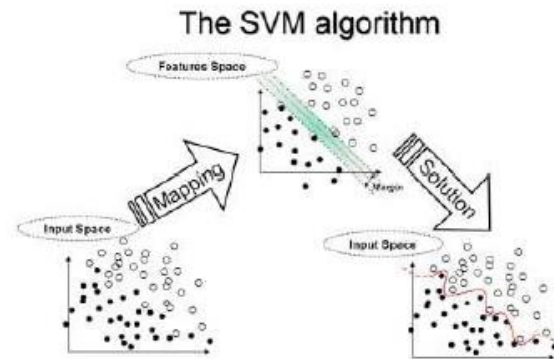


For a test sample $x^* = (x_1^*, x_2^*, \dots, x_p^*)$

$$\delta_k(x^*) = \sum_{i=1}^p \frac{(x_i^* - \bar{x}'_{ik})^2}{s_i^2} - 2 \log \pi_k$$

$$C(x^*) = \ell \text{ if } \delta_\ell(x^*) = \min_k \delta_k(x^*)$$

Support Vector Machine (e1071)



$$\text{maximize } M \text{ subject to } \sum_{j=1}^p \beta_j^2 = 1,$$

$$y_i \left(\beta_0 + \sum_{i \in S} \hat{\alpha}_i \langle x, x_i \rangle \right) \geq M(1 - \varepsilon_i)$$

$$\sum_{i=1}^n \varepsilon_i \leq C \quad \varepsilon_i \geq 0$$

Elastic net (CVGLMNET)

$$\hat{\beta} = \arg \min_{\beta} \|y - \mathbf{X}\beta\|^2 + \lambda_2 \|\beta\|^2 + \lambda_1 \|\beta\|_1$$

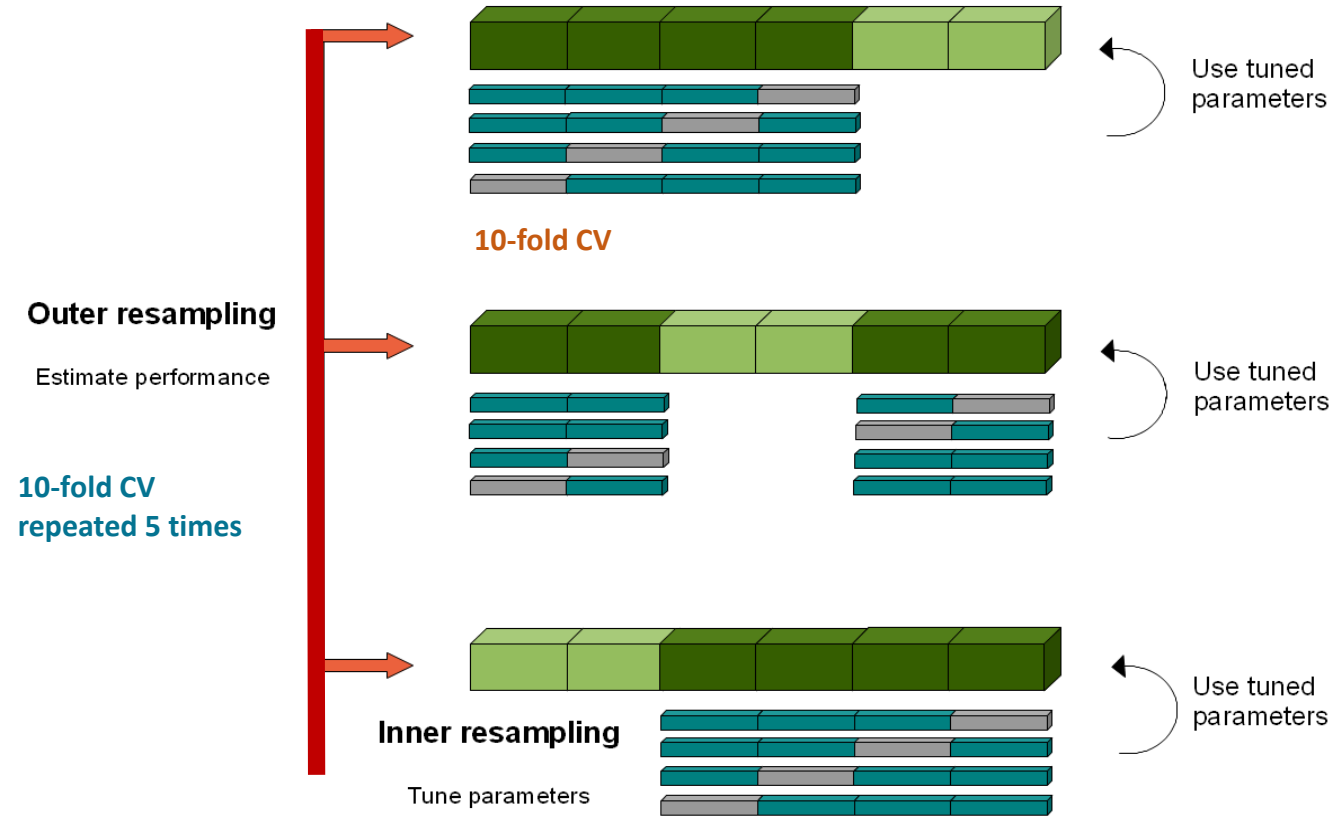
The elastic net penalty

$$J(\beta) = \alpha \|\beta\|^2 + (1 - \alpha) \|\beta\|_1$$

(with $\alpha = \frac{\lambda_2}{\lambda_2 + \lambda_1}$)

$$\min_{\beta} \|y - \mathbf{X}\beta\|^2 \text{ s.t. } J(\beta) \leq t.$$

Nested Resampling



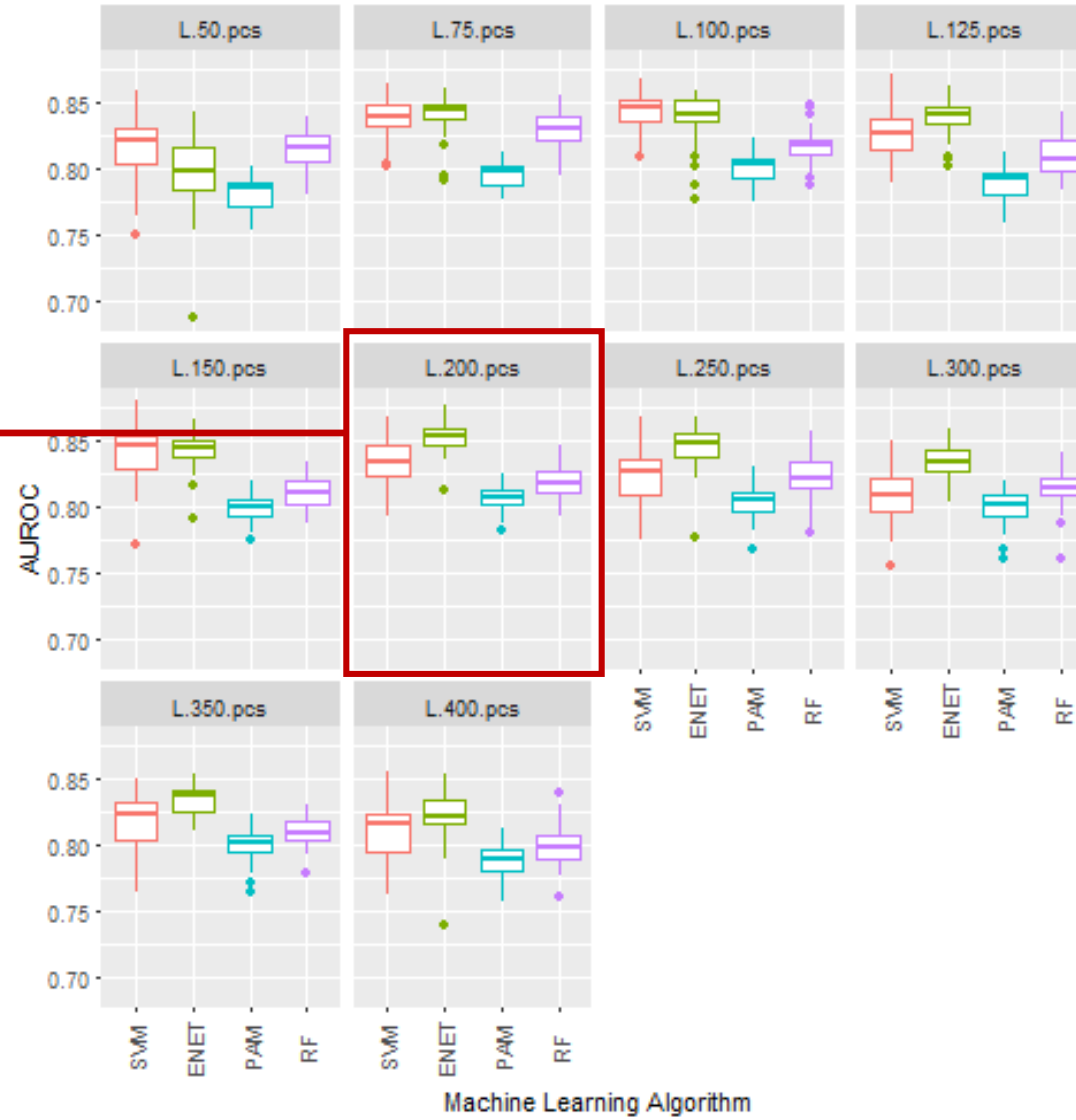
Legend for the diagram components:

- Training set outer resampling
- Test set outer resampling
- Training set inner resampling
- Test set inner resampling

Classification Performance

LIVER
AUROC

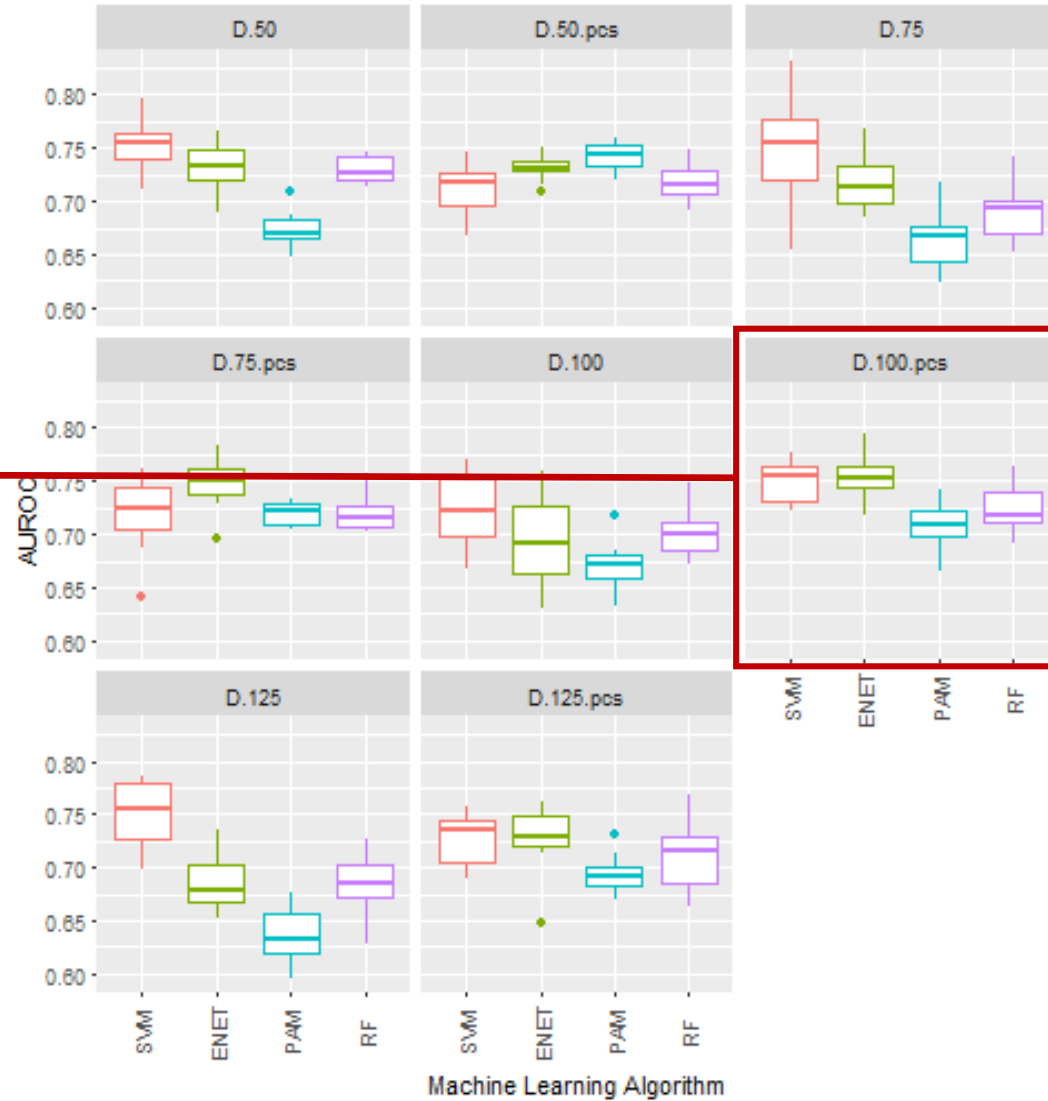
AUROC = 0.85
Accuracy = 0.78



Classification Performance

DUODENUM AUROC

AUROC = 0.76
Accuracy = 0.69



Functional analysis: most significant overrepresented canonical pathways in liver

Canonical Pathways	$-\log(\text{p-value})$	Molecules
Melatonin Degradation II	3,41E00	SMOX,IL4I1
Sirtuin Signaling Pathway	2,01E00	WRN,TUBA1A,GLS,TOMM20,CLOCK,TUBA3E,PFKM
PPAR α /RXR α Activation	1,8E00	PRKAR2B,CLOCK,GNA14,CYP2C8,AIP
Tryptophan Degradation X (Mammalian, via Tryptamine)	1,76E00	SMOX,IL4I1
GPCR-Mediated Nutrient Sensing in Enteroendocrine Cells	1,2E00	PRKAR2B,GNA14,CCK
Xenobiotic Metabolism Signaling	6,81E-01	SMOX,IL4I1,CYP2C8,AIP

Functional analysis: most significant overrepresented canonical pathways in duodenum

Canonical Pathways	$-\log(p\text{-value})$	Molecules
NRF2-mediated Oxidative Stress Response	3,04E00	DNAJC6,MAPK8,DNAJC1,PRKD3
Role of Pattern Recognition Receptors in Recognition of Bacteria and Viruses	2,45E00	MAPK8,CASP1,PRKD3
Aldosterone Signaling in Epithelial Cells	2,2E00	DNAJC6,DNAJC1,PRKD3
Cholecystokinin/Gastrin-mediated Signaling	1,67E00	MAPK8,PRKD3
IL-8 Signaling	1,14E00	MAPK8,PRKD3
Production of Nitric Oxide and Reactive Oxygen Species in Macrophages	1,15E00	MAPK8,PRKD3
Unfolded protein response	9,35E-01	MAPK8
Protein Ubiquitination Pathway	9,25E-01	DNAJC6,DNAJC1

Conclusions

- Good performance of ML algorithms and RNA-Seq expression data for classifying pigs into high or low RFI groups.
- Expression difference of genes involved in feed efficiency are clearer in liver than in duodenum tissues.
 - Among biological pathways identified in liver and duodenum tissues, those related to response to oxidative stress, aldosterone signaling and melatonin degradation seemed consistently involved in FE related traits.

Funding

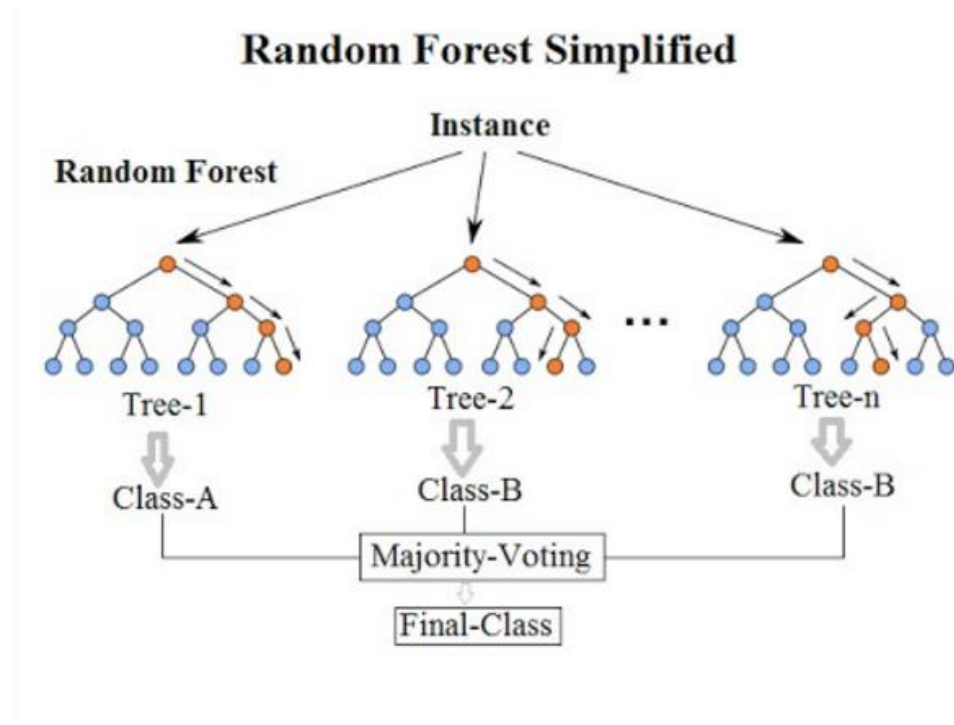
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Machine learning algorithms



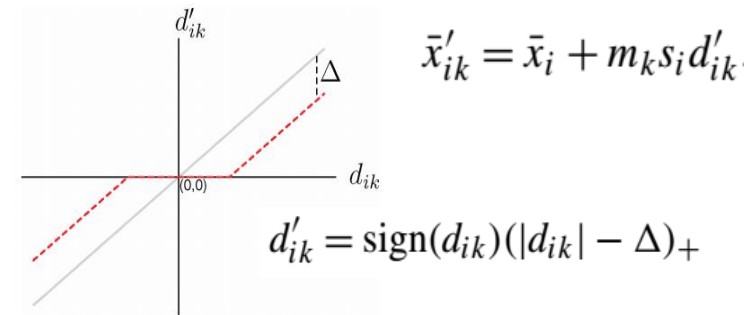
Random Forest



Machine learning algorithms



Nearest Shrunken Centroids (PAMR)



For a test sample $x^* = (x_1^*, x_2^*, \dots, x_p^*)$

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Machine learning algorithms



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The elastic net penalty

$$J(\beta) = \alpha \|\beta\|^2 + (1 - \alpha) \|\beta\|_1$$

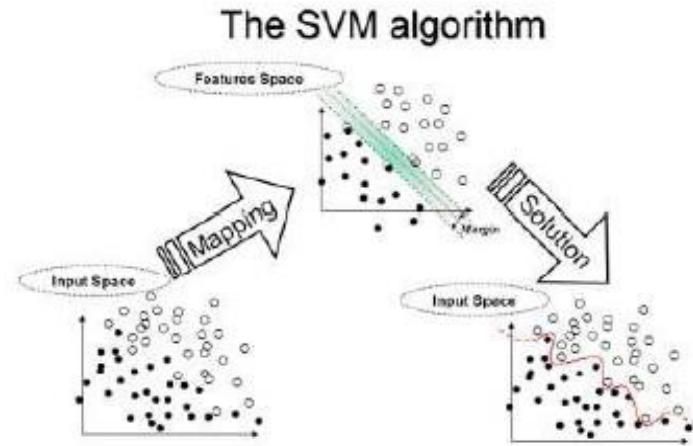
$$\text{(with } \alpha = \frac{\lambda_2}{\lambda_2 + \lambda_1} \text{)}$$

$$\min_{\beta} \|\mathbf{y} - \mathbf{X}\beta\|^2 \text{ s.t. } J(\beta) \leq t.$$

Machine learning algorithms



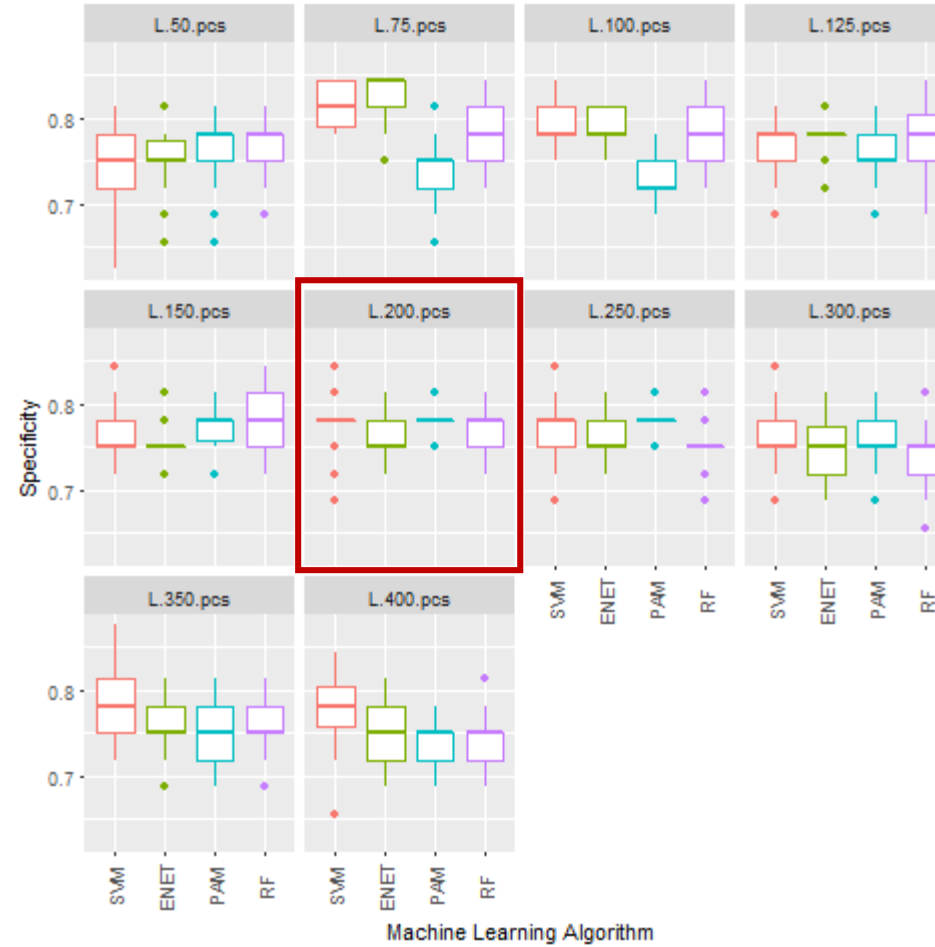
Support Vector Machine (SVM)



$$\begin{aligned} & \text{maximize}_{\beta_0, \beta_1, \dots, \beta_p, \epsilon_1, \dots, \epsilon_n} M \quad \text{subject to} \quad \sum_{j=1}^p \beta_j^2 = 1, \\ & y_i \left(\beta_0 + \sum_{i \in S} \hat{\alpha}_i \langle x, x_i \rangle \right) \geq M(1 - \epsilon_i) \\ & \sum_{i=1}^n \epsilon_i \leq C \quad \epsilon_i \geq 0 \end{aligned}$$

Classification Performance

LIVER Specificity



Classification Performance

DUODENUM Specificity

