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IN VIVO ASSESSMENT OF THE ANTHELMINTIC EFFECTS OF BY-PRODUCTS (PEELS) FROM THE CHESTNUT INDUSTRY.

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EAAP – Ghent 2019



GASTROINTESTINAL NEMATODES (GINs) of SMALL RUMINANTS

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Haemonchus



Teladorsagia



Trichostrongylus

- Grazing = GINs
- Ubiquitous
- Main pathological consequences
 - Production losses
 - Clinical signs and possible deaths
- According to the FAO, the main parasitic infections in Ruminants, worldwide.



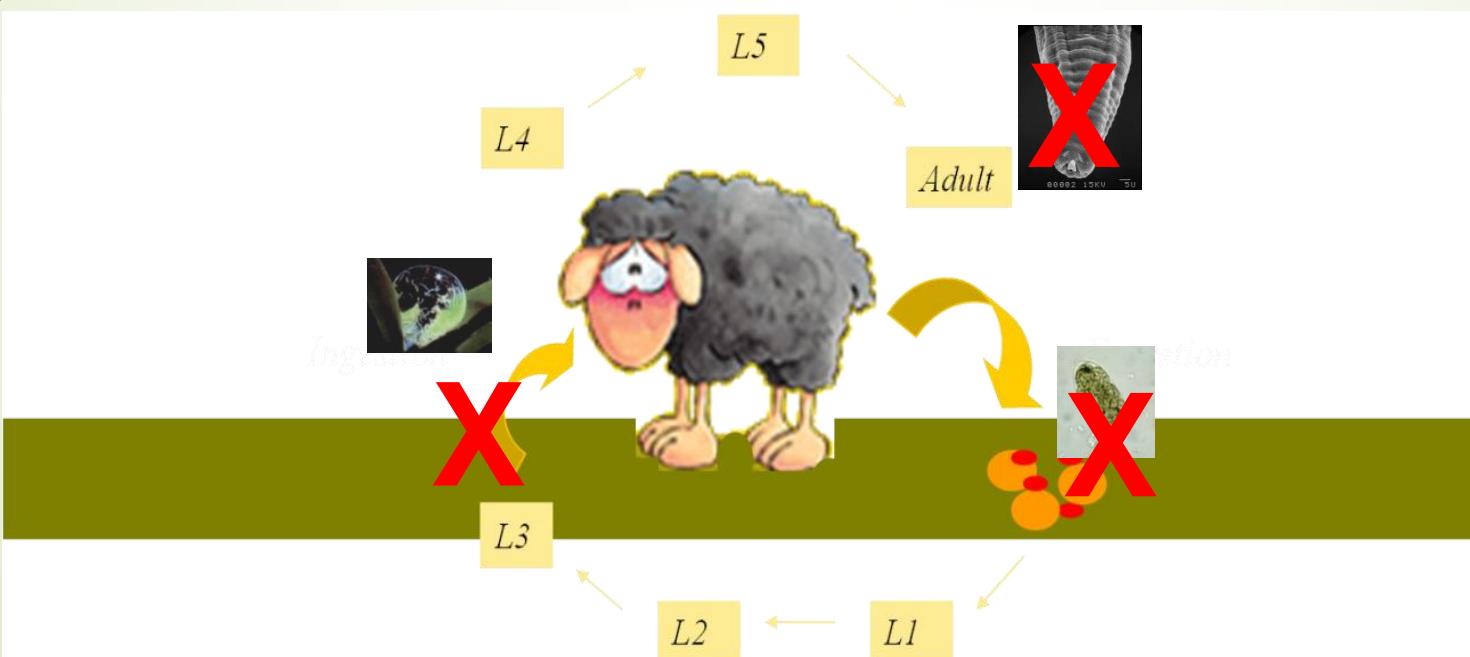
CONTROL of GINs = SYNTHETIC ANTHELMINTIC DRUGS

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ANTHELMINTIC (AHs) TREATMENTS



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INCREASED LIMITS on the USE of AHs and NEED for ALTERNATIVE SOLUTIONS of CONTROL.

Limits of AHs

- ▶ Worldwide development and diffusion of AH resistance in GINs
 - ▶ Increasing number of multiresistant isolates in small ruminants
 - ▶ **Exclusive reliance on AH is not a sustainable option**
-
- ▶ Increased expectations to reduce chemicals in agriculture (e.g OF products)
 - ▶ Increased constraints in the regulation to use commercial AHs

Seek for alternative solutions

- ▶ Biological control, vaccine, genetic selection
- ▶ interaction host nutrition
- ▶ **Natural bioactive compounds of plants**

Sainfoin and other tannin rich Legumes as nutraceuticals

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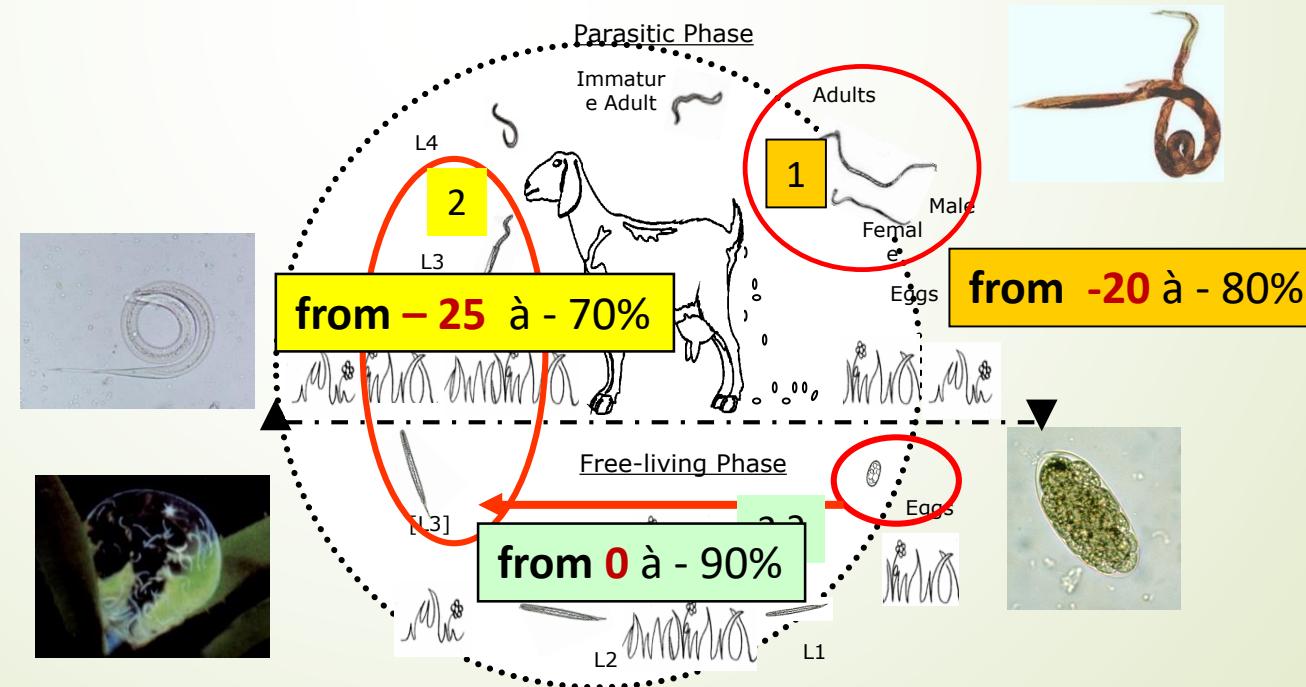
NUTRACEUTICALS in VETERINARY SCIENCES

A feed (a forage) which combined effects on animal **nutrition + health** (Andlauer & Furst, 2002).

It has to be consumed at a certain level of concentration for several days

A nutraceutical is proposed but not imposed to the animals. The effects depend on the animal
Voluntary Feed Intake (VFI)

The **bioactivity** is related to plant secondary metabolites = **PSMs** (e.g. Tannins)



AGRO INDUSTRIAL BY-PRODUCTS : *in vitro* and *in vivo* Results

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- Extracts of tannin rich barks of different trees
 - Quebracho (*Athanasiadou et al, 2000, 2001, Paolini et al 2003 a,b*)
 - Acacia sp (*Cenci et al, 2007, Minho et al 2008; Max et al, 2010, Zabre et al, 2018*)
- By products of carob industry
(*Silanikove et 2006, Manolaraki et al, 2010, Sotiraki et al, 2013*)
- By products of Nuts : Hazel nut peels (*Desrues et al, 2012*)
Use of Chest nuts « skins » ?



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in vitro RESULTS

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ID	Plant	Type	Tannin Content (TT)	Phenol Content (TP)	EC 50
1	Skin of Chestnut	BIO	5.13%	5.71%	67.4
2	Skin of Chestnut	BIO	2.46 %	2.80%	63.2
3	Skin of Chestnut	Conventional	8.12%	8.96%	82.19
4	Skin of Chestnut	Conventional	4.27%	4.81%	75
5	Skin of Chestnut	OF	4.60%	5.05 %	30.6
6	Skin of Chestnut	OF	3.99%	4.57%	75
7	Skin of Chestnut	Conventional	4.43%	4.91%	95.5
8	Skin of Chestnut	Conventional	6.78%	7.53%	80
9	Bugs of Chestnut	Conventional	1.70%	1.84%	140
10	Chips of Chestnut	France	5.19%	5.65%	90.2
11	Chips of Chestnut	Chile	2.50%	2.68%	> 150
12	Leaves of Chestnut	Conventional	NA	NA	124
13	Skin of Chestnut	Spain	6.13%	6.54%	90, 6
14	Skin of Chestnut	Spain	5.11%	5.45%	70

Sainfoin (LEIA method): Mean EC50 = 364 µg/ml (from 84 µg/ml to 630 µg/ml)

OBJECTIVES = To confirm these preliminary data in *in vivo* conditions

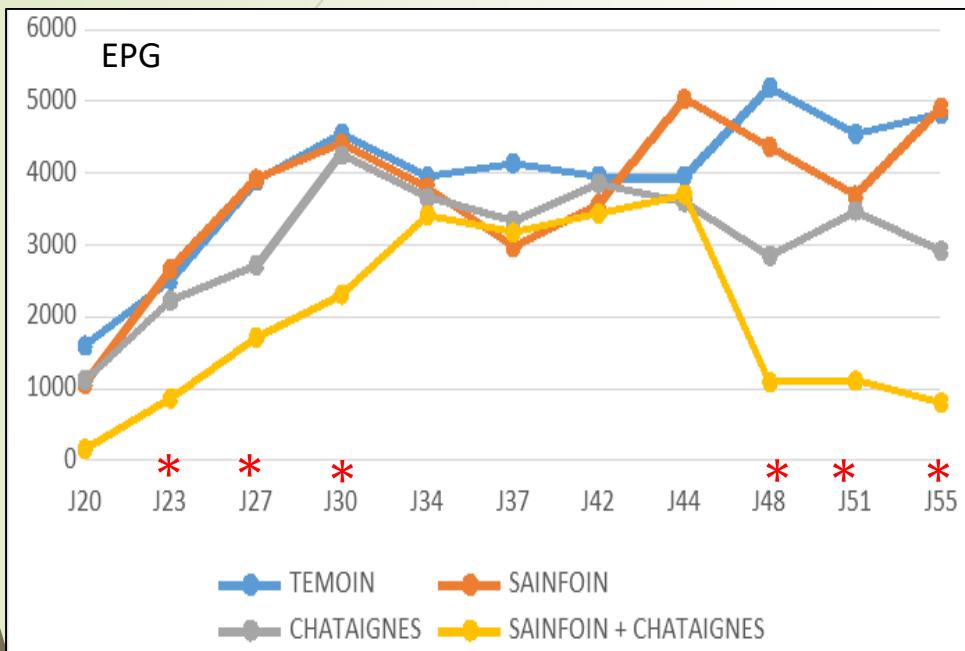
In vivo STUDY = MATERIALS and METHODS

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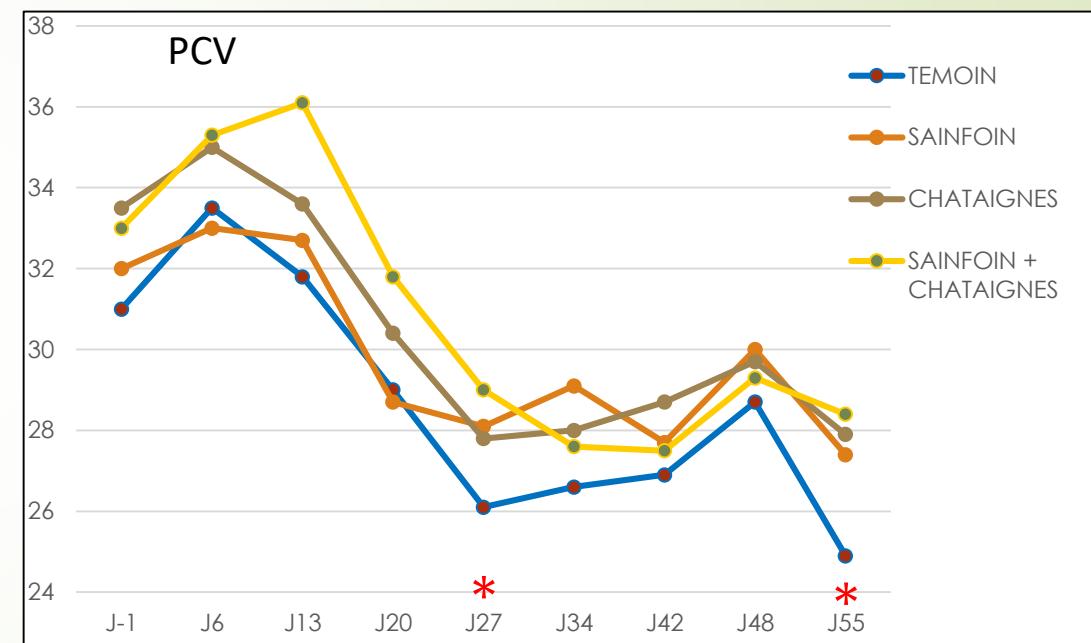
- ▶ 4 groups of 7 experimentally Infected lambs (on D0)
5000 L3 *Haemonchus contortus* + 2300 L3 *T colubriformis*
- ▶ Distribution of diets starting on D-5. Necropsies on D56
- ▶ 4 isoproteic pellets
Control / Sainfoin / Chestnut « skins » / Sainfoin + Chestnut « skins »
- ▶ Measurements of Total Phénols (TP) and Total Tannins (TT)
- ▶ Analyses of the structure of tannins



Fecal Egg Counts



Packed Cell Volume



In vivo RESULTS

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	Total Phenols	Total Tannins	<i>Haemonchus contortus</i>	<i>Trichostrongylus colubriformis</i>
Control	0	0	1716	372
Sainfoin	2.2	2.08	2155	340
Chestnut	4.1	3.9	2300	537
Sainfoin + Chestnut	1.6	1.4	864 *	432
			P < 0,05	NS

Mean establishment rate for *Haemonchus* larvae = 35 %; for *T colubriformis* = 18,3 %

in vivo Results = larval cultures

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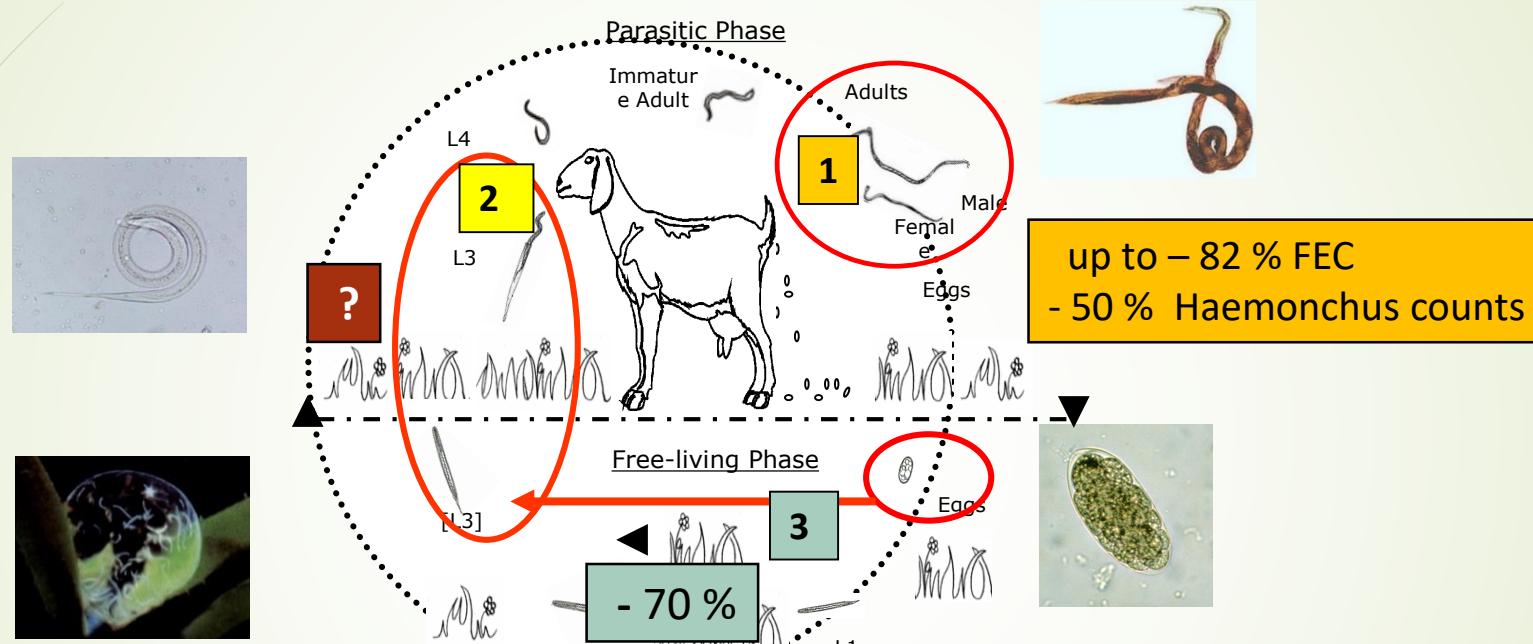
- Ratio *Haemonchus* / *Trichostrongylus* = 70- 80 % vs 30 -20 %
Difference NS entre les groupes
- Yield of group larval culture

	J27	J34	J41	J48	mean
Group C	15	10,4	10	11	11,6 %
Group S	16	20,6	16	15	16,9 %
Group Ch	8	9,7	6	8	7,9 %
Group S+Ch	4	1,2	6	3	3,6 %

Discussion / Conclusions

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- Parasites populations : No effects of Sainfoin and chestnut pellets
Statistical effects with Sainfoin + Chestnut pellets



- Resilience (PCV) some effects on PCV for tannin containing pellets vs controls
- Total tannins: $S + C = 1,4$; $S = 2,1$; $C = 3,9$
- Possible interactions between tannins ???

Thank you for your attention ! Any Questions ?



THANK YOU FOR YOUR ATTENTION



MODE of ACTION against the different STAGES of GINs ?

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- The main hypothesis on the mode of action is a **direct (pharmacological-like) effect**



FUNCTIONAL CONSEQUENCES

1) Infective larvae (L3)

Reduced establishment

2) Adult worms

Motility

Nutrition and Reproduction

Viability

MODE of ACTION against the GINs ?

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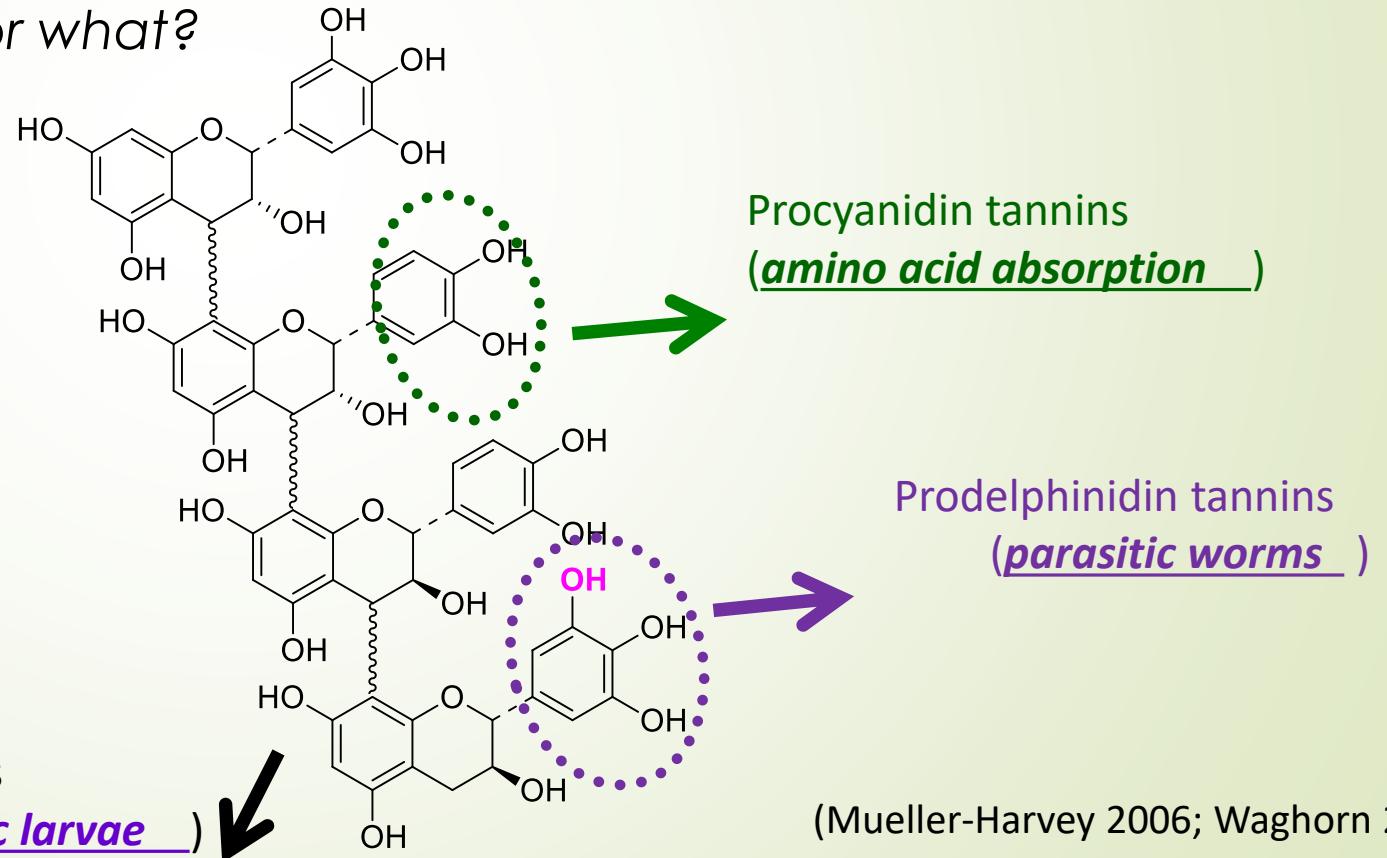
1 CT and also some flavonoids are responsible for the AH properties

(Molan et al, 2003, (Barrau et al 2006, Brunet et Hoste, 2006, Novolbisky et al, 2013, Quijada et al, 2015)

2 *in vitro* and *in vivo* results : Dose dependent activity – Threshold

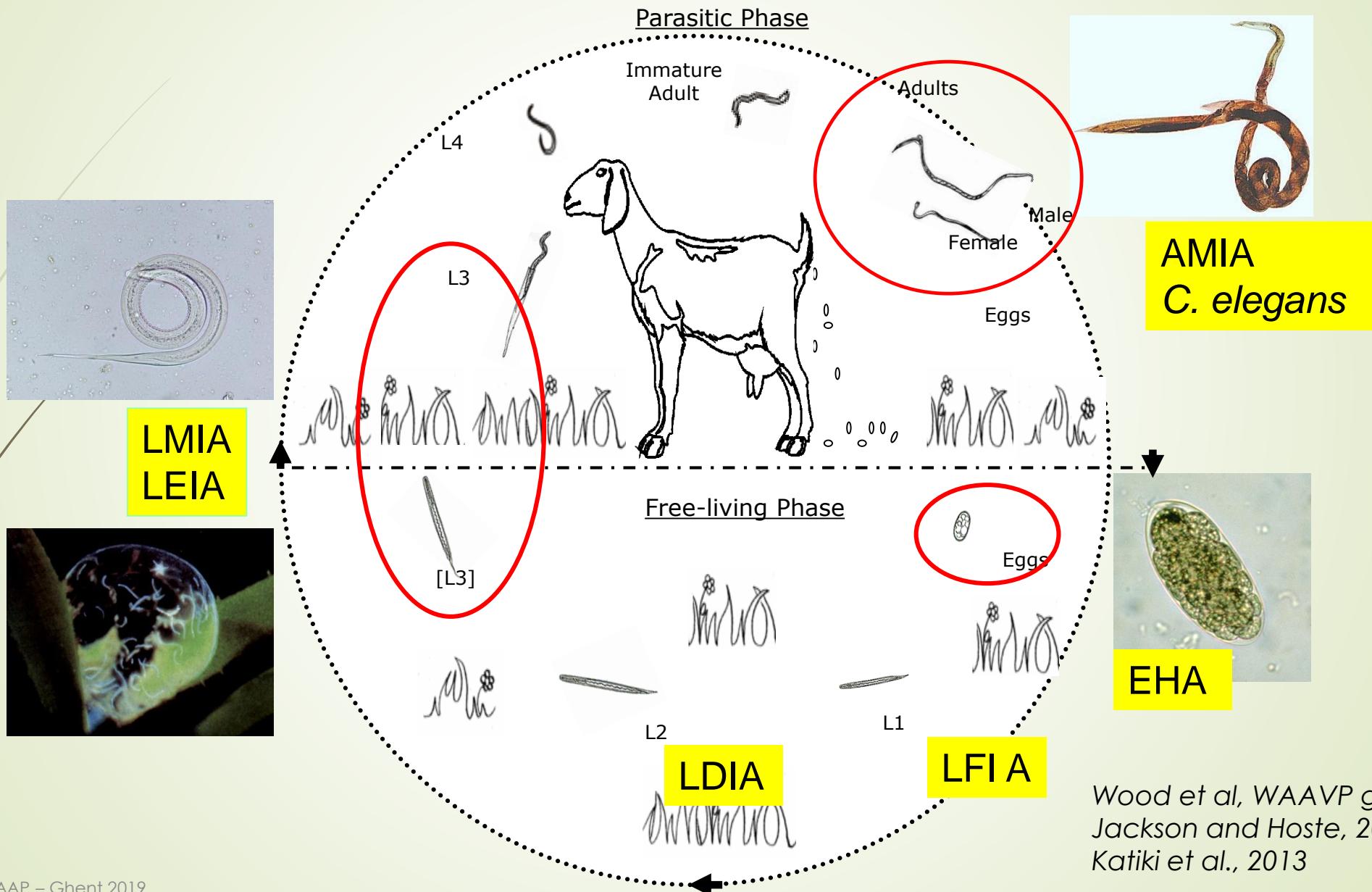
(Terrill et al, 2007, Brunet el, 2007)

3 Which tannins are best for what?



IN VITRO METHODS TO SCREEN AH PROPERTIES

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Other RESSOURCES ? Other LIVESTOCK SPECIES ? Other PARASITES ?

1. Cultivated Legume forages

- Sulla (*Hedysarum coronarium*),
- Sainfoin (*Onobrychis viciifoliae*)
- Sericea lespedeza (*Lespedeza cuneata*)



2. Browsing tropical Legumes



3 Exploiting agro industrial by-products (Caroob, nuts, cocoa..)

4 GIN in cattle and pigs (Williams et al, 2014; Desrues et al, 2015)



5 Coccidia in small ruminants and rabbits