



Walloon Agricultural Research Centre

DEVELOPMENT AND EVALUATION OF REAL-TIME PCR TARGETS FOR THE AUTHENTIFICATION OF INSECT FLOURS

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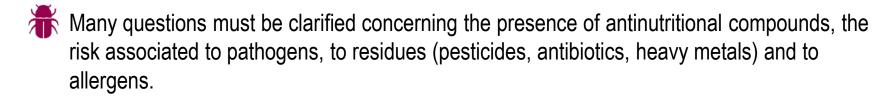




CONTEXT

- In Europe, the processed animal proteins obtained from <u>seven insect species</u> have been authorised <u>for aquaculture</u> by Commission Regulation (EU) 2017/893 since 1 July 2017.
 - Yellow Mealworm (*Tenebrio molitor*)
 - Black Soldier Fly (Hermetia illucens)
 - Common Housefly (Musca domestica)
 - Lesser Mealworm (Alphitobius diaperinus)
 - House Cricket (Acheta domesticus)
 - Banded Cricket (Gryllodes sigillatus)
 - Field Cricket (*Gryllus assimilis*)
- The stakeholders are now hoping future modifications of the legislation to allow the feeding of poultry and pig.





Methods to detect if a product really contains insects and to authenticate insect products will also be mandatory.

European Commission Regulation No 51/2013 added the Polymerase Chain Reaction as a reference method to determine the constituents of animal origin in feed.

To fulfill the need in authentication methods, we started to develop real-time PCR methods specific to the insects listed above

Most of the work here was done in the framework of the FARMYNG project











FlAgship demonstration of industrial scale production of nutrient Resources from Mealworms to develop a bioeconomY New Generation

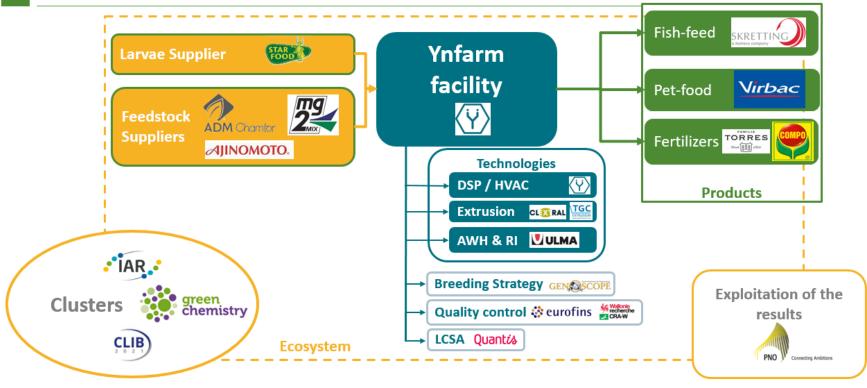
EU project gathering 20 partners





FARMYNG: a full value chain consortium

An integrated biorefinery valorising existing side streams from crops biorefineries into premium products





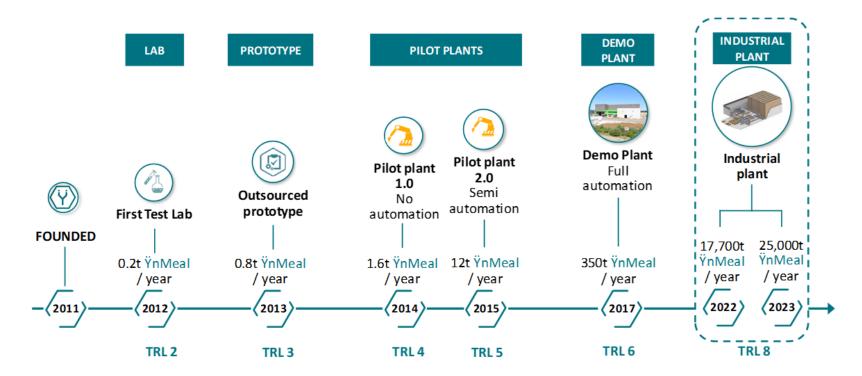






YNSECT in FARMYNG

An innovative and emerging agro-industrial leader dedicated to premium insect-based resources









FARMYNG: Tenebrio molitor insect

Tenebrio molitor is the most important insect species farmed in the feed application sector



- √ High protein content: 72%
- √ significant content of fat : 20%
- \checkmark a very low proportion of ashes.









recherche is active in the Work Package 6 in collaboration with Curofins



WP6 is based on Quality, Safety and Authenticity of insects based-products

This WP includes real-time PCR and High Troughput Sequencing (HTS) methods for the authentication of insect derived products





Step 1: Searching for sequences of interest, design of primers and probes, in silico testing

Insect	Selected gene	Amplicon size (bp)	Reference
All insects	185	81	Debode et al., 2017
Yellow Mealworm (Tenebrio molitor)	wingless and cadherin	87	Debode et al., 2017
Black Soldier Fly (Hermetia illucens)	COX3	67	Marien et al., 2018
Common Housefly (Musca domestica)	white	72	/
Lesser Mealworm (Alphitobius diaperinus)	cadherin	134	/
House Cricket (Acheta domesticus)	cytB	150	/
Field Cricket (Gryllus assimilis)	juvenile hormone esterase	87	/
Silkworm (<i>Bombyx Mori</i>)	cadherin	98	/

Note that some methods are based on low copy targets and other on high copy targets : both are interesting





Step 2 : Evaluation of the performances of the PCR tests

Amplificability

Check that the PCR test is able to amplify the targeted DNA with a good efficiency

Specificity

Test against other insect species (DNA of 52 insect species available in the laboratory) but also mammals, crustacean, fish, birds, mollusca plant species

Applicability

Able to work on real-life samples as processed products or industrial samples



Step 2 : Evaluation of the performances of the PCR tests

Sensitivity

Check that the PCR test is able to detect low amounts of the target

Tests: LOD6 or LOD10, LOD95%

Minimal performance criteria: 20 copies of the target

Robustness

Check that the test is still working with slight deviations to the standard experimental conditions (concentrations of primers and probes, volume of master mix, annealing temperature, different real-time PCR machine, different commercial master mixes)

<u>Acceptance criterion</u>: all deviations to the standard protocol must give a positive result at a level of 20 copies of the target

(Digital PCR)

Better control of copy numbers used in PCR reactions

Check the absence of similar sequences (that could generate a second PCR product)





Step 2 : Evaluation of performance - Results

Insect	Efficiency	Specificity	Applicability	Sensitivity	Robustness
All insects	V	V (due to plant species)	V	/	/
Tenebrio molitor	V	V*	V	LOD6 = 20	/
Hermetia illucens	V	V *	V	LOD6 = 5 LOD95% = 5	
Musca domestica	V	/ (?)	/	/	/
Alphitobius diaperinus	V	V *	V	LOD6 = 5 LOD95% =10	V
Acheta domesticus	V	V *	V	LOD ₁₀ = 5 LOD _{95%} =10	V
Gryllus assimilis	V	V*	V	LOD6 = 10 LOD95% = 20	V
Bombyx Mori	V	V*	V	LOD6 = 10 LOD95% =10	V

(*) late signals for few insect species are sometimes encountered







Real-time PCR methods: targeted method, we only find what we are looking for



Need to develop untergated methods: High Troughput Sequencing (HTS) is a solution

- → Development of approach by metagenomics / barcoding
- → CRA-W is already working on approaches for animals and plants detection
- → Need of bioinformatic skills (available at CRA-W)

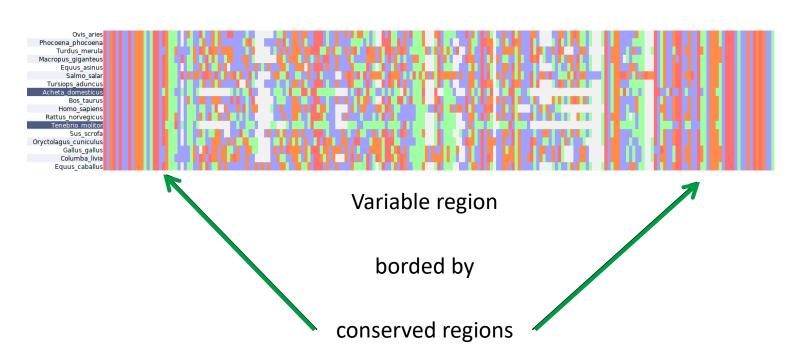




High Troughput Sequencing (HTS)

First tests realised on products containing insects with a 16S DNA region used for detection and distinction of animal species

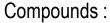
16S DNA





High Troughput Sequencing (HTS)

Pasta with Tenebrio meal (commercial food product)



- Tenebrior molitor (10%)
- Chickpea (43%)
- Brown rice (43%)
- Egg whites (4%)



All animal reads (> 35000) are assigned to *Tenebrio molitor*

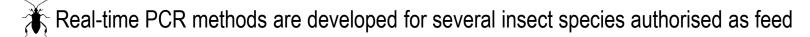


**With the vegetal targets, chickpea and rice were also detected



CONCLUSIONS on DNA based methods

Real-time PCR



Low and high copy number targets have both an interest

General target for insect must be improved

Concerning Musca domestica, many close species are erroneously considered to be Musca domestica

High Troughput Sequencing (HTS)

W Research is only starting

W Promising method

Several targets will be necessary to confirm the results





Thanks for your attention



















