



Faculty of Science



# Insect diseases in production systems: prevention and management

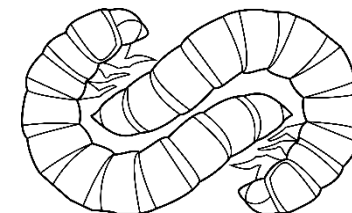
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Department of Plant and Environmental Sciences  
University of Copenhagen, Denmark

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Innovation Fund Denmark



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## Diseases in insects produced for food and feed

J. Eilenberg<sup>1\*</sup>, J.M. Vlask<sup>2</sup>, C. Nielsen-LeRoux<sup>3</sup>, S. Cappellozza<sup>4</sup> and A.B. Jensen<sup>1</sup>

*Journal of Insects as Food and Feed*, 2015; 1(2): 87-102

### Diseases of insects in European production systems: Diagnosis, prevention and management

*Krankheiten von Insekten in Europäischen Produktionssystemen: Diagnose, Prävention und Management*

Antoine Lecocq, Annette Bruun Jensen, Jørgen Eilenberg

Berl Münch Tierärztl Wochenschr  
DOI 10.2376/0005-9366-18061

14

### Prevention and Management of Diseases in Terrestrial Invertebrates

Jørgen Eilenberg and Annette Bruun Jensen

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In Hajek, A.E. & Shapiro-Ilan, D. (2018): Ecology of Invertebrate Diseases, John Wiley and Sons



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## Natural enemies in insect production systems

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In Huis, A. (ed) 2017: Insects as Food and Feed from Production to Consumption. Wageningen University Press, 201-222.

### Towards a coordination of European activities to diagnose and manage insect diseases in production facilities

J. Eilenberg, M.M. van Oers, A.B. Jensen, A. Lecocq, G. Maciel-Vergara, L.P.A. Santacoloma<sup>1</sup>, J.J.A. van Loon and H. Hesketh



*Journal of Insects as Food and Feed*, 2018; 4(3): 157-166

SPECIAL ISSUE: *Insects in European feed and food chains*



Article

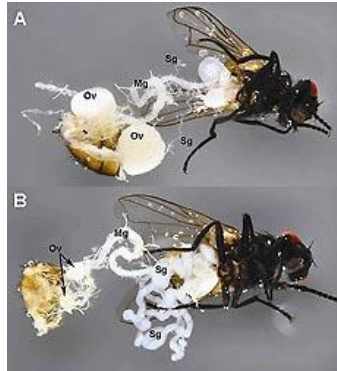
### Cannibalism as a Possible Entry Route for Opportunistic Pathogenic Bacteria to Insect Hosts, Exemplified by *Pseudomonas aeruginosa*, a Pathogen of the Giant Mealworm *Zophobas morio*

Gabriela Maciel-Vergara<sup>1,2,3,\*</sup> , Annette Bruun Jensen<sup>1</sup>  and Jørgen Eilenberg<sup>1</sup>

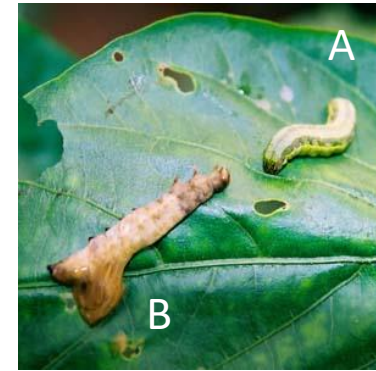
# Virus infections in insects



RNA Virus: Honey  
Bee Deformed  
Wing Virus DWV


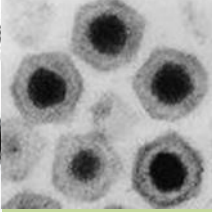
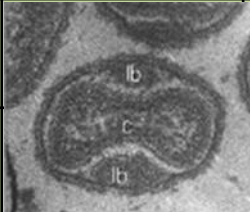
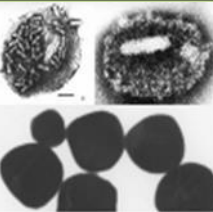
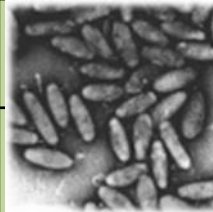


DNA virus: House fly  
*Musca domestica* infected  
(B) with SGHV

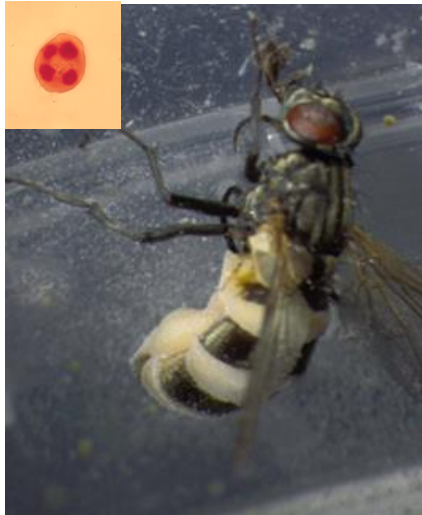


DNA Virus: Caterpillar  
(B) infected with  
Baculovirus

## Examples of large invertebrate DNA viruses

Virus	Asco-virus	Irido-virus	Entomo-poxvirus	Baculo-virus	Nima-virus
Virion					

# Insect pathogenic fungi: specialists/generalists



House fly *Musca domestica*  
infected with  
*Entomophthora schizophorae*

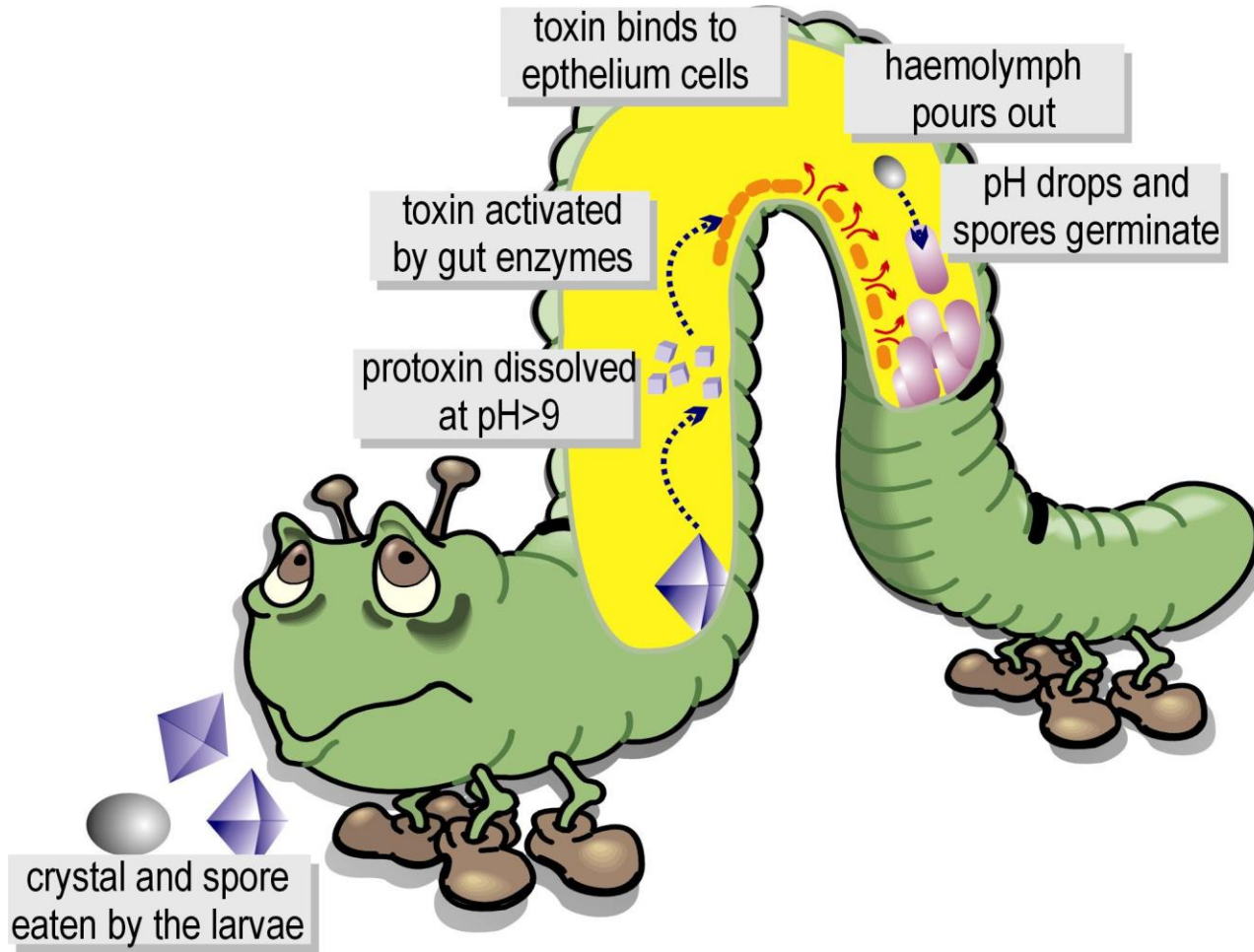


Grasshopper nymph  
*Schistocerca americana*  
infected with  
*Entomophaga grylli*

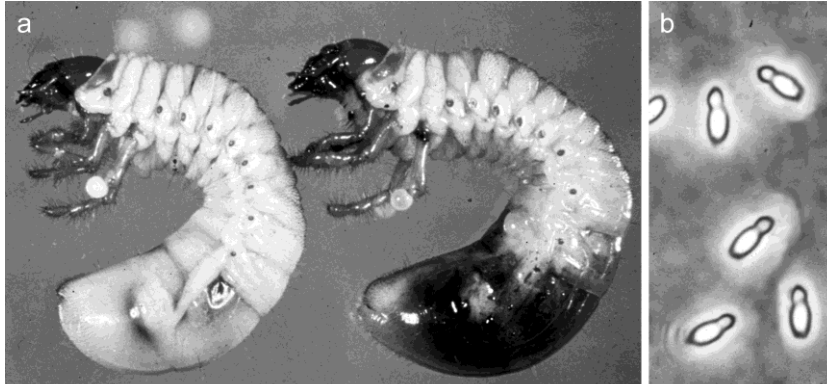


Adult weevil  
*Strophosoma* sp.  
infected with *Beauveria bassiana*

# *Bacillus thuringiensis*



## A specialist insect pathogenic bacterium



*Bacillus popilliae*, 'Milky White Disease', a pathogen on scarabs

## An opportunistic insect pathogenic bacterium

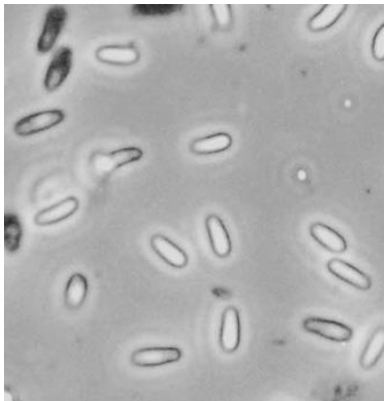


Probably genus *Pseudomonas*, infecting mealworm *Tenebrio molitor*

# Microsporidia

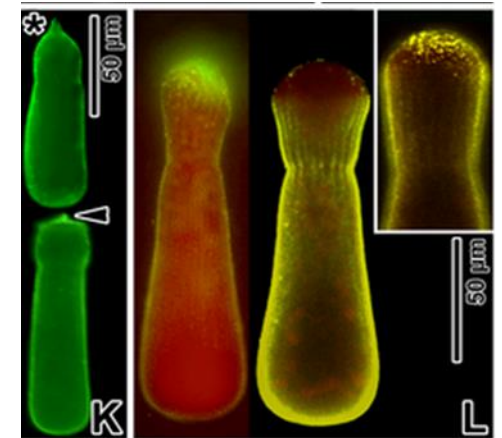
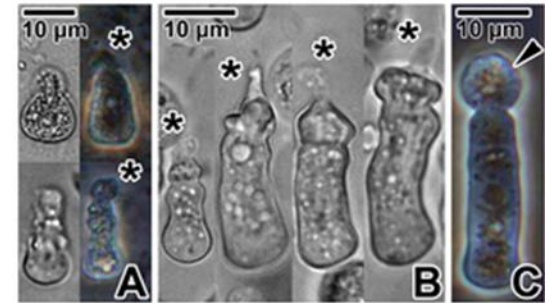


Grasshoppers infected with *Nosema locusta*



Spores of *Cystosporogenes deliaradicae* from infected fly *Delia radicum*

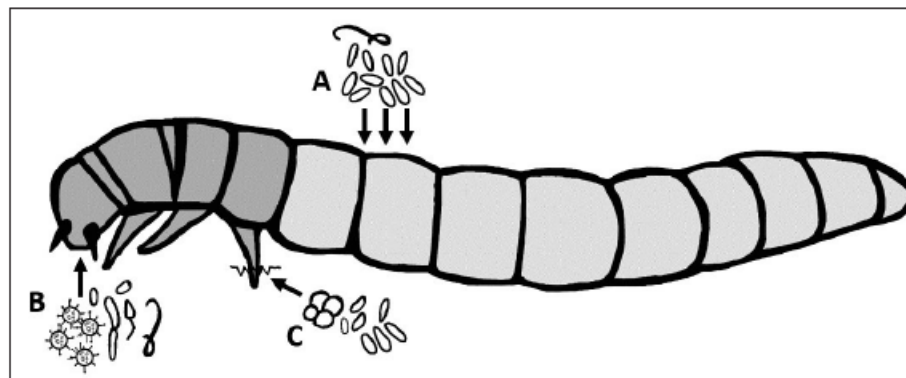
# Neogregarines



*Gregarina cuneata* in infected Mealworm *Tenebrio molitor*

# Some characteristics of important groups of insect pathogens

Type	Infection route	Specialists or generalists
Virus	Oral	Specialists
Bacteria	Oral	Generalists/specialists
Fungi	Through cuticle/oral	Generalists/specialists
Microsporidia	Oral	Specialists
Nematodes	Through body openings	Generalists



**FIGURE 1:** Infection routes for insect pathogens. (A) Entomopathogenic fungi and nematodes can enter the insect's body cavity through the cuticle. (B) Pathogens such as viruses and bacteria, but also microsporidia, protozoa and nematodes, infect their host upon ingestion, alongside feed or faeces. (C) Injury to an insect's limb or body, often as a result of aggression among conspecifics, can allow opportunistic pathogens like certain bacteria and fungi, to enter the body and replicate. (source: Antoine Lecocq)



**Table 14.2** Selected results from a survey (2014–16) of insect diseases found by employees in production facilities for insects used as food and feed.

Production insect	Disease	Symptoms	Action
<i>Acheta domesticus</i>	Bacterium sp.	Increased mortality, red appearance	Cleaning of cages
<i>Acheta domesticus</i>	<i>Metarhizium</i> sp. and <i>Beauveria bassiana</i>	Some mortality in population	Quarantine, new breeding stock
<i>Acheta domesticus</i>	Cricket paralysis virus (CrPV)	Collapse of cricket population: the virus seems to spread globally	Switching to new breeding stock or even new cricket species
<i>Gryllus bimaculatus</i>	<i>Gryllus bimaculatus</i> iridovirus	Swollen abdomen, strikingly sluggish, mortality close to 100%	Occurs occasionally
<i>Tenebrio molitor</i>	<i>Beauveria bassiana</i>	Some mortality in population	Cleaning, removal of dead larvae, quarantine
<i>Zophobas morio</i>	<i>Pseudomonas</i> sp.	Increased mortality: recurrent problem	Removal of dead larvae
<i>Musca domestica</i>	<i>Entomophthora muscae</i>	Dead adult flies with spores, epidemic	Cleaning, removal of dead flies, quarantine
<i>Hermetia illucens</i>	Unknown (bacterium?)	Elongated, rounded mature larvae, moving slowly before dying	Quarantine



## A typical situation:

What is wrong with my *Tenebrio molitor* in culture ?

Some get discolored, some die

Is it an insect disease or some saprophytic microorganisms?

Any action needed?



## Diagnosis needed

Bacterium, unknown species



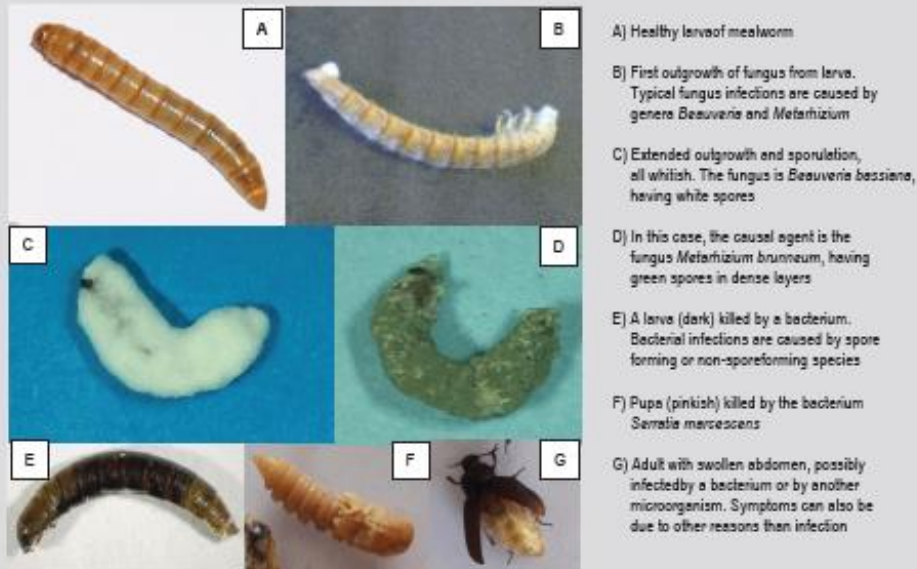
Fungus (*Beauveria* sp.) infected



Nematode (*Steinernema feltiae*)  
infected

Uninfected

## Common insect diseases in mealworm, *Tenebrio molitor*



- A) Healthy larva of mealworm
- B) First outgrowth of fungus from larva. Typical fungus infections are caused by genera *Beauveria* and *Metarhizium*
- C) Extended outgrowth and sporulation, all whitish. The fungus is *Beauveria bassiana*, having white spores
- D) In this case, the causal agent is the fungus *Metarhizium brunneum*, having green spores in dense layers
- E) A larva (dark) killed by a bacterium. Bacterial infections are caused by spore forming or non-sporeforming species
- F) Pupa (pinkish) killed by the bacterium *Serratia marcescens*
- G) Adult with swollen abdomen, possibly infected by a bacterium or by another microorganism. Symptoms can also be due to other reasons than infection

*Tenebrio molitor* can be affected by lethal insect pathogens, both in larval, pupal and adult stages. Commonly found are fungi from the genera *Beauveria* and *Metarhizium*, and bacteria (genus *Serratia* and others). To diagnose fungal infections, allow dead larvae with early symptoms (B) to stay in humid chamber (moist filter paper added) up to two weeks. If infected by the *Beauveria* or *Metarhizium*, extensive outgrowth will be observed (C and D). First diagnosis is based on color of fungus and other visible symptoms, later to be accompanied by spore morphology to determine genus name. If species name is needed, PCR can be added. For bacteria, it might be difficult to distinguish between true infections caused by an insect pathogen and growth caused by opportunistic bacteria. Due to few prominent morphological characters like color and swelling of host (E, F and G), often PCR is needed to determine to genus or species level of bacteria. Other microorganisms like insect virus and protozoa can also infect mealworms, although little is known about their effects on host. Fungal growth on dead insects can be due to mold fungi. Malfunctions, discoloration and death of *Tenebrio* may also be due to other reasons than infections.

A main reason for insect pathogens to occur and spread is a lack of cleaning and insufficient removal of dead insects. Also, a stressed population may be more susceptible to opportunistic bacterial infections. Management: Check carefully for dead insects, remove cadavers immediately. Note external symptoms, eventually also symptoms on living individuals. Immediate cleaning using standard washing and sterilizing liquids of all equipment is needed, potentially UV light as well (Eilenberg et al., 2015). Smaller cohorts of insects should be kept in quarantine to avoid further spread. Contact an INSECTPATH laboratory for further assistance and advice.

Professor Jørgen Eilenberg, University of Copenhagen, Denmark. [Jei@plen.ku.dk](mailto:Jei@plen.ku.dk), phone + 45 23 30 20 24.



Reference: Eilenberg et al. 2015, Journal of Insects as Food and Feed, 2: 87-102  
 inVALUABLE deliverable prepared by University of Copenhagen, December 2017.  
 University of Copenhagen: all photos, except F and G (Delphine Calas-List, Entomo Farm)

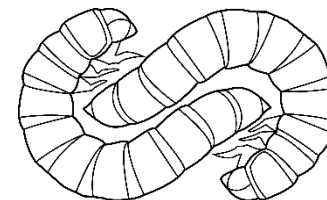
## Fact sheet: *Tenebrio molitor*

For use in production facilities

Freely available [jei@plen.ku.dk](mailto:jei@plen.ku.dk)

Feedback from users is important

We plan to prepare similar sheets for other important insects in production



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## Closed production facilities



A) **Closed production facilities** for lesser mealworm (*Alphitobius diaperinus*). Larvae are reared in plastic boxes on separate shelves. The Netherlands.

B) **Automatic separation** of *Tenebrio molitor* larvae from substrate. The Netherlands.

C) **Closed production facilities** for black soldier fly (*Hermetia illucens*). Larvae are reared in plastic boxes stacked at several meters height. South Africa.

## Semi-open production facilities



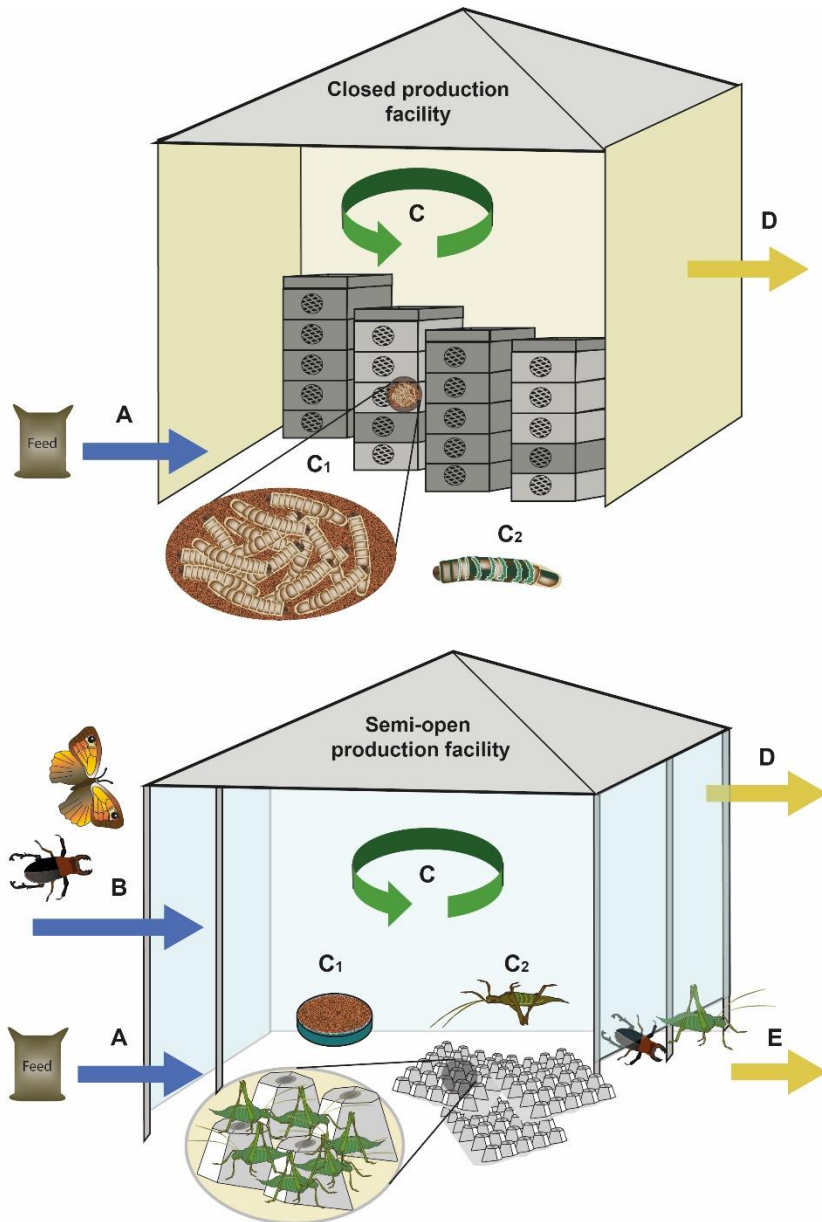
D), E) **Semi-open production facilities** for house cricket (*Acheta domesticus*) and black cricket *Gryllus bimaculatus*. New cohorts are started in the different sections at time intervals. Thailand.

# Outdoor production facilities



- F) **Outdoor hives** for honeybee (*Apis mellifera*) in almond plantation with low biological diversity. California, USA
- G) **Outdoor hives** for *A. mellifera* in an agricultural landscape with medium biological diversity. Denmark
- H) **Outdoor hives** for *A. mellifera* near tropical forest with high biological diversity. Tanzania.
- I) **Outdoor production/foraging** of Mopane Moth *Gonimbrasia belina*, Zimbabwe





Main transportation routes of insect pathogens from the external environment and into facility, movement within facility and transportation from facility to the external environment



# Recommendation

- Check physical structure of production facilities. Do they prevent natural enemies to enter?
- Check substrates to enter the facilities. Any risk of microorganisms?
- Ensure optimal rearing conditions. Batches in separate units or continuous production systems? Consider genetical variation in stock insects, inbreeding
- Frequent observations of production facilities and insects are needed
- Viruses, bacteria, fungi, microsporidia and protozoa all need different methods to be diagnosed , covert infections are tricky to discover
- Contact insect pathology laboratories: [Jej@plen.ku.dk](mailto:Jej@plen.ku.dk)

# INSECT DOCTORS

Educating tomorrow's insect pathologists to solve problems caused by diseases in the insect-rearing industry

An EU supported IJD-ITN project. Fifteen PhD projects to be initiated in spring 2020, covering a range of basic and applied aspects.

Coordinator Prof. Monique van Oers,  
Wageningen University  
[Monique.vanoers@wur.nl](mailto:Monique.vanoers@wur.nl)

## Beneficiaries

(= Universities hiring the students)

- Wageningen University\*
- University of Copenhagen\*
- Centre National de la Recherche Scientifique\*
- University Valencia\*
- National Institute for Agricultural Research
- University of Exeter\*
- UK Centre for Ecology & Hydrology
- Julius Kühn Institute

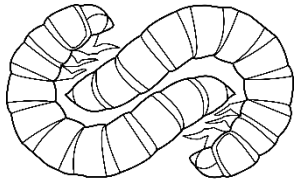
\* Dual Degree Awarding Universities

## Partners

- University of Leeds\*
- Grande École AgroParis Tech\*
- Tours University\*
- Technical University of Darmstadt\*
- Food and Agriculture Organisation (FAO)
- Koppert BV
- Proti-Farm R&D BV
- YNSECT
- Entec Nutrition Ltd.
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