



# Sequence-based genome-wide association study of milk mid-infrared wavenumbers in dairy cattle

**Kathryn Tiplady**

---

T. Lopdell<sup>1</sup>, E. Reynolds<sup>2</sup>, R. Sherlock<sup>1</sup>, M. Keehan<sup>1</sup>, T. Johnson<sup>1</sup>, J. Pryce<sup>3,4</sup>, H. Blair<sup>2</sup>, S. Davis<sup>1</sup>, M. Littlejohn<sup>1,2</sup>, D. Garrick<sup>2</sup>, R. Spelman<sup>1</sup> and B. Harris<sup>1</sup>

<sup>1</sup>Livestock Improvement Corporation, Hamilton, NZ

<sup>2</sup>Massey University, AL Rae Centre, Hamilton, NZ

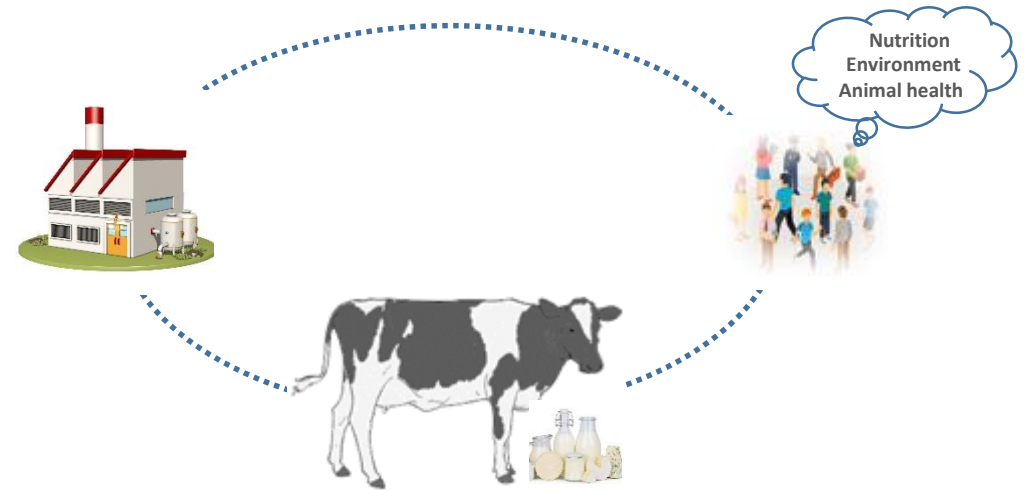
<sup>3</sup>Agriculture Victoria, AgriBio, VIC, Australia

<sup>4</sup>La Trobe University, VIC, Australia



# Introduction

- Milk is highly complex.

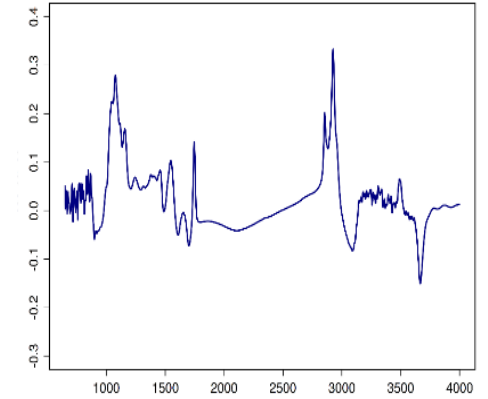
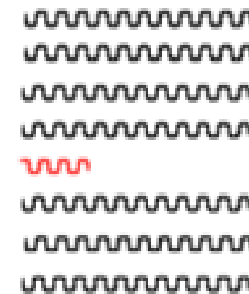
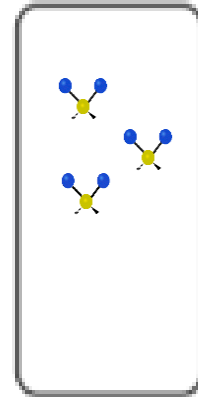
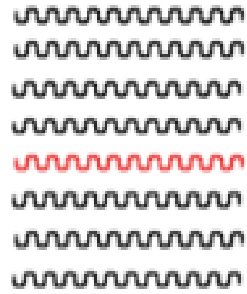


- Fourier-transform infrared (FTIR) spectroscopy is widely used to estimate milk composition and other indicator traits.
- High throughput, inexpensive, readily available from routine milk testing.

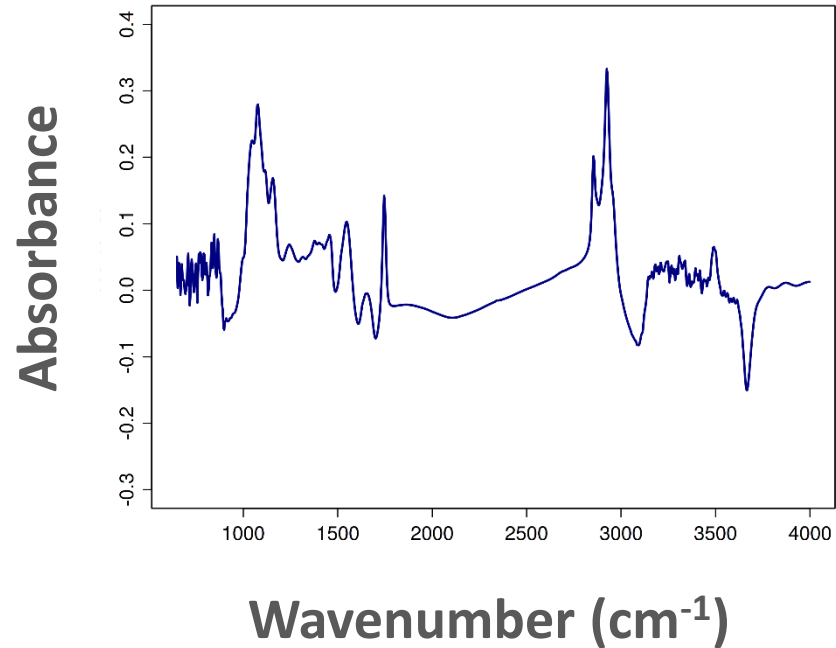
# FTIR spectroscopy



Infrared  
light



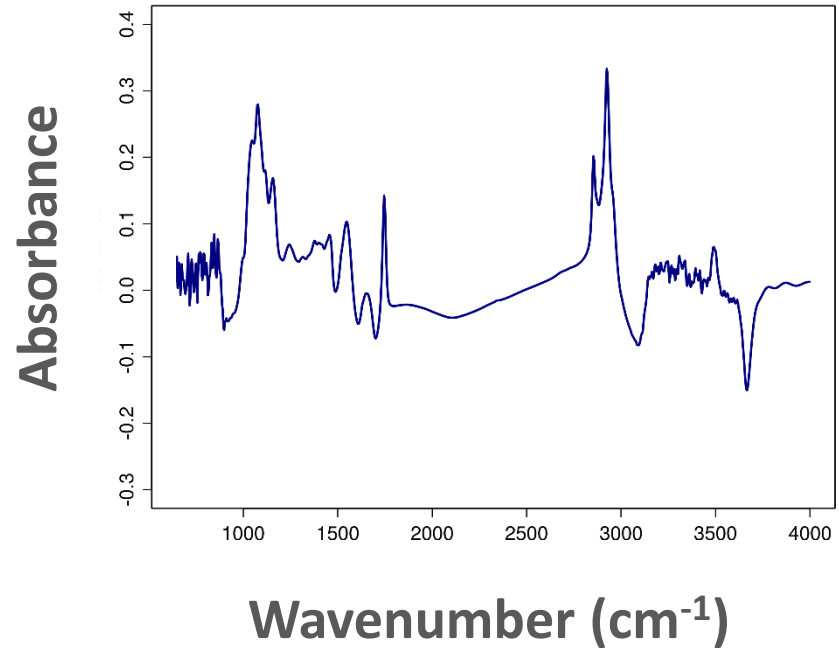
# FTIR spectroscopy



$$Fat = F_0 + F_1w_1 + F_2w_2 + \dots + F_kw_k$$

$$Protein = P_0 + P_1w_1 + P_2w_2 + \dots + P_kw_k$$

# FTIR spectroscopy



Energy balance  
Fertility  
Fatty acid profiles

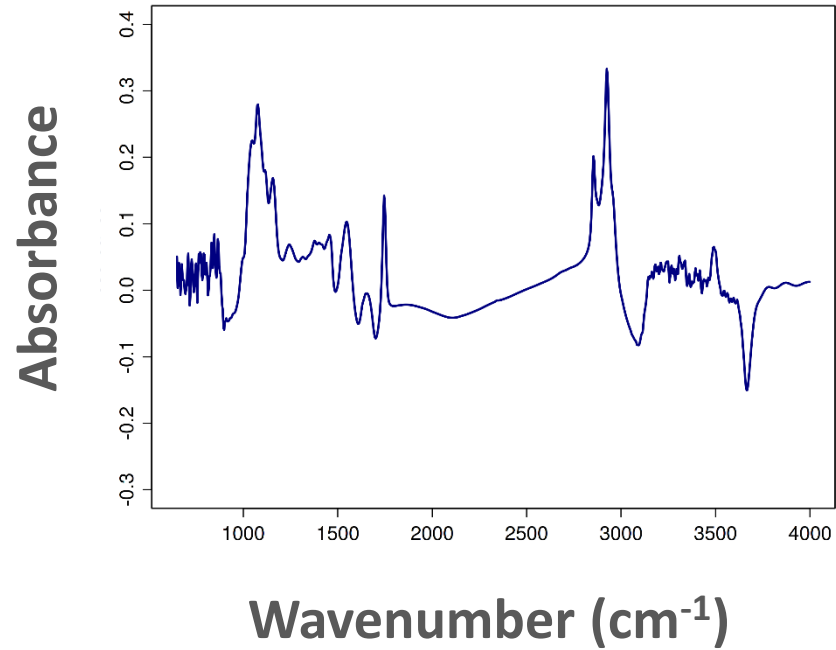


Nitrogen  
Methane

$$Fat = F_0 + F_1w_1 + F_2w_2 + \dots + F_kw_k$$

$$Protein = P_0 + P_1w_1 + P_2w_2 + \dots + P_kw_k$$

# FTIR spectroscopy



$$\text{Fat} = F_0 + F_1w_1 + F_2w_2 + \dots + F_kw_k$$
$$\text{Protein} = P_0 + P_1w_1 + P_2w_2 + \dots + P_kw_k$$



Energy balance  
Fertility  
Fatty acid profiles



Nitrogen  
Methane

# Genetics of FTIR data

---

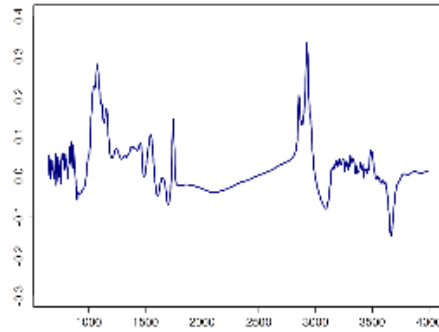


- Individual wavenumbers have moderate to high heritability across the mid-infrared region (Bittante & Cecchinato 2013; Soyeurt et al. 2010).
- Some promising association peaks have been identified in other studies (Wang & Bovenhuis 2018; Bendet et al. 2019).

## **Aim of our study**

- Identify associations between individual mid-infrared wavenumbers and regions of the genome.

# Identify genomic associations

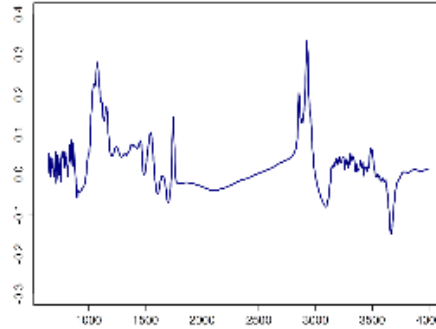


← GWAS →





# Identify genomic associations



← GWAS →



## Dataset

- 100,671 FTIR records: 38,085 mixed-breed NZ dairy cattle.
- Imputed sequence: 17,776,293 variants.
- Individual wavenumbers standardized (Grelet et al., 2015; Tiplady et al., 2019).
- Adjusted for parity, days in milk, breed, heterosis and herd by test date (ASReml mixed linear model).

# Identify genomic associations

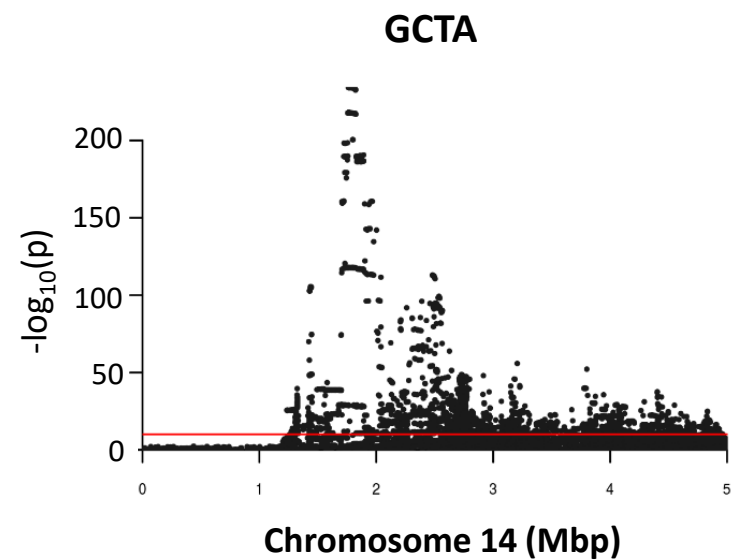
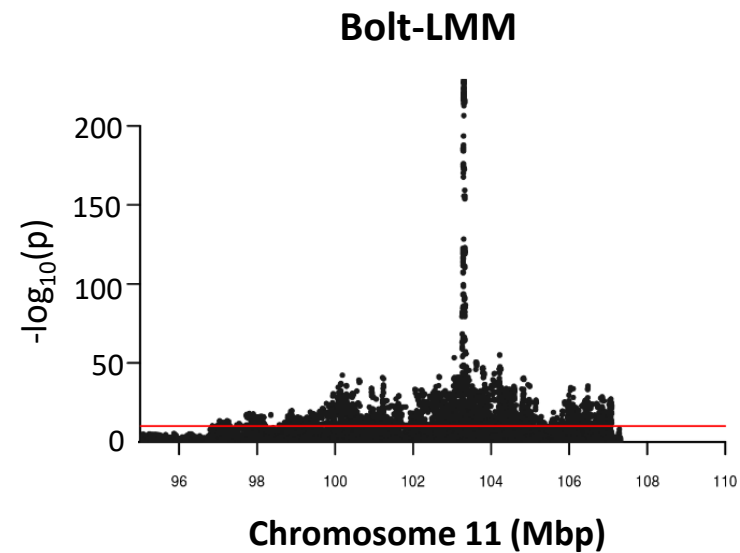
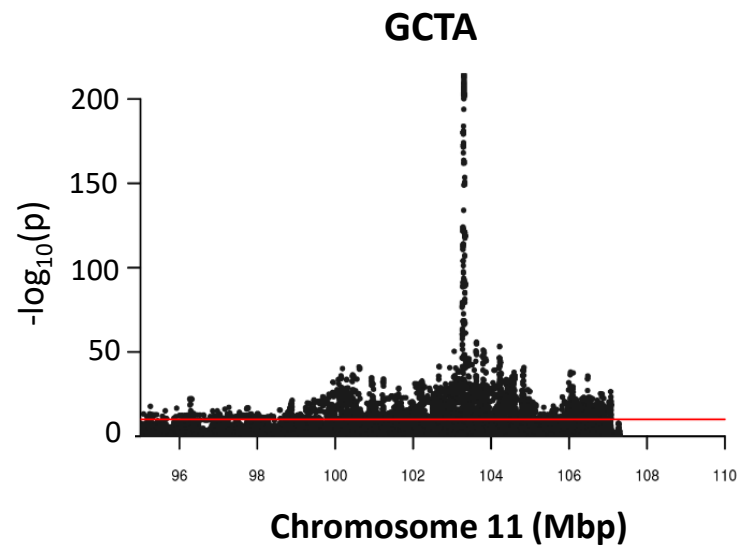
**Sequence-based GWAS: ~38k animals, ~17.7m variants, 895 traits**

## Computation challenges

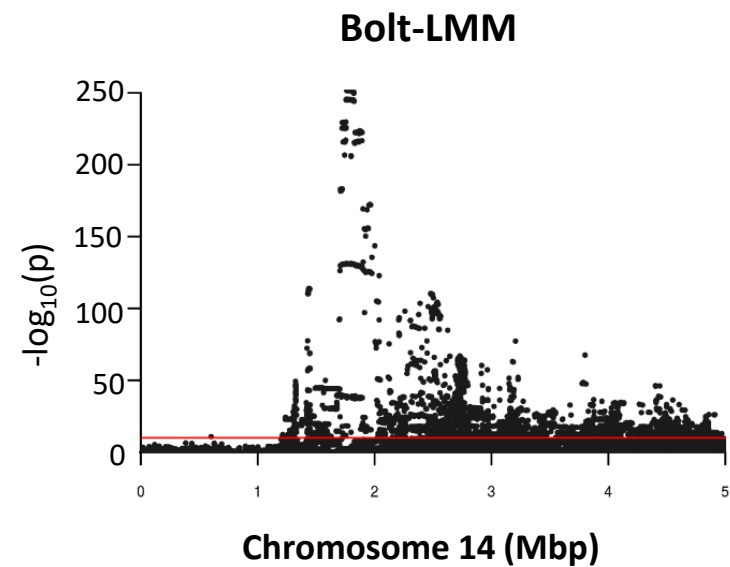
- GCTA (Yang et al., 2011): 1800 hours per trait  $\Rightarrow$  ~ 6 years
- Bolt-LMM (Loh et al., 2015): 6 hours per trait  $\Rightarrow$  ~ 7 days
- Comparisons made using 43,851 SNP to account for population stratification.



# Software comparisons

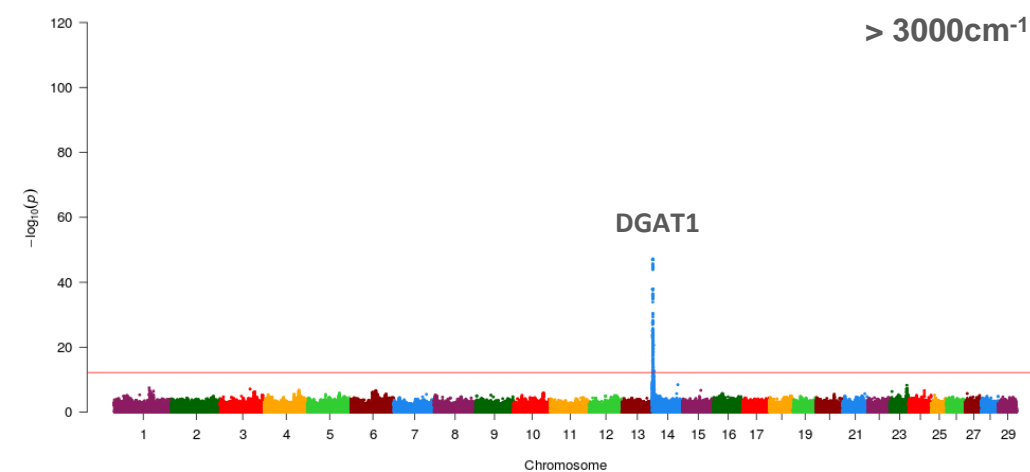
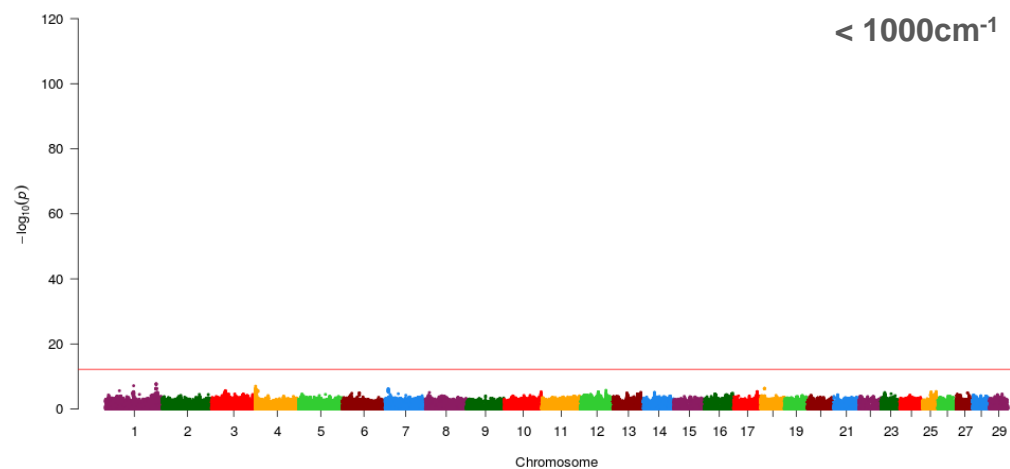
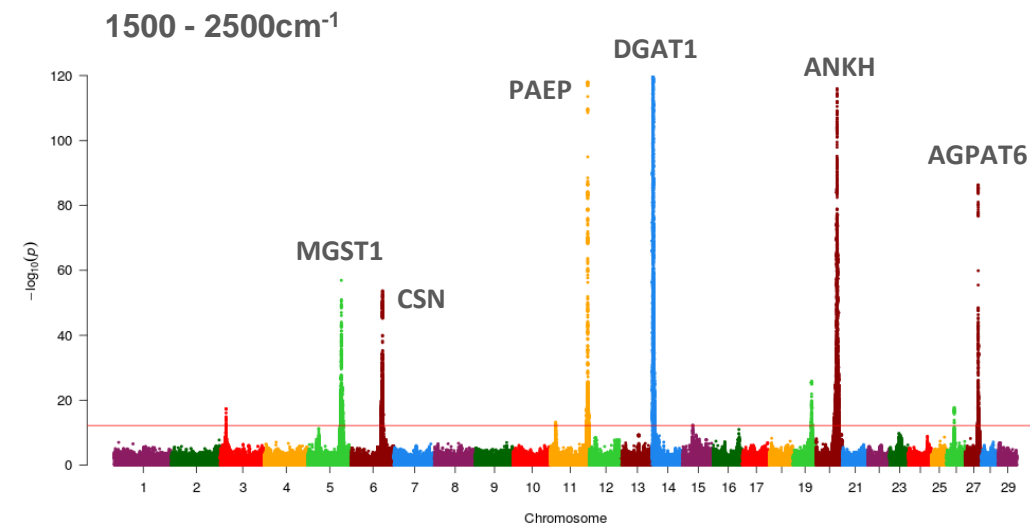
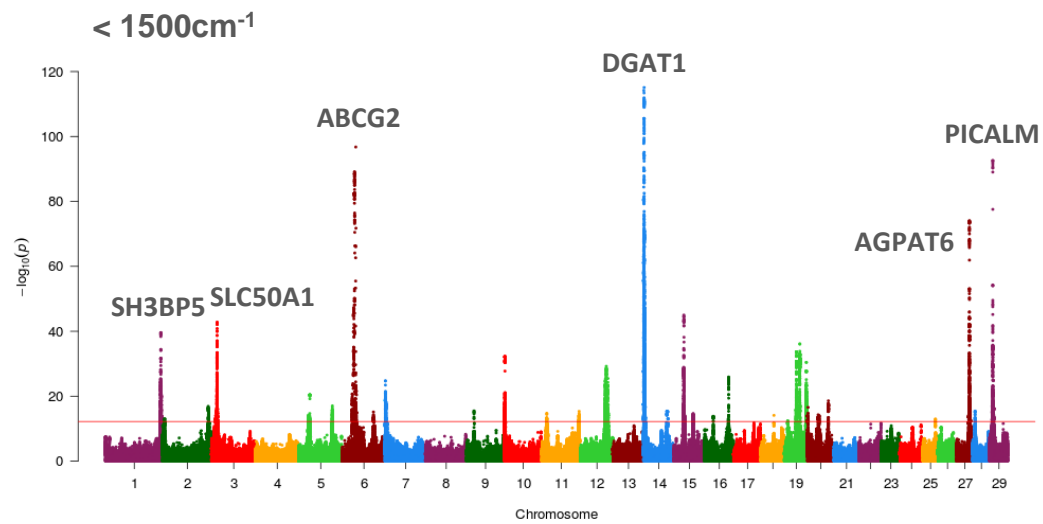


~1800 hours/trait



~6 hours/trait

# Results



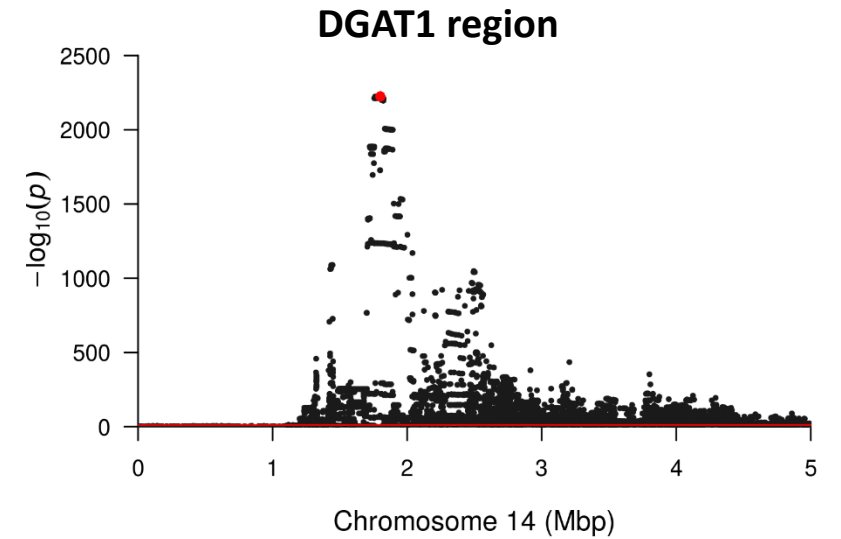
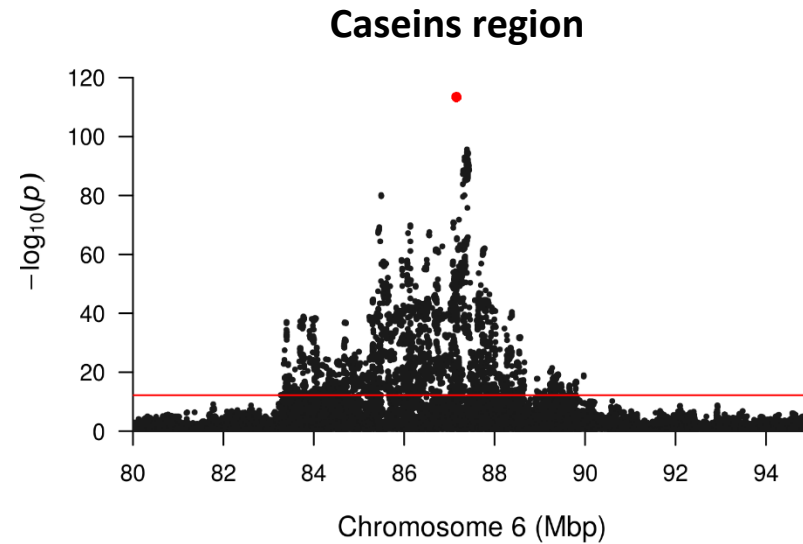
Bonferroni-threshold:  $0.01/17.7m/895=6.3 \times 10^{-13}$ .

Significant associations conserved across multiple wavenumbers: DGAT1 significant for over 750 wavenumbers!

# Peak identification

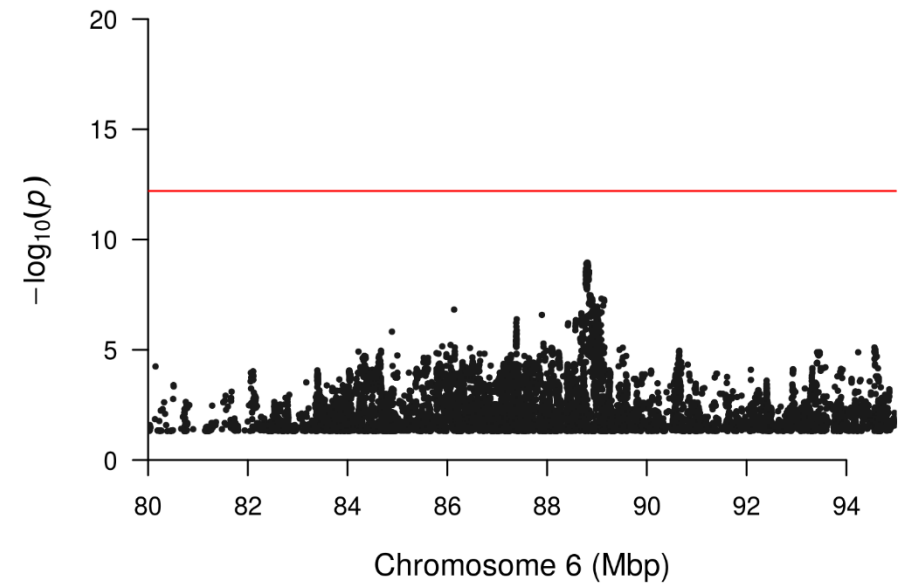
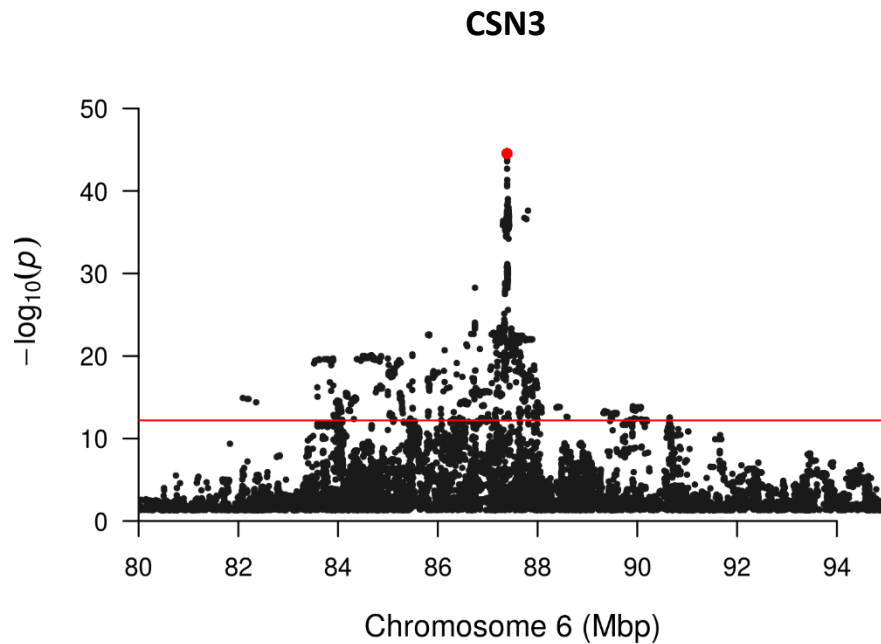
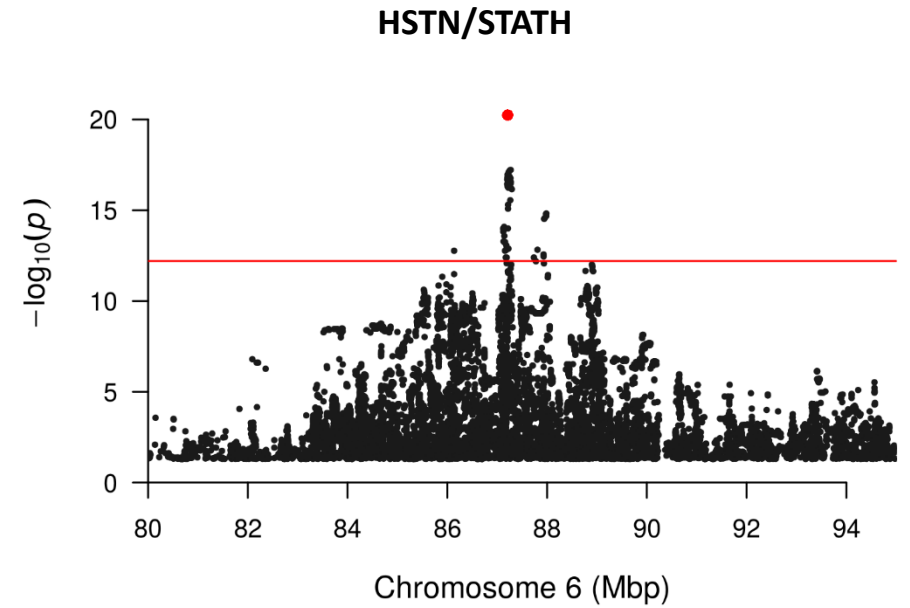
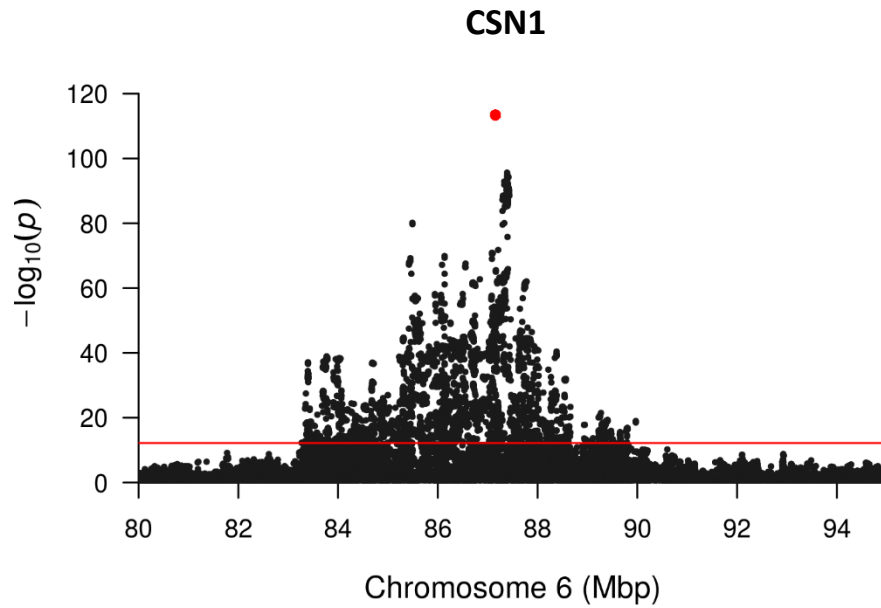


- How to deal with complex regions?  
e.g: Caseins region on BTA6; DGAT1 region at the start of BTA14.

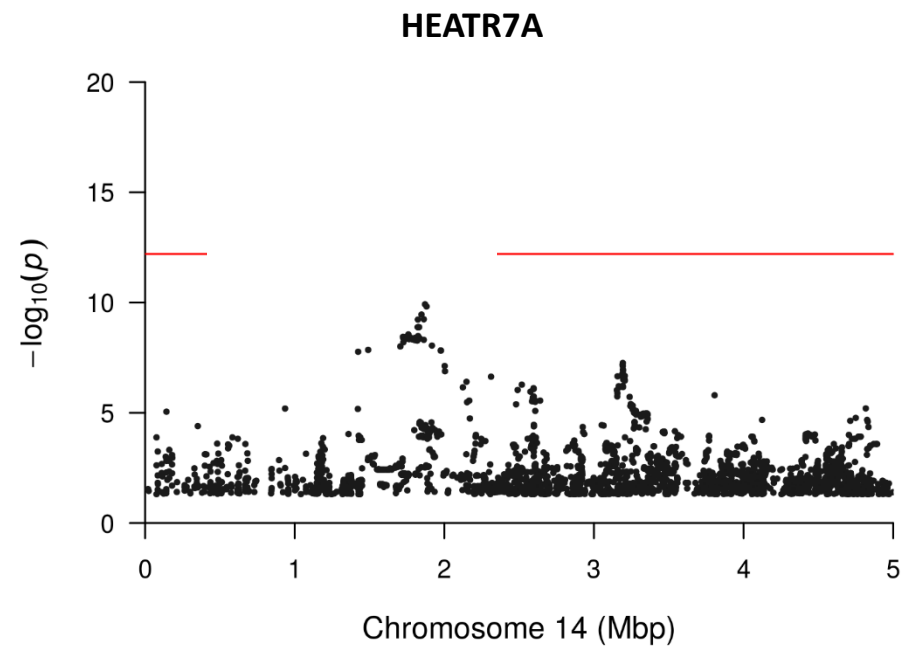
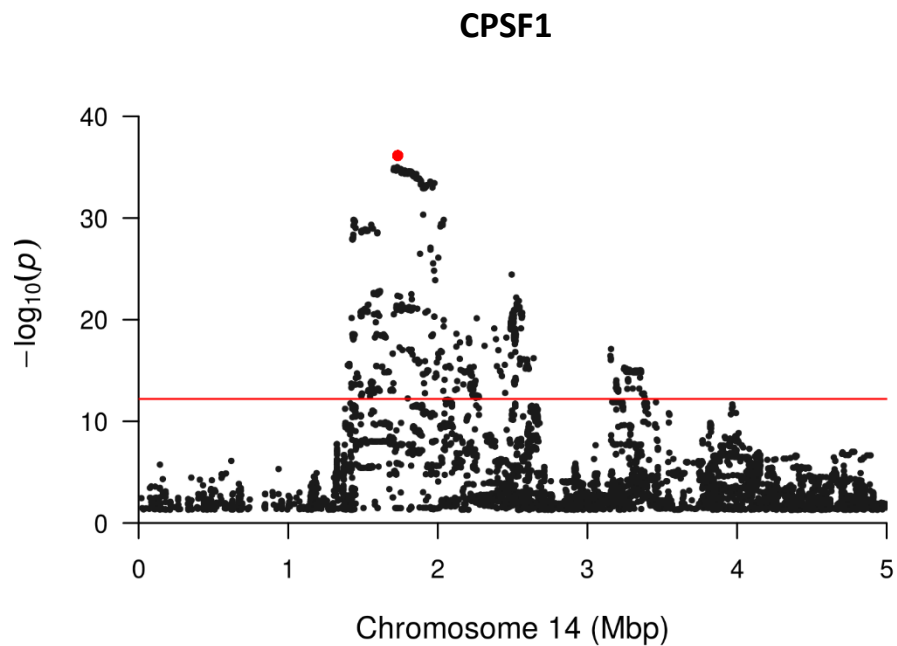
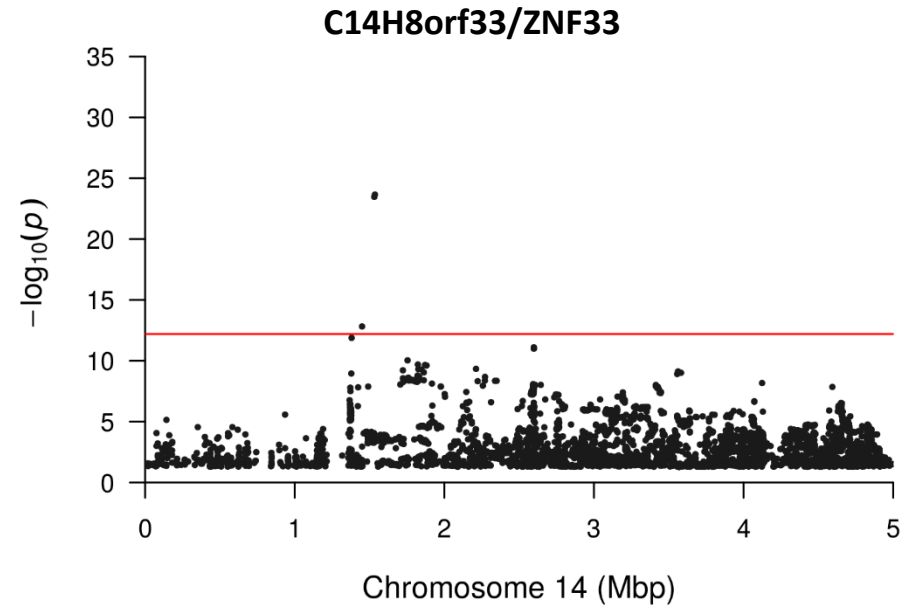
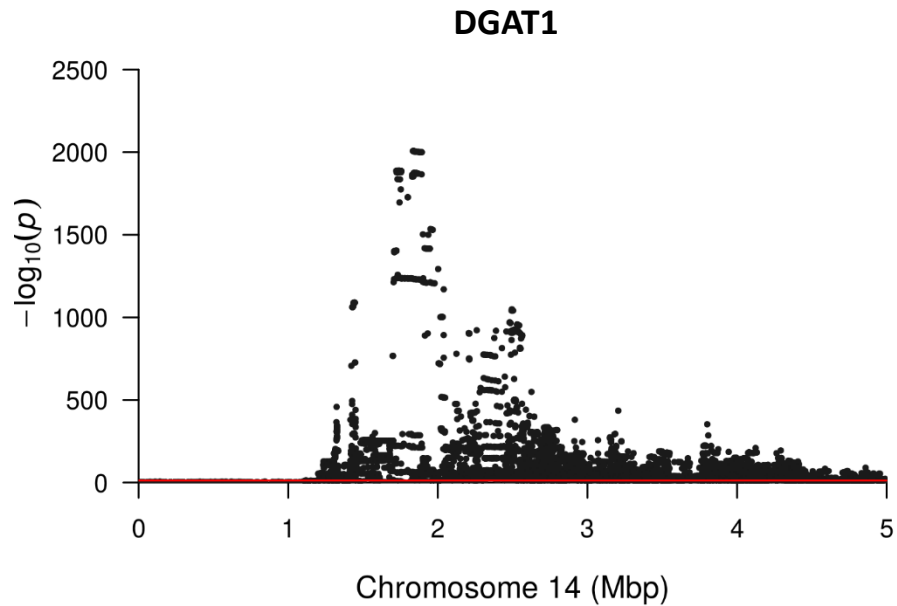


- Conditional iterative GWAS for each wavenumber.
  - Top SNP on each chromosome fitted as a covariate.
  - Iteratively run until there no signal left.

# Peak identification

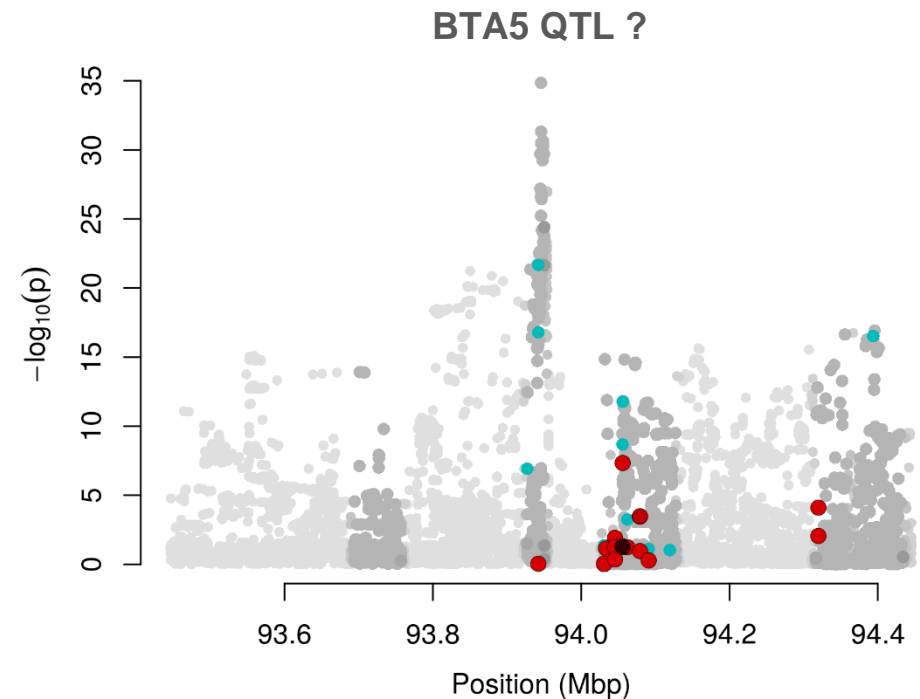
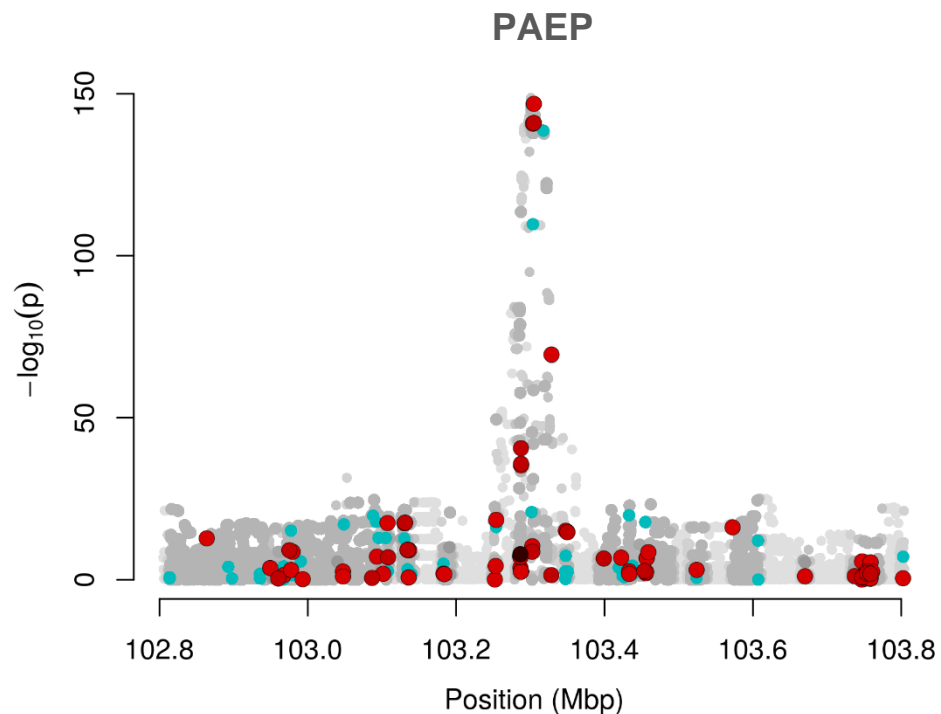


# Peak identification



# Determine likely candidates

Link associations to SnpEff annotation - assess the candidacy of variants in terms of their predicted impact on protein sequence.

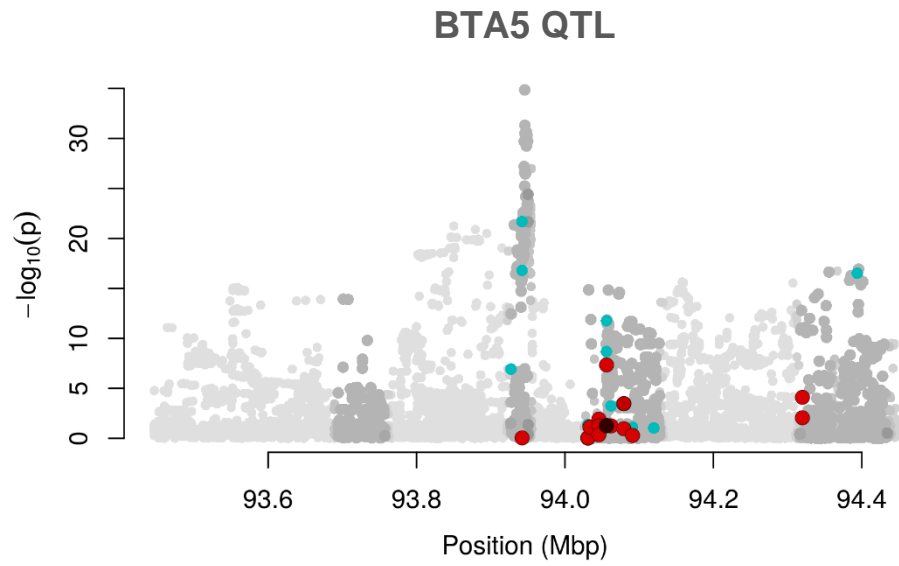


- High impact: Slice donor/acceptor, frameshift variants
- Moderate: Missense, splice region variants

- Low impact: Synonymous variants, non-coding exons
- Modifiers: Intron and up/downstream variants, UTRs.

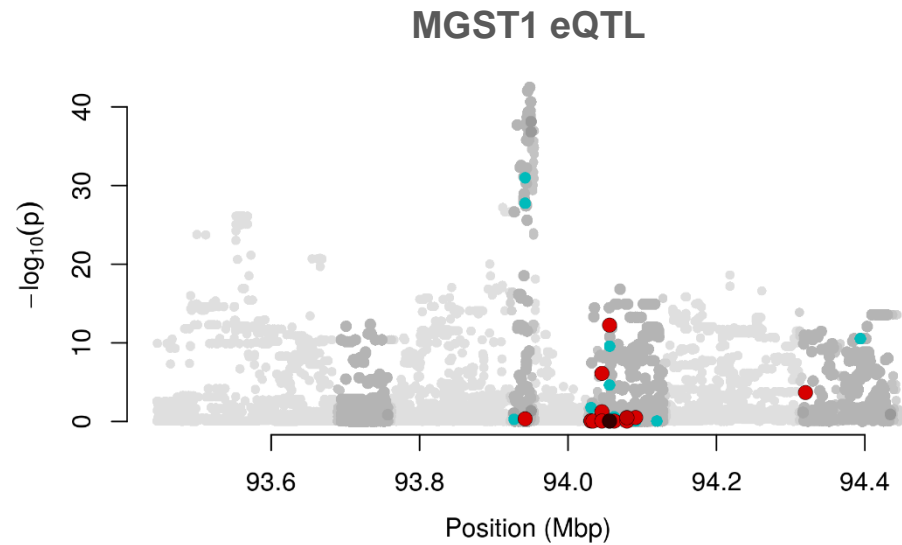
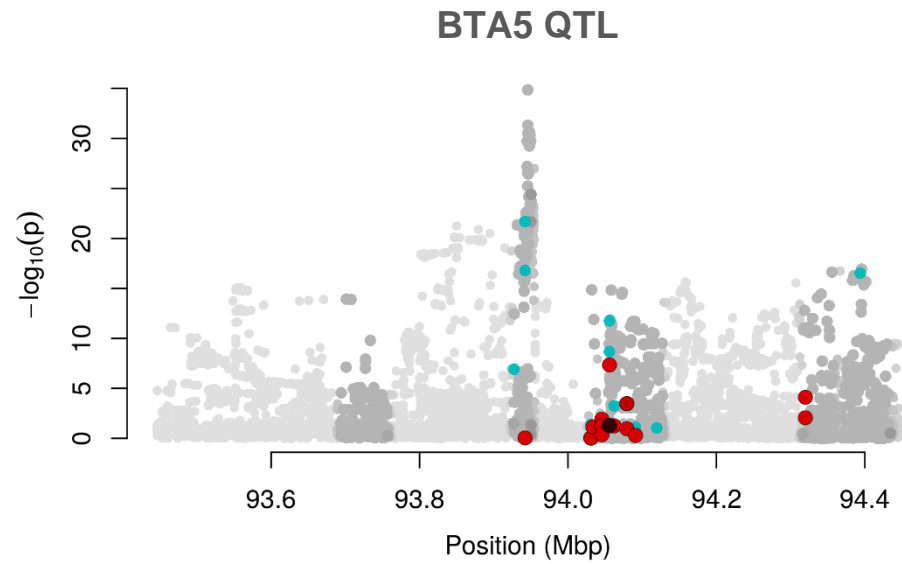


# Co-locating eQTLs



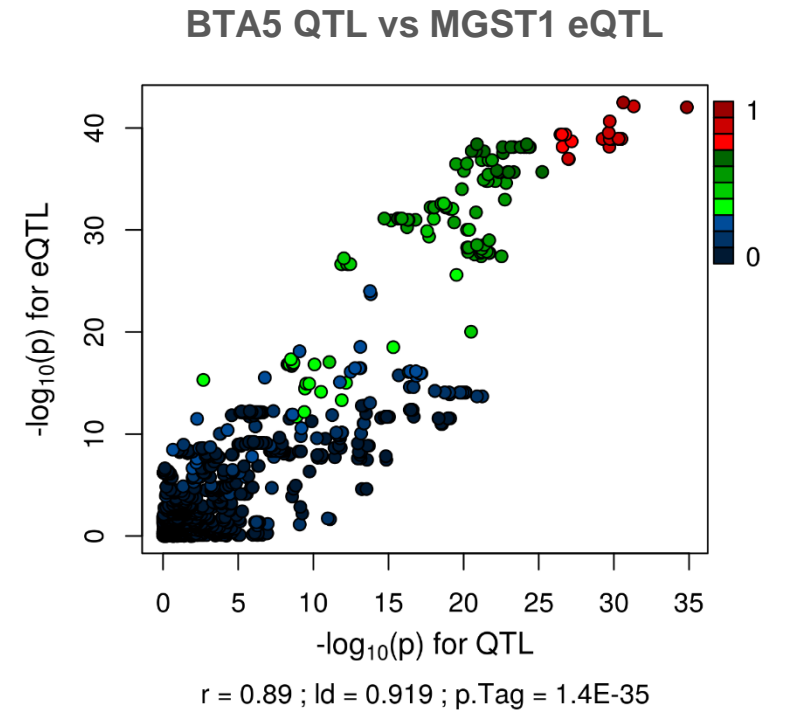
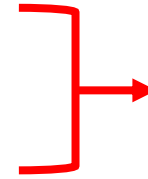
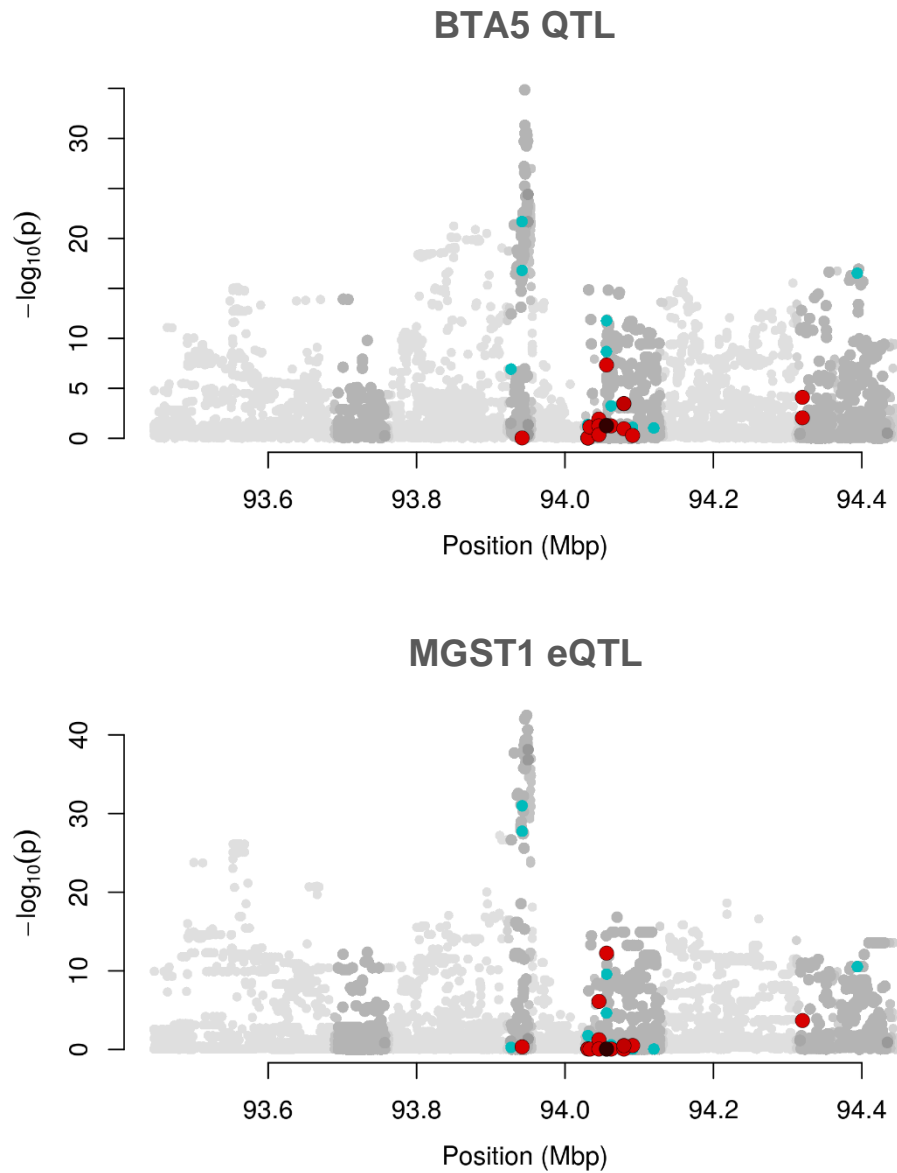
- Gene expression data from mammary tissue in 375 animals (Lopdell et al., 2017).
- A co-locating eQTL indicates genetic regulation of the gene in the same region that we see the trait QTL.

# Co-locating eQTLs



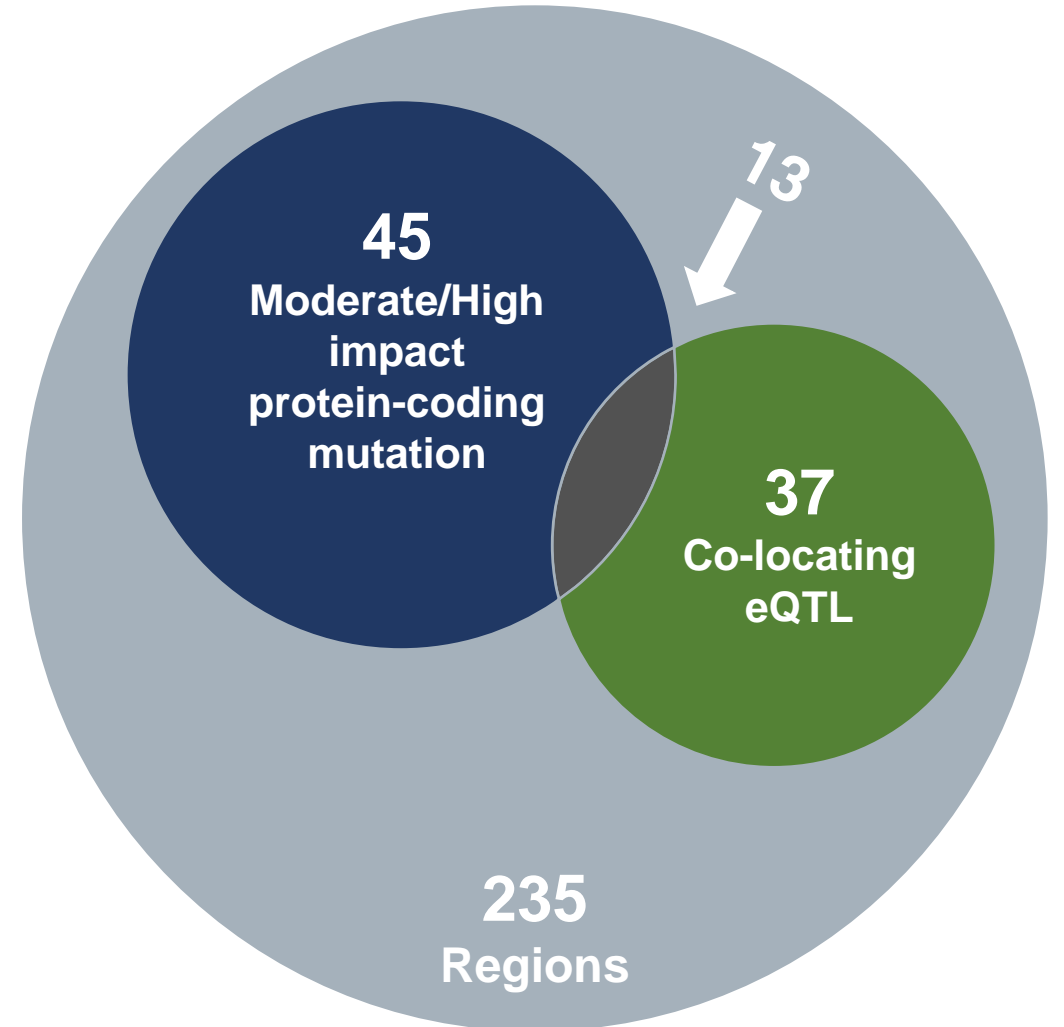
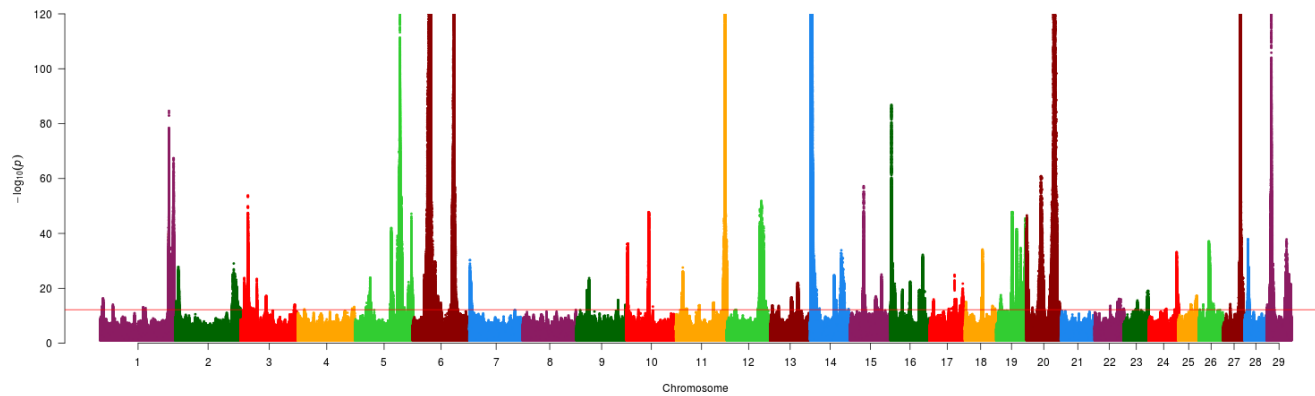
- Gene expression data from mammary tissue in 375 animals (Lopdell et al., 2017).
- A co-locating eQTL indicates genetic regulation of the gene in the same region that we see the trait QTL.

# Co-locating eQTLs



- Gene expression data from mammary tissue in 375 animals (Lopdell et al., 2017).
- A co-locating eQTL indicates genetic regulation of the gene in the same region that we see the trait QTL.

# Peak overview



# Summary

---

- GWAS on mid-infrared wavenumbers conducted
  - Bolt-LMM: ~38k animals, ~17.7m variants, 895 traits
  - Multiple QTL in complex regions  $\Rightarrow$  Conditional iterations.
- Promising peaks with candidate variants and co-locating eQTLs identified.
- Ongoing research to consolidate results and determine how we can use this information to improve genomic selection.



# Acknowledgements

## Sequence-based genome-wide association study of milk mid-infrared wavenumbers in dairy cattle

T. Lopdell, E. Reynolds, R. Sherlock, M. Keehan, T. Johnson, J. Pryce, H. Blair, S. Davis, M. Littlejohn, D. Garrick, R. Spelman and B. Harris

Tracey Monehan (Project manager)

Dorian Garrick (Massey, AL Rae Centre)

Matt Littlejohn (Massey/LIC)

Jennie Pryce (Agriculture Victoria)

Hugh Blair (Massey)

