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Tissue distribution and pharmacological characterization of bovine free fatty acids-sensing GPCRs

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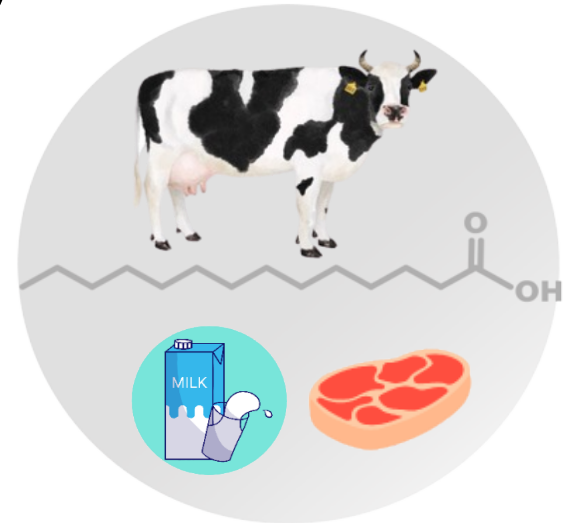
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INRAE

Introduction

Fatty acids (FA):

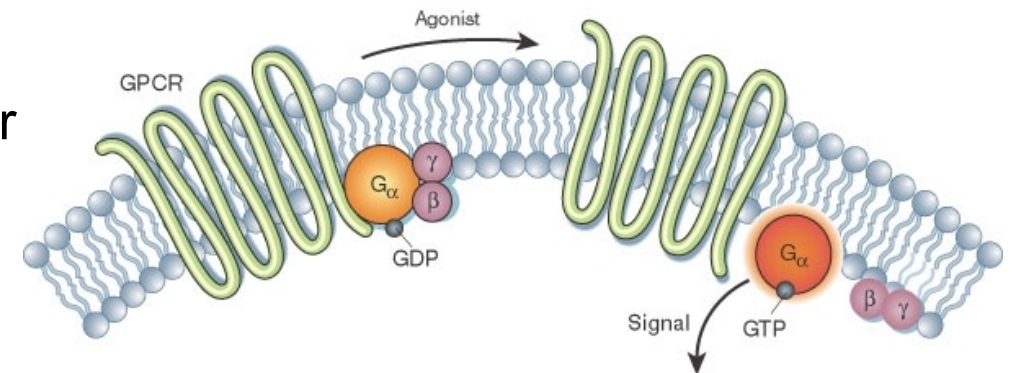
- Growth, production, and metabolism adaptation (e.g., peripartum)
- **More than energy sources**
- **Signaling molecules** → regulation of metabolic functions
- Binding to different receptors:
 - SREBP, PPARs, liver X receptor (LXR) – nuclear
 - TLR4, TLR2, CD36 – cell membrane
 - G protein-coupled receptors



Introduction

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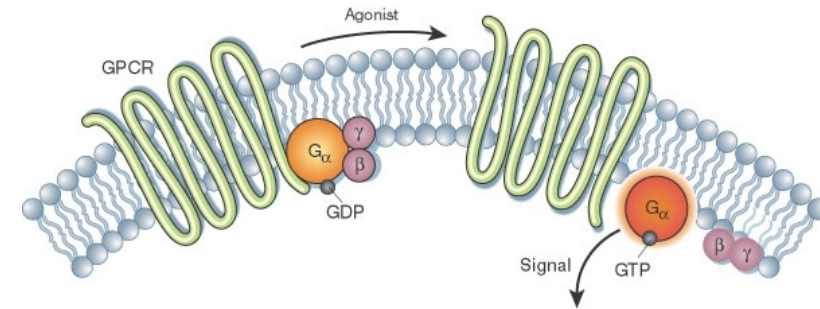
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 - **G protein-coupled receptors**



Introduction

G protein-coupled receptors (GPCR):

- Many GPCRs are activated by FFA
- FFAR1, 2, 3 and 4, GPR84



Humans and mice

- Tissue distribution/pharmacological properties
- Activation of signaling pathways and their associated biological outcomes:
e.g., insulin secretion, inflammation, lipolysis
- FFAR as pharmacological targets for the treatment of different diseases

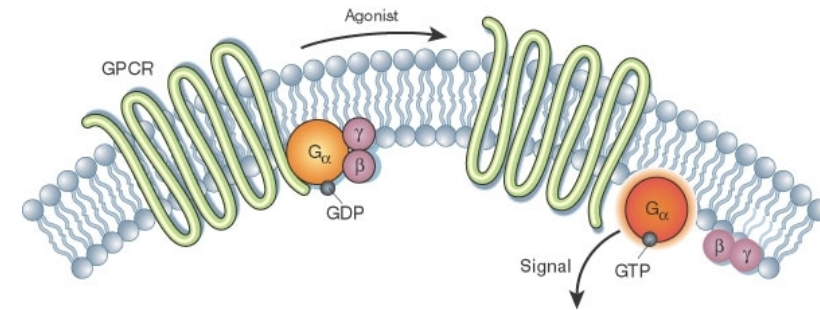
Dairy cows



Introduction

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Dairy cows

- Scarce information
- Gene expression on different tissues

Objectives

- The objective of our study was to characterize 5 bovine FFARs (FFAR1 to 4, and GPR84) in regards of tissue distribution
- Moreover, we aimed to characterize the pharmacological properties of FFARs (FFAR1 and FFAR2)



Materials and methods

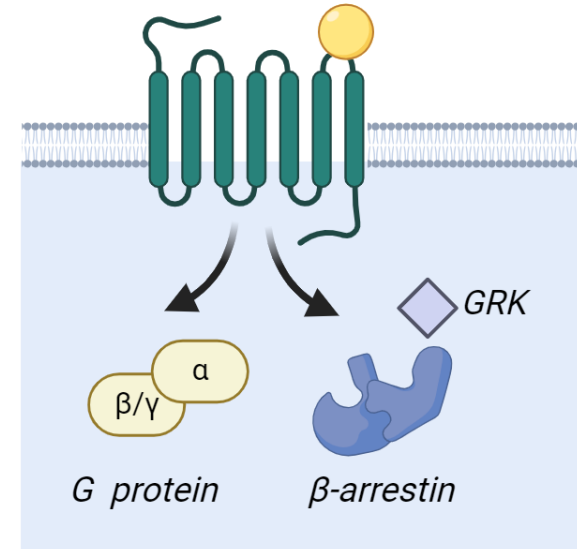
Tissue distribution:

- 16 Charolais bulls (16-18 months old)
- Samples from 6 tissues:
 - Liver
 - Ileum
 - Rectum
 - Spleen
 - Longissimus thoracis (LT)
 - Perirenal adipose tissue (PRAT)
- Total RNA was extracted and gene expression assessed by RT-qPCR

Materials and methods

Pharmacological properties:

- HEK293a cells
- FFAR transfected individually
 - mG proteins (mGq, mGi, mG12, mGs)
 - B arrestin



Unbiased ligand: $G = B$
Bias ligand: $G > B$ or $G < B$

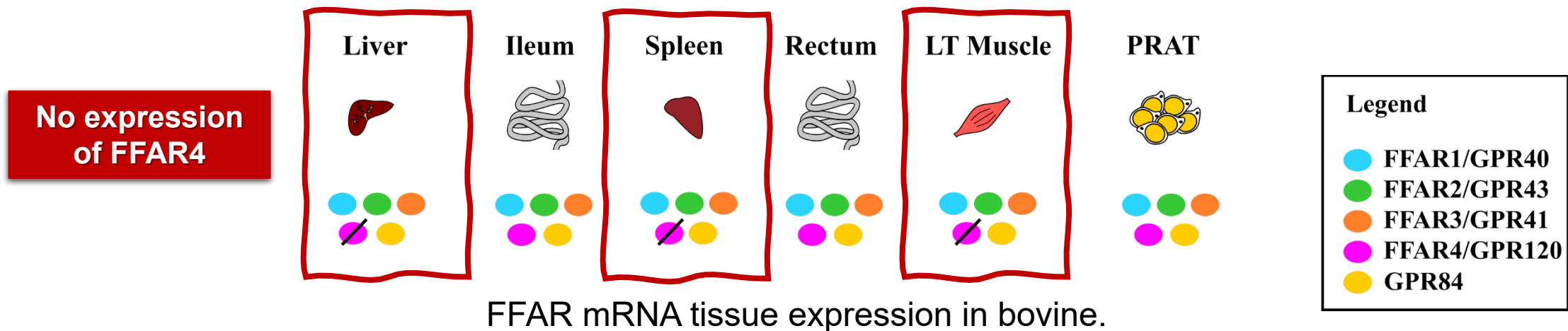
- Bioluminescence Resonance Energy Transfer (BRET)



Results

Tissue distribution:

- FFAR1, FFAR2, FFAR3 and GPR84 – expressed in all studied tissues
- FFAR4 expression was restricted to ileum, rectum, and PRAT



Results

Pharmacological properties:

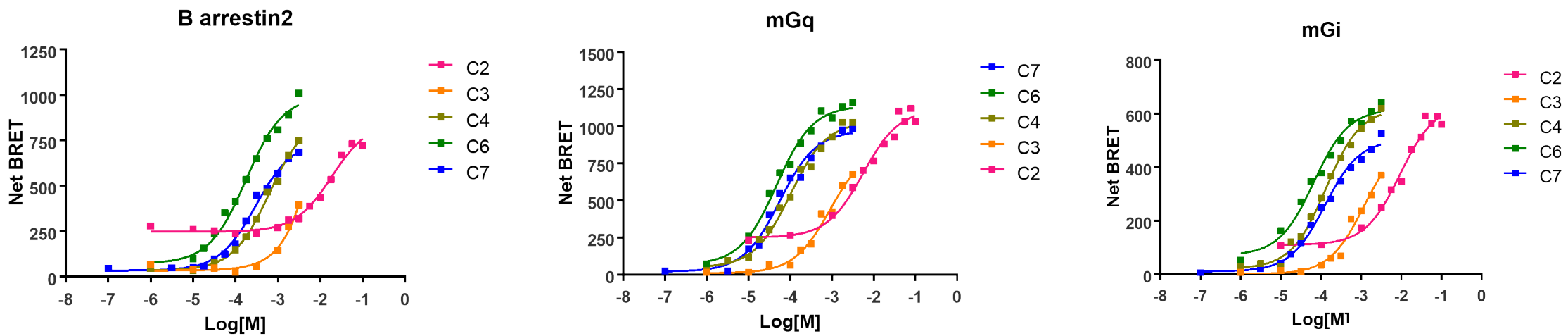
Potency ($-\log EC_{50} \pm SEM$) of SCFA in G proteins and B arrestin signaling in bovine FFAR2.

Fatty acid	mGi	mGq	B arrestin 2
C2	1.961 \pm 0.07	1.961 \pm 0.07	1.729 \pm 0.09
C3	2.211 \pm 0.07	2.211 \pm 0.07	1.661 \pm 0.06
C4	3.906 \pm 0.06	3.906 \pm 0.06	3.035 \pm 0.06
C6	4.036 \pm 0.07	4.036 \pm 0.07	3.842 \pm 0.06
C7	3.901 \pm 0.08	3.901 \pm 0.08	3.518 \pm 0.07
C8 and C10	-	-	-

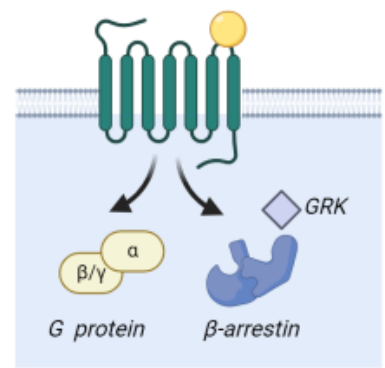
- C8 and C10: Did not showed a typical dose-response curve

Results

Pharmacological properties:



- C6: Greatest efficacy (Emax) and potency (-logEC50)
- No bias observed for SCFA in activating mGq, mGi or B arrestin 2 signaling



Unbiased ligand: G = B

Results

Pharmacological properties:

- LCFA conjugated or not with BSA (4:1 molar ratio)
- Fatty acids from 7 to 22 carbons
- G protein recruitment restricted to mGq:

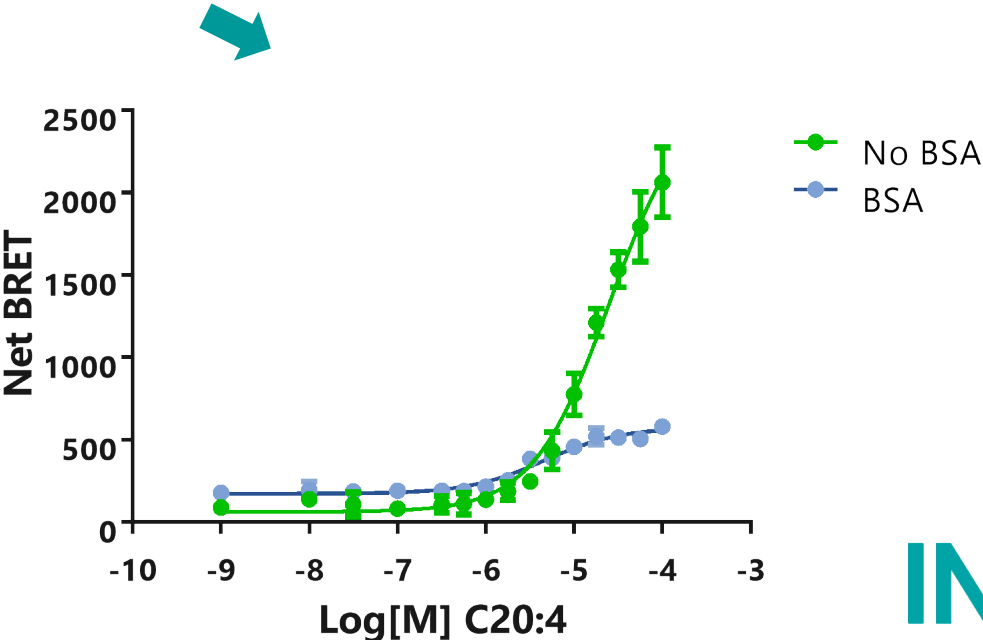
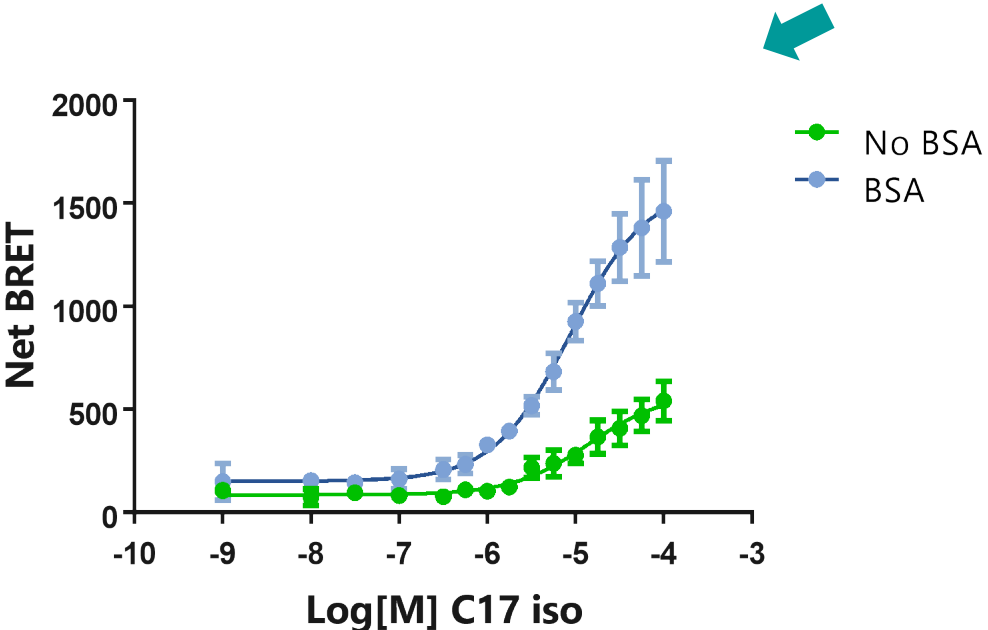
C22:6/BSA: Greatest potency (EC50) → Omega-3, Anti-inflammatory and antioxidant properties

C18:3 (ALA): Greatest efficacy (Emax) → Omega-3, Essential fatty acid

Results

Pharmacological properties:

- Efficacy (Emax) and potency (-logEC50) affected by FFA and BSA
- Efficacy (Emax): Either increased or decreased by BSA conjugation



Conclusions and Future Perspectives

Bovine FFAR2:

- Greater potency in SCFA with more carbons (C6)
 - Contrast to what is observed in humans (C2)

Bovine FFAR1:

- Conjugation with BSA affects FFAR1 response: Possible implications?
 - Activation of receptors in the GI tract by dietary FFA
 - Periods of alterations in FFA:albumin ratio (e.g., during transition period)



Conclusions and Future Perspectives

Future perspective:

- Assess B arrestin recruitment of FFAR1
- Determine pharmacological properties of the others bovine FFAR
- Further understand the biological outcomes associated with the activation of FFARs in cattle
- Possible association with, e.g., metabolic disorders, adaptation around parturition?



Acknowledgments



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Dr. Guillaume Durand

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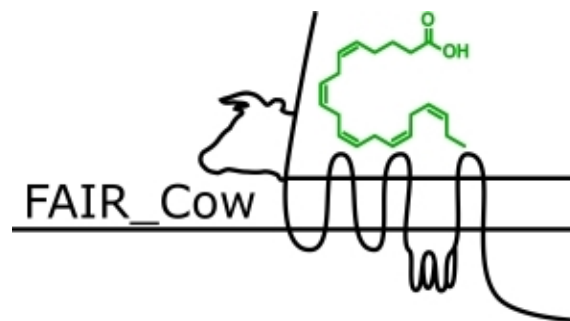
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Thank you!