



Animal Production System Area

The characterization of the microbiome and resistome in feces and slurry of dairy cows in different livestock production systems

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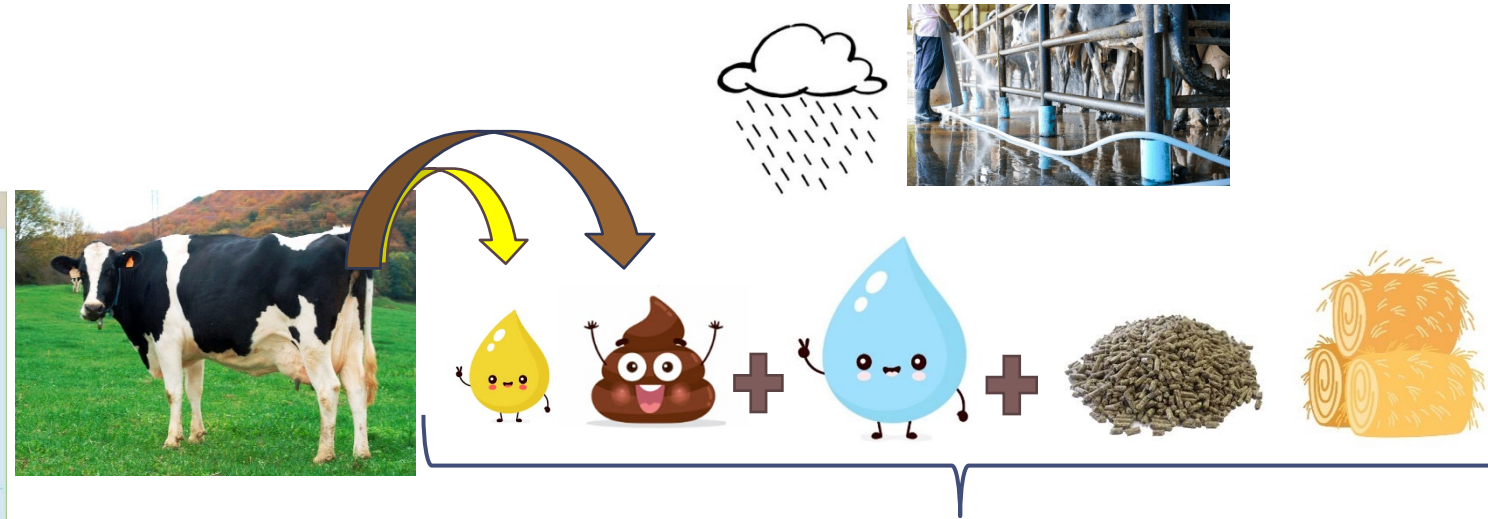
September, 01st 2024 (Florence, Italy)

Cantabria, Spain



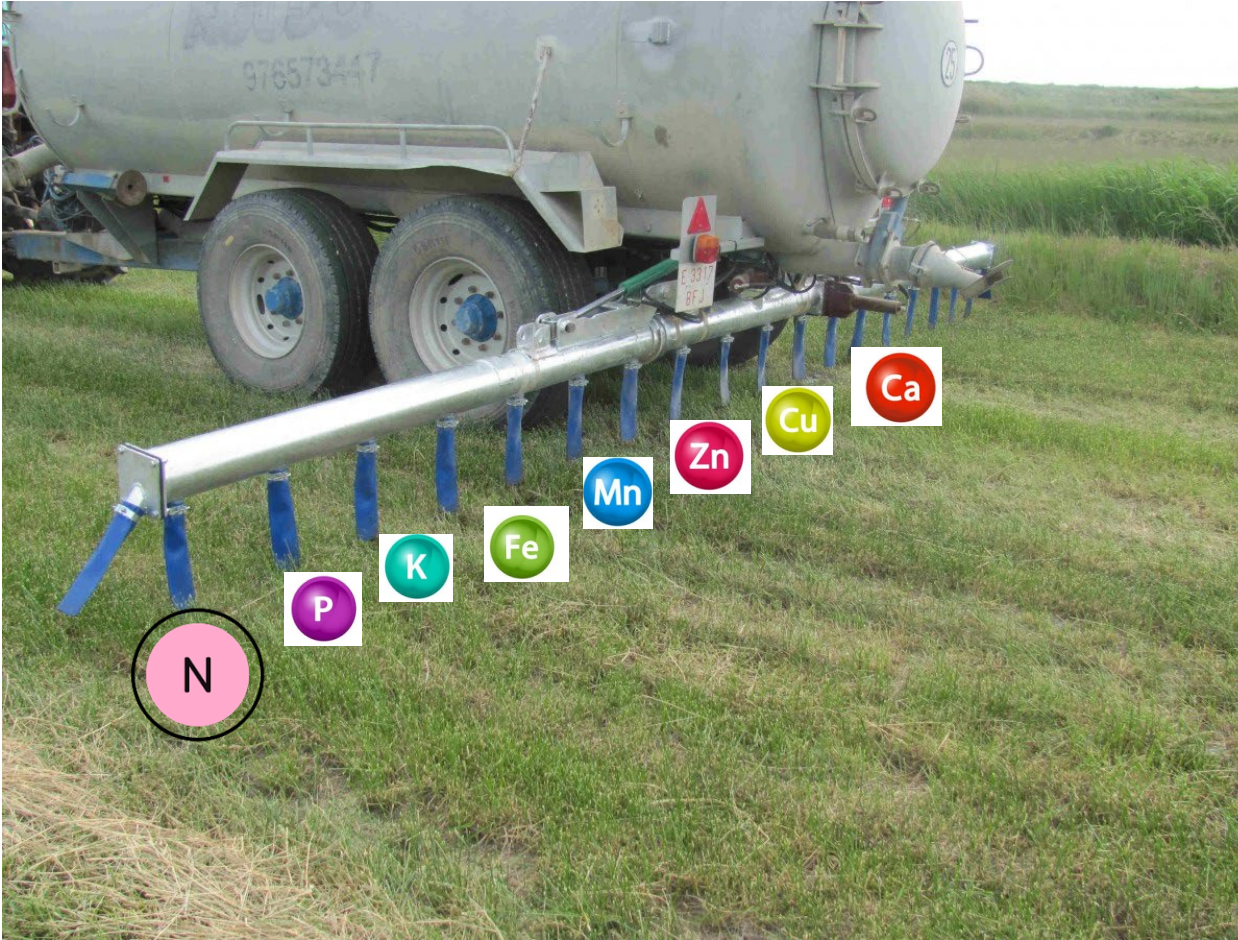
Figure 1: The 17 autonomous regions of Spain
(<http://mapasenpdf.com/descarga?pdf=maps%2Feducation%2Fespana%2Fpdfs%2Fmapa-politico-espana-para-imprimir.pdf>)

- Heads of dairy cattle: **164,406** (MAFF, 2024)
- Number of dairy cattle farms: **838** (MAFF, 2024)



55 l/cow/day ➡ 9 000 tons

Used as an organo-mineral fertilizer



- High agronomic value;
 - Soil quality improvement
 - Yield and quality improvement of crop and pasture





- Pathogenic and non-pathogenic bacteria
- Bacteria and antimicrobial resistance genes



World Health
Organization

By **2050**, multi-resistant bacteria could kill **10 million people** per year, surpassing cancer as our main health concern (WHO, 2021).



Hypothesis

H_0 : Livestock farms present the same ARG and ARB load according to the production system, and there are no significant differences.



H_a : Livestock farms present different ARG and ARB load according to the production system, and there are significant differences.



Microbiome



Resistome



Intensive system



Conventional grazing system



Organic grazing system



During 4 years (2021-2024)

Livestock Production Systems

Conventional grazing



Organic grazing

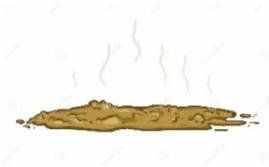


Intensive



Total feces samples = 300

Total slurry samples = 90



Collection from the rectum

Collection from piles of accumulated slurry

1- Selection of production systems and farms

2- Questionnaires realization

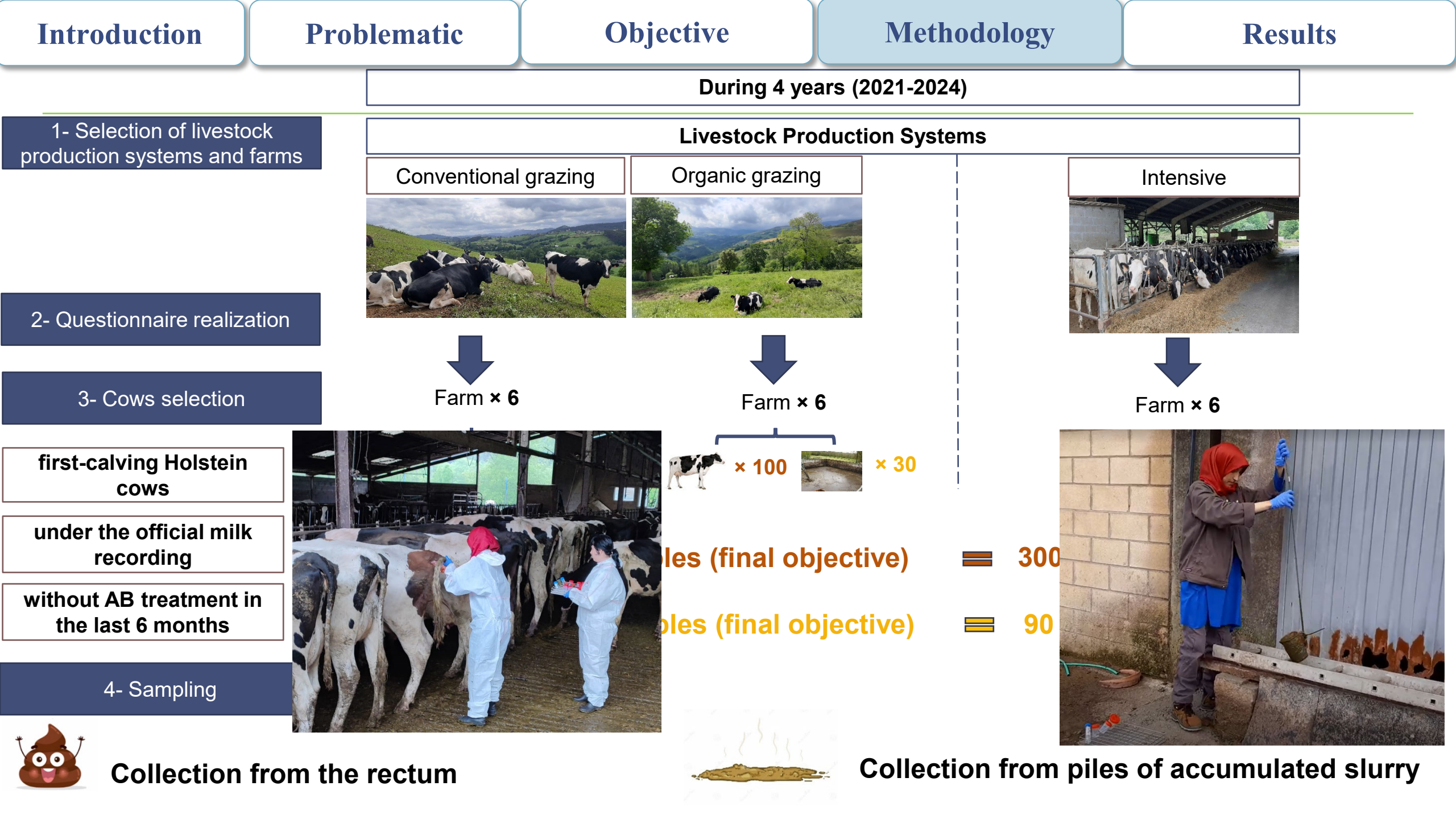
3- Cows selection

first-calving Holstein cows

under the official milk recording

without AB treatment in the last 6 months

4- Sampling



Physicochemical conditions?

Physicochemical analysis

Physical parameters

- pH
- Electrical conductivity
- Total dissolved solids (TDS)

Chemical parameters

- Dry Matter(drying oven at 103°C)
- Ash (muffle furnace at 500°C)
- Cations (atomic absorption)
- Organic nitrogen (Kjeldahl method)
- Ammoniacal nitrogen (distillation)



Multiparameter (HI 991301, Hannah Instruments)



Microorganisms ?

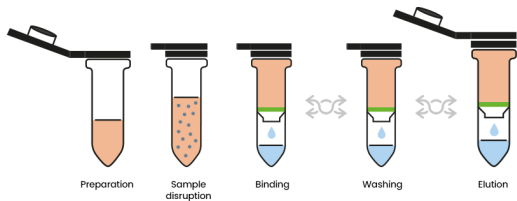
1- DNA extraction and purification



**QIAamp PowerFecal Pro
DNA Kit**

2- DNA quantification

Estimate the concentration of extracted DNA by **fluorescence** using a fluorometer (Qubit 4.0)



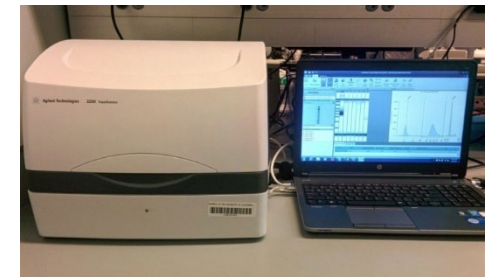
3. DNA purity control

Verify the purity of DNA and the presence of other components (proteins, salts, etc.) using a spectrophotometer (V-730, Jasco) with 260/280 and 260/230 nm ratios

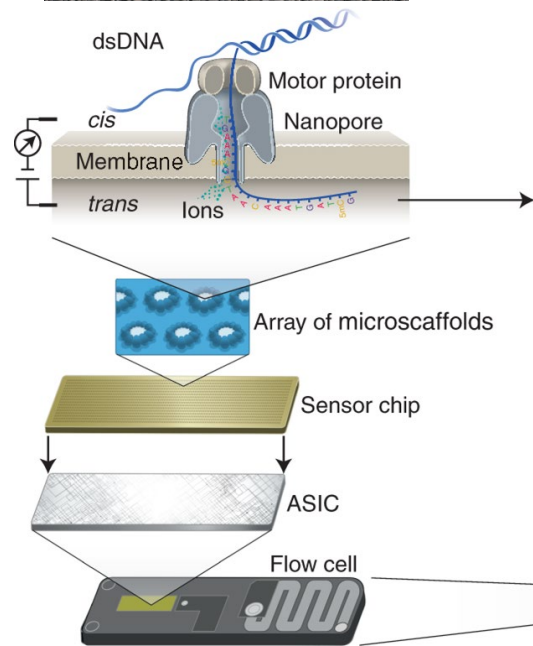
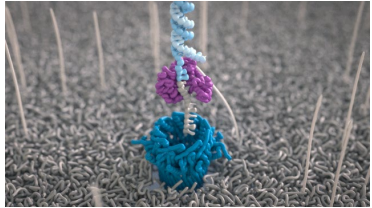


4. DNA quality control

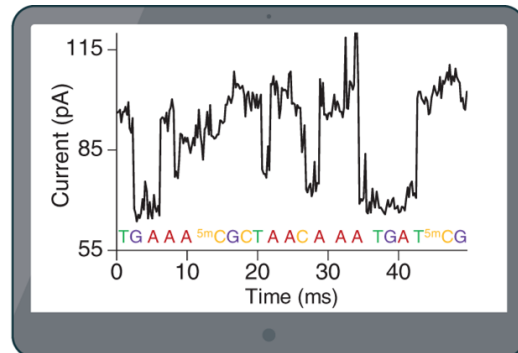
Assess the degree of integrity using automated electrophoresis (Agilent TapeStation) to determine the DNA integrity number (**DIN**)



5- Library construction and sequencing



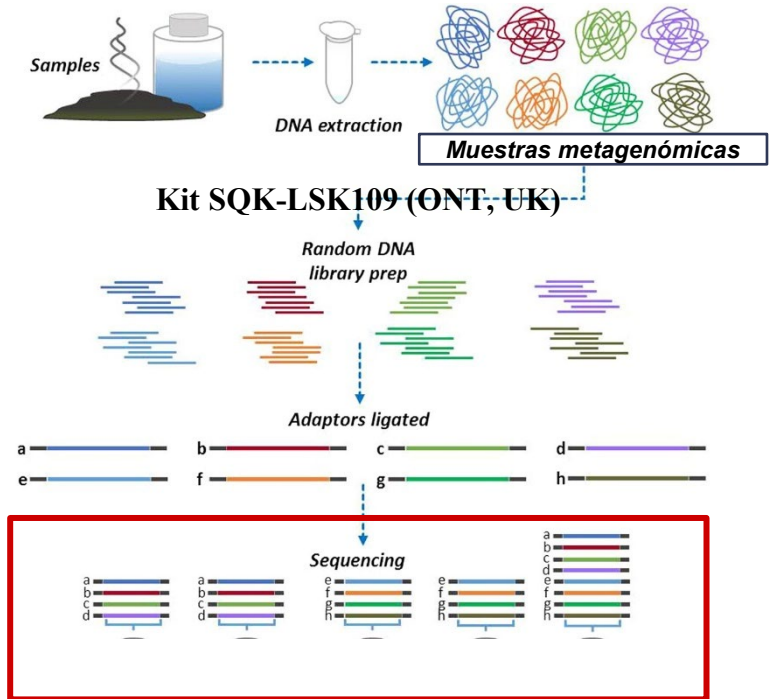
Basecalling (Guppy, SUP)



Fastq



Flow Cell



MinIon Mk1C device
(Oxford Nanopore Technology)

6- Bioinformatic analysis

Metagenomics pipeline:
SqueezeMeta (Tamames
and Puente-Sánchez, 2019)

A fully automatic pipeline
for metagenomics, covering
all steps of the analysis:
assembly, ORF prediction,
annotation and binning.

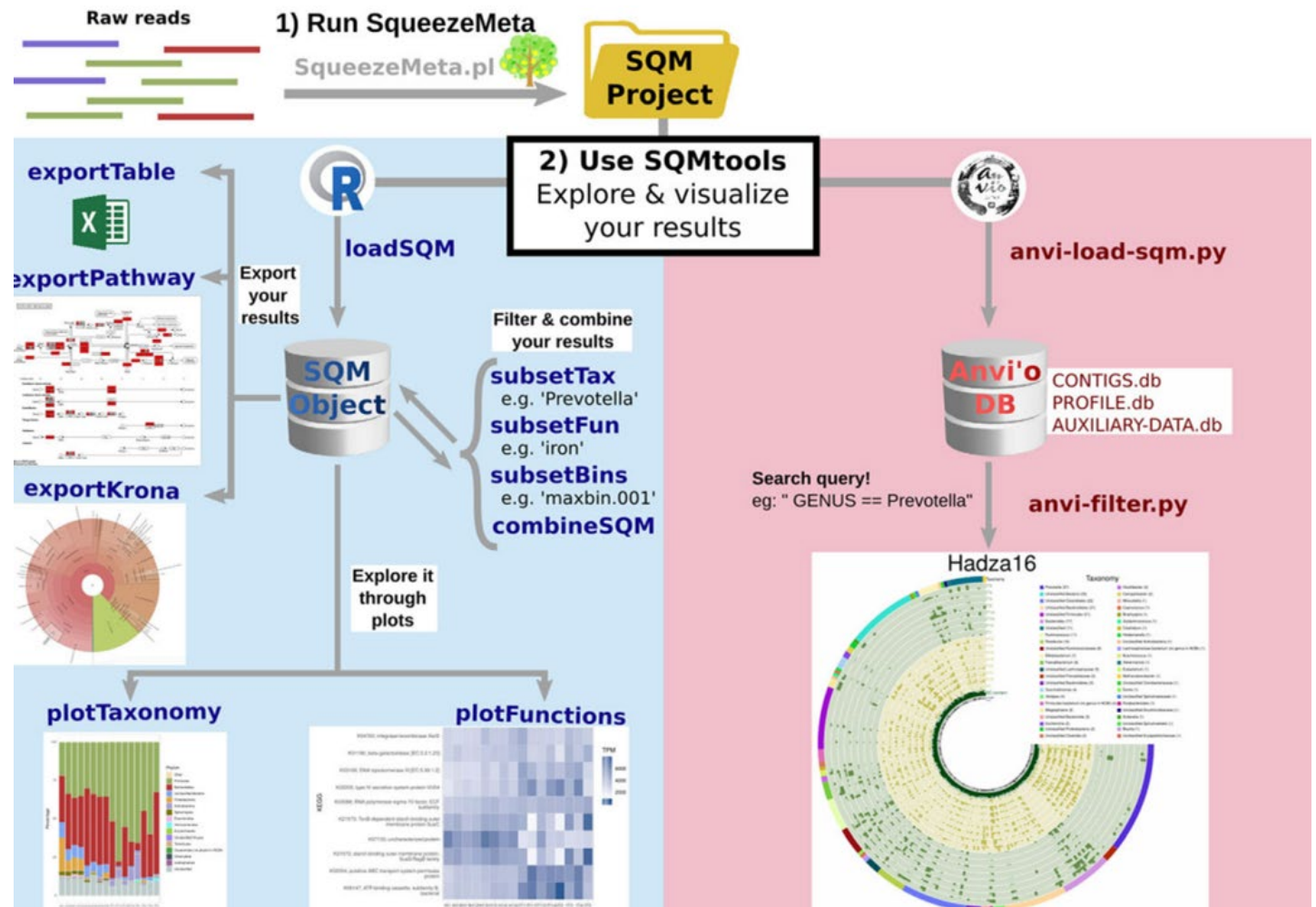


Figure2. Main workflow of the SQMtools pipeline (Tamames and Puente-Sánchez, 2019)

- Metagenomics pipeline: SqueezeMeta (Tamames and Puente-Sánchez, 2019)
- High Performance Computing (HPC) of Castilla and León (SCAYLE.

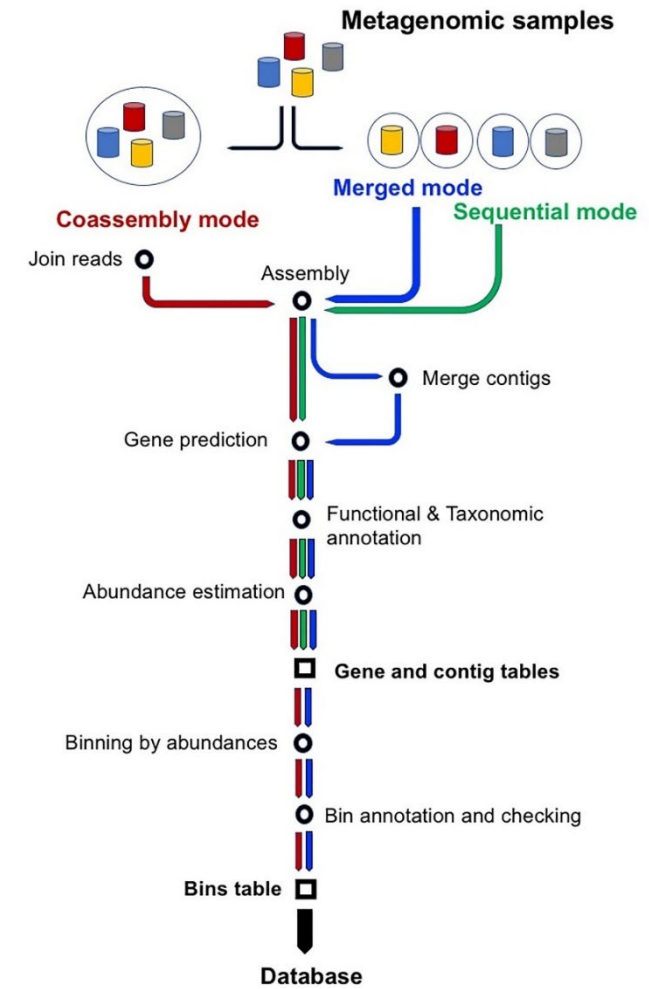
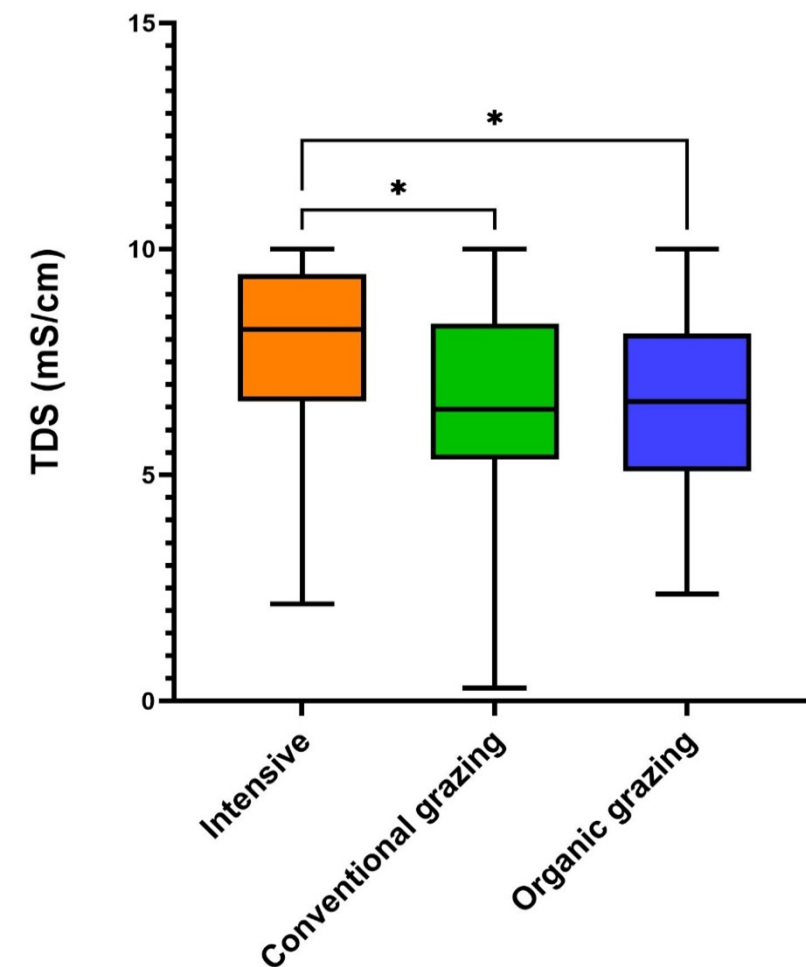
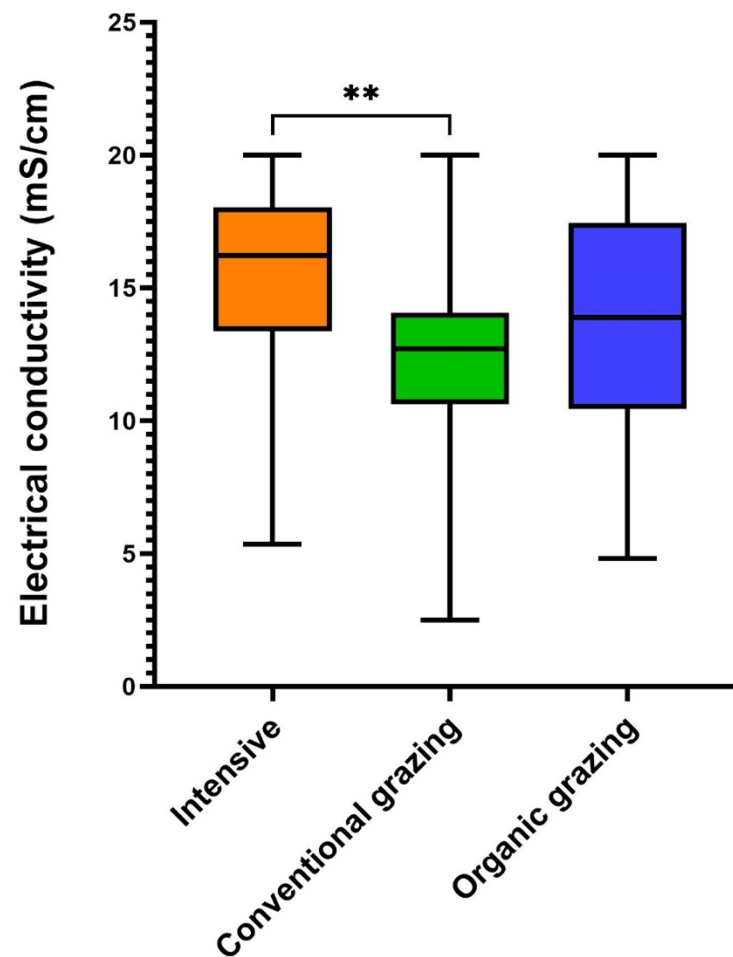
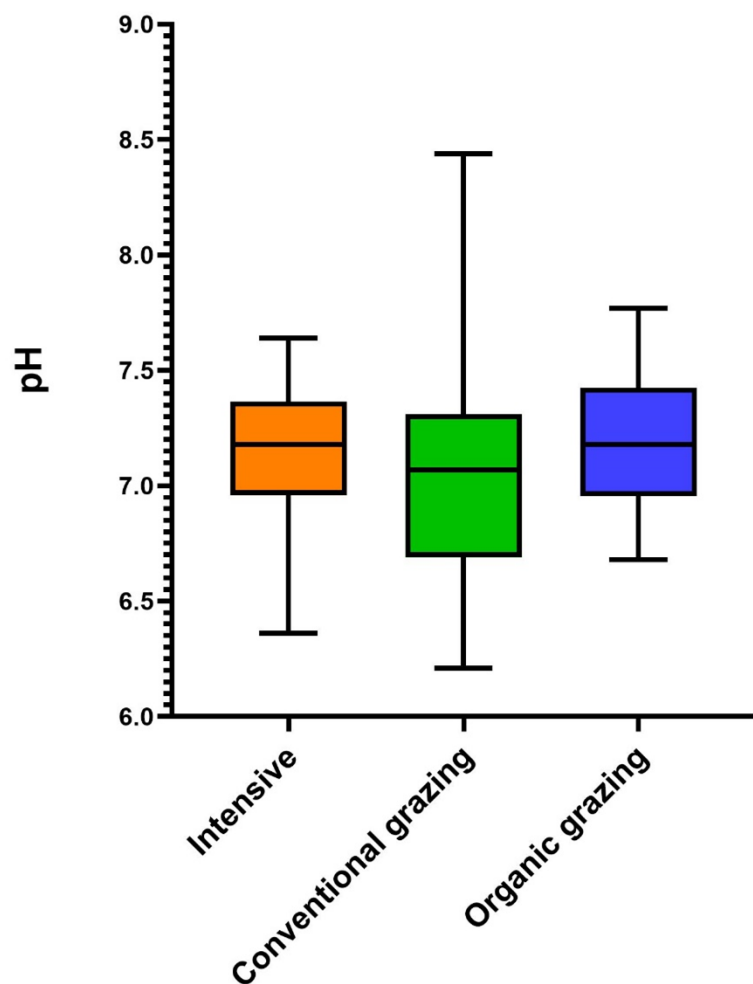


Figure 3. Workflow of the three modes of operation of SqueezeMeta: sequential, co-assembly and merged (Tamames and Puente-Sánchez, 2019)



Intensive; N= 38

Conventional grazing; N= 33

Organic grazing; N= 30

* $p < 0,05$

** $p < 0,01$



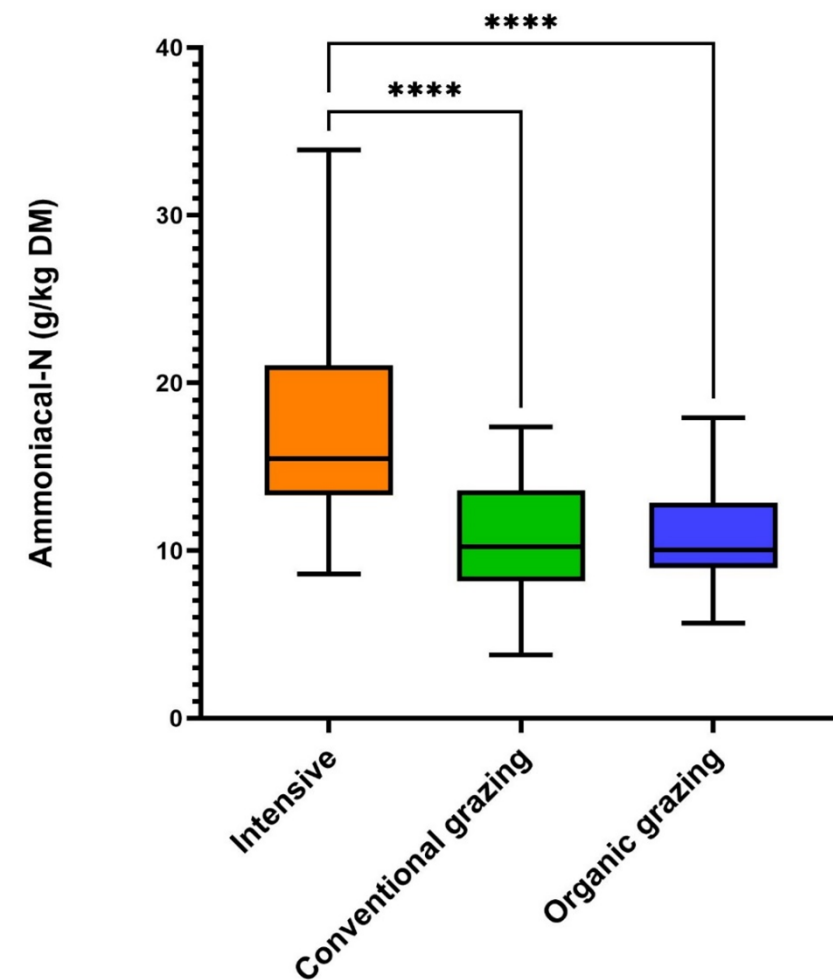
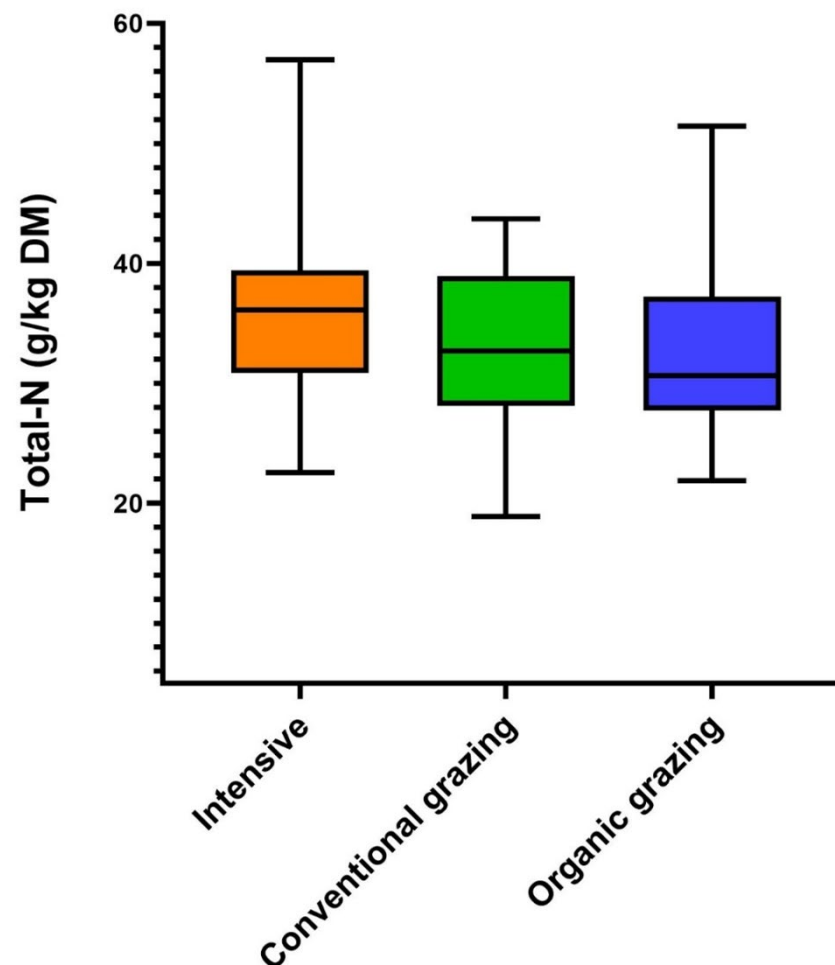
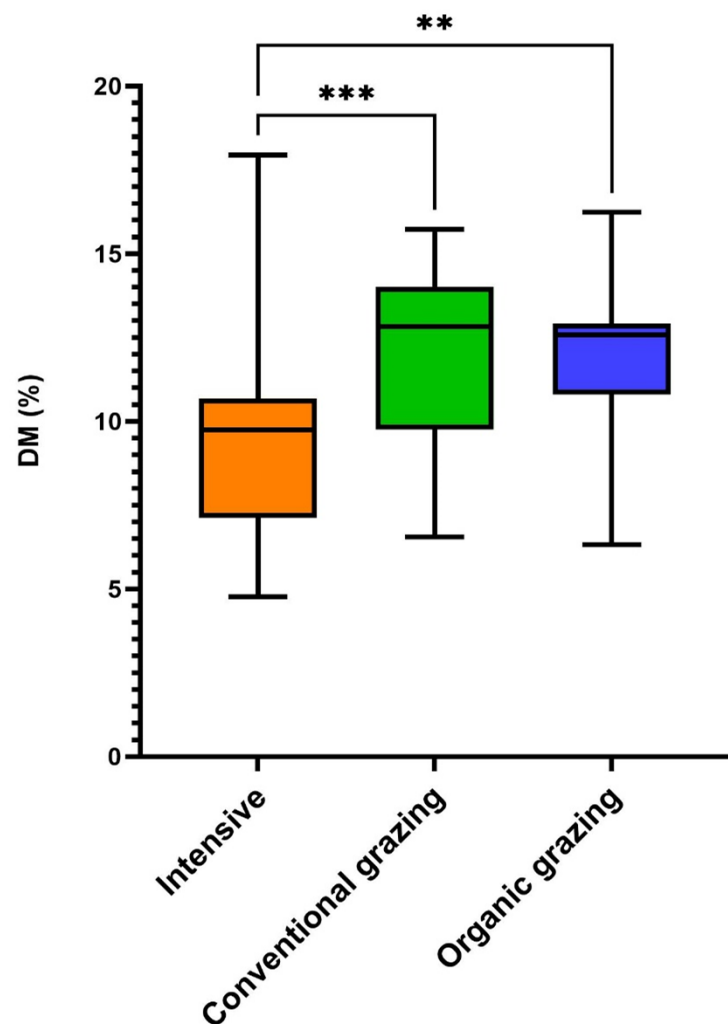
Introduction

Problematic

Objective

Methodology

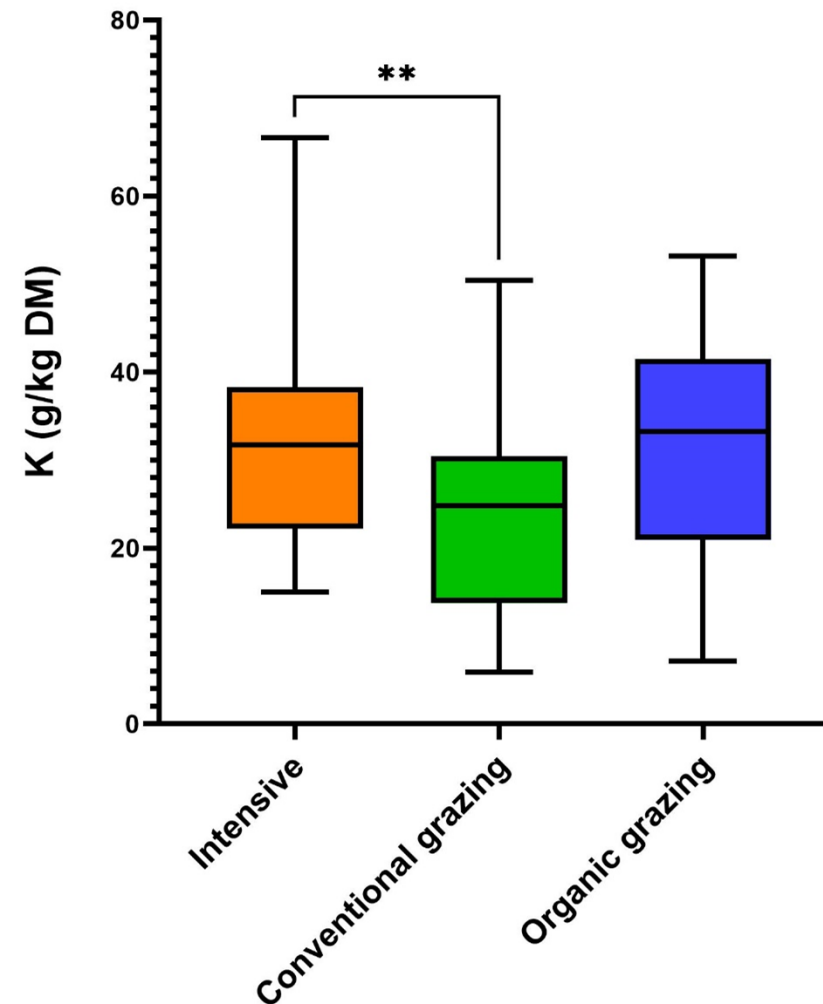
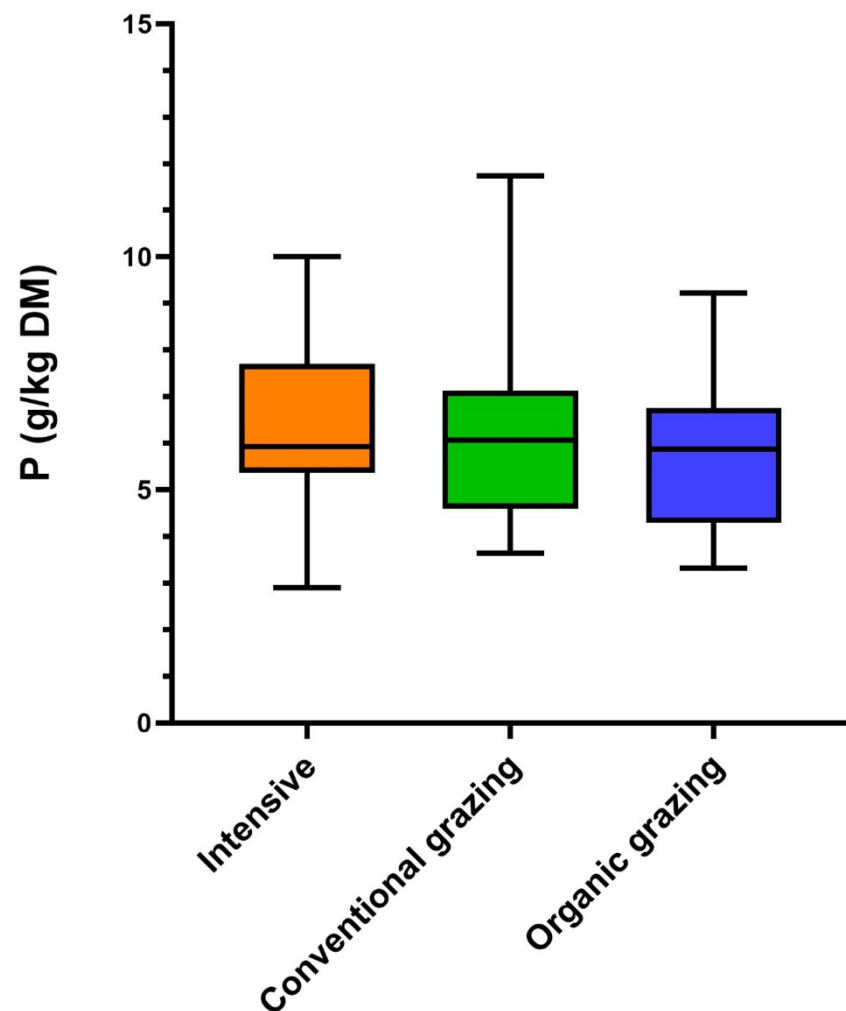
Results



Intensive; N= 38
Conventional grazing; N= 33
Organic grazing; N= 30

** p<0,01
*** p<0,001
**** p<0,0001





Intensive; N= 38
Conventional grazing; N= 33
Organic grazing; N= 30

** p<0,01



Sequences characteristics of all samples (N=101)

Parameters	Average	Standard deviation
Mean read length (bp)	2,797	1,061
Mean read quality	13.31	0.21
Median read length (bp)	1,252	844
Median read quality of reads	14.30	0.27
Mean number of reads	319,894	298,769
Mean read length N50 (bp)	6,792	1,089
Mean of total bases (Gb)	7,80	6,80

- Long reads with a mean length of 2,797 bp and a median read quality >13
- Super accuracy basecalling mode (guppy version 6.4.8)



#----- Statistics on contigs

Assembly

Number of contigs 276,472

Total length 4,364,454,890

Longest contig 1,431,698

Shortest contig 200

N50 29,031

N90 7,323

Contigs at superkingdom (k) rank 247,395 (89.5%), in 4 superkingdoms

Contigs at phylum (p) rank 221,583 (80.1%), in 76 phyla

Contigs at class (c) rank 186,684 (67.5%), in 81 classes

Contigs at order (o) rank 138,371 (50.0%), in 135 orders

Contigs at family (f) rank 78,734 (28.5%), in 238 families

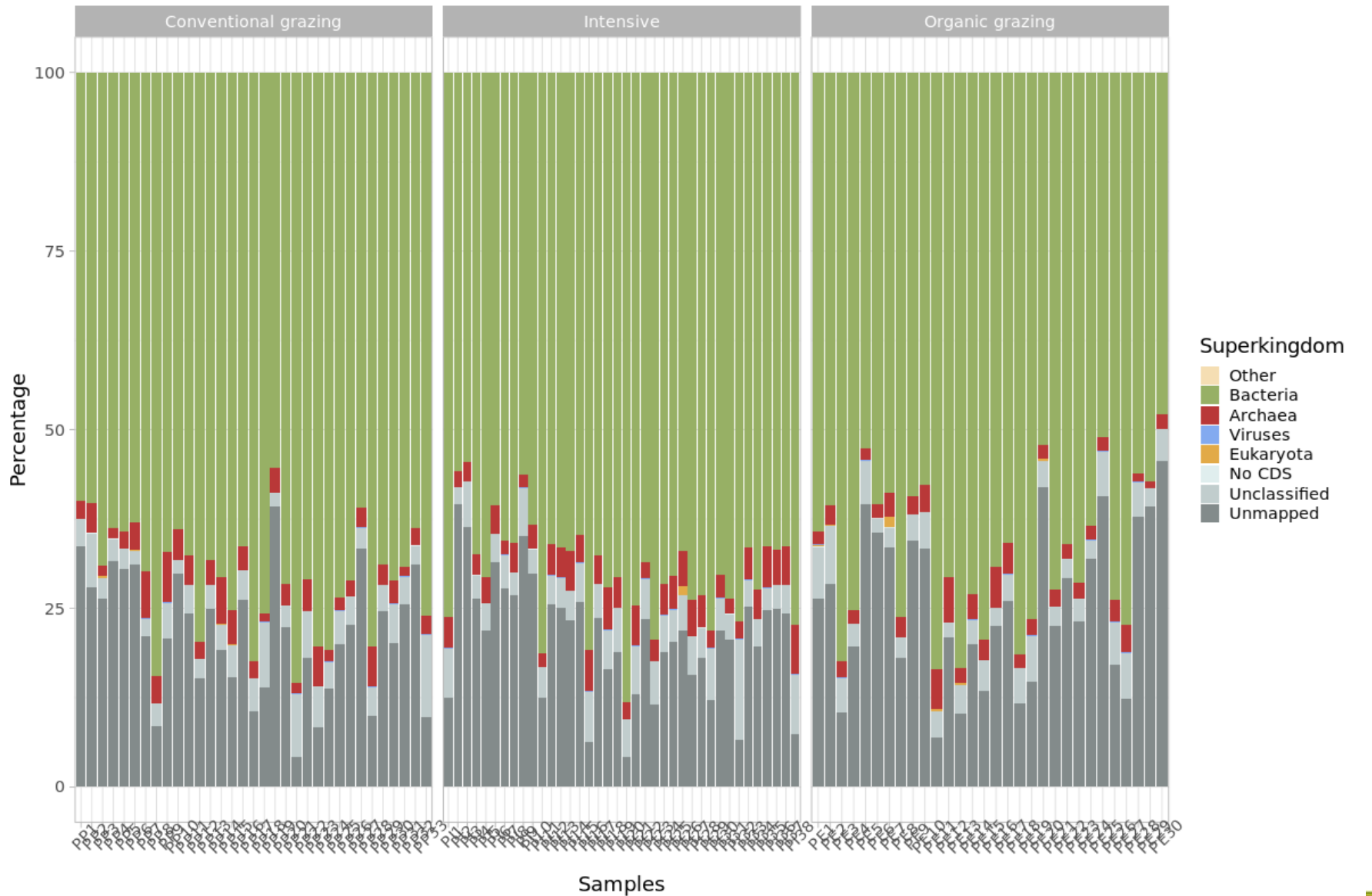
Contigs at genus (g) rank 35,193 (12.7%), in 531 genera

Contigs at species (s) rank 26,439 (9.6%), in 667 species

Congruent 222,199 (80.4%)

Disparity >0 54,274 (19.6%)

Disparity >= 0.25 13,230 (4.8%)



73% classified and mapped:

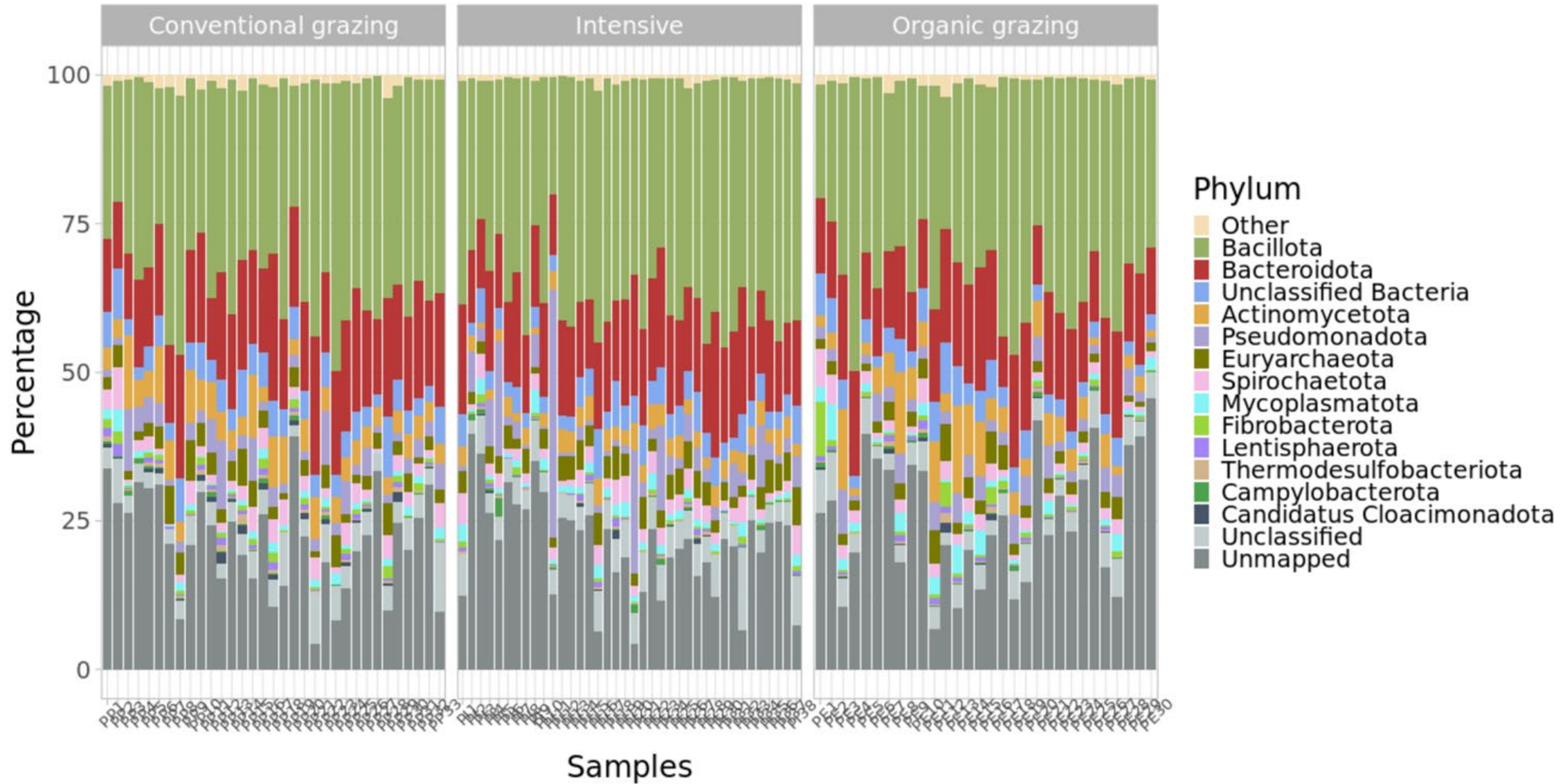
Archaea:4.61%

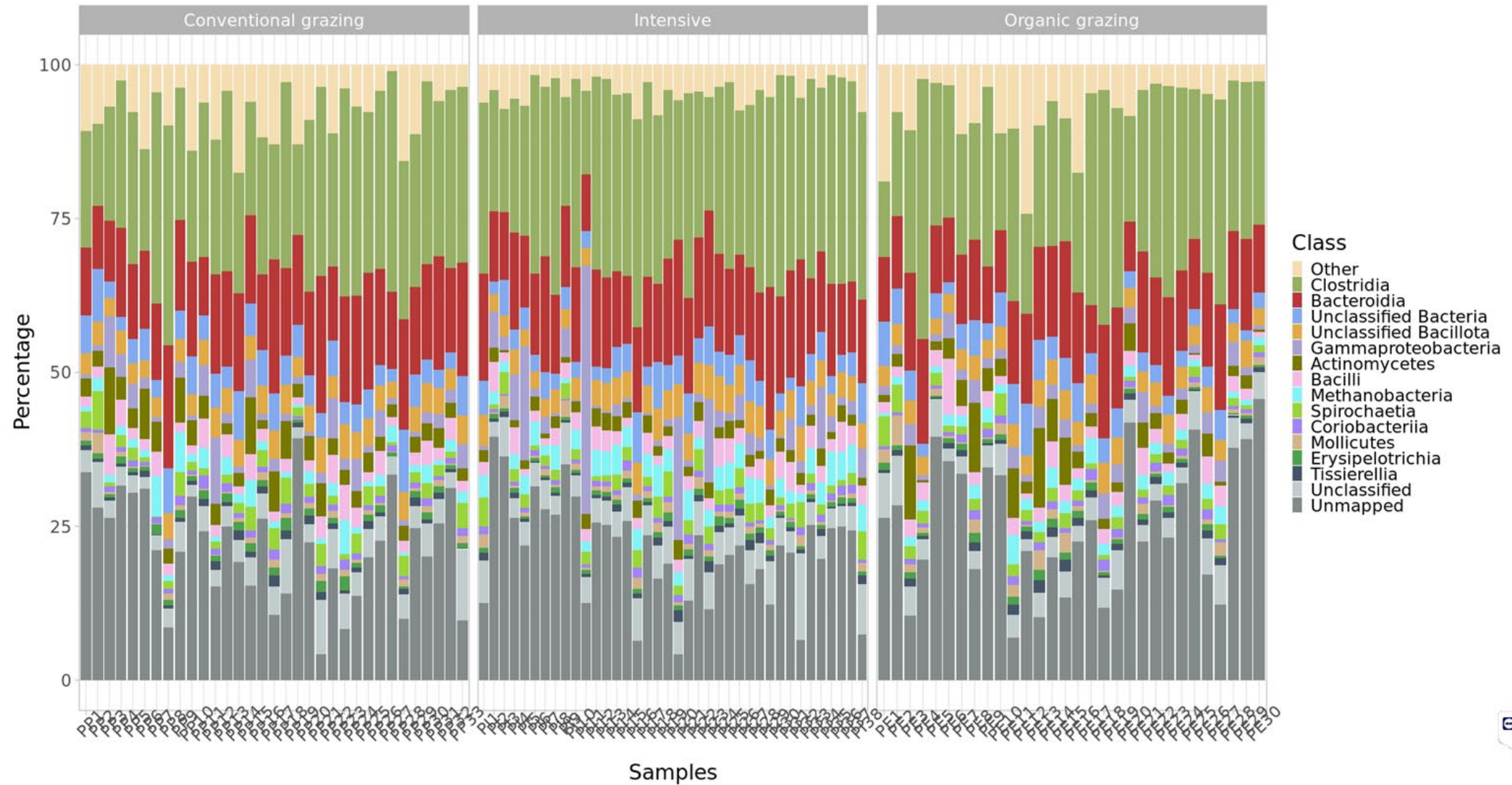
Bacteria: 95.22%

Eukaryota:0.07

Viruses:0.09







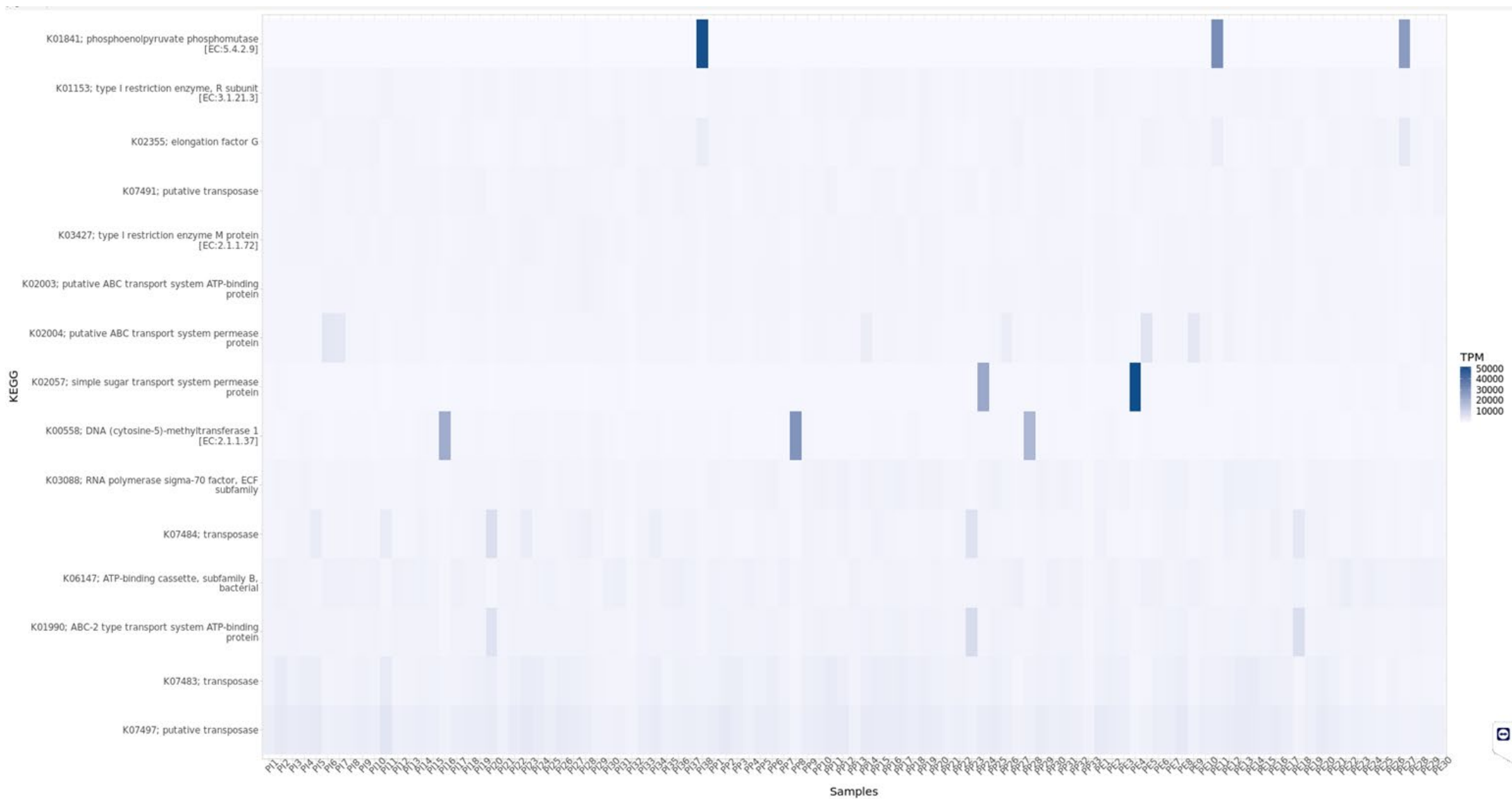
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Conclusions

- Physicochemical analyses showed that there are significant differences for the slurry between the three different livestock production systems regarding EC, TDS, DM, ammoniacal-N and K;
- The preliminary results of coassembly mode of slurry samples showed that 73% of microorganisms were classified and mapped and mainly formed by bacteria (95%) and the most abundant phylum are Bacillota, Bacteroidota and unclassified Bacteria.
- The sequential mode will be used in the further analysis to compare results with coassembly mode.

Acknowledgments



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Thank you for your attention

