

monoguthealth

Optimal gut function in monogastric livestock

Glutamine supplementation during the suckling period and its influence on piglet growth and intestinal metabolism

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Challenges of modern pig production.

Environmental impact (NH_3 , N_2O , CO_2 , CH_4 direct and indirect (50 – 70%).

Animal welfare (farrowing crates, lack of space).²

Negative public perception of mass production pig farming (European Livestock Voice).¹

Increased production costs (feed, energy, staff).

Loss of breed diversity (95% of modern production is just a few breeds) – bred for productivity.¹

Increased litter sizes.



- ✓ Has decreased the number of litters required to produce 1000 weaned piglets from **126** in 2009 to **80** in 2021²
- ❖ Has led to increased numbers of low birthweight piglets, which have a...²
- ❖ ...Higher pre-weaning mortality²

Low birthweight piglets – definition and problems.

Definition: There is no commonly agreed definition for what constitutes a low birthweight piglet. Researchers have used birthweight cutoffs or individual / group litter birthweight variation as methods to define and identify low birthweight piglets in their studies.

- Slower growth:^{5,6} ✓

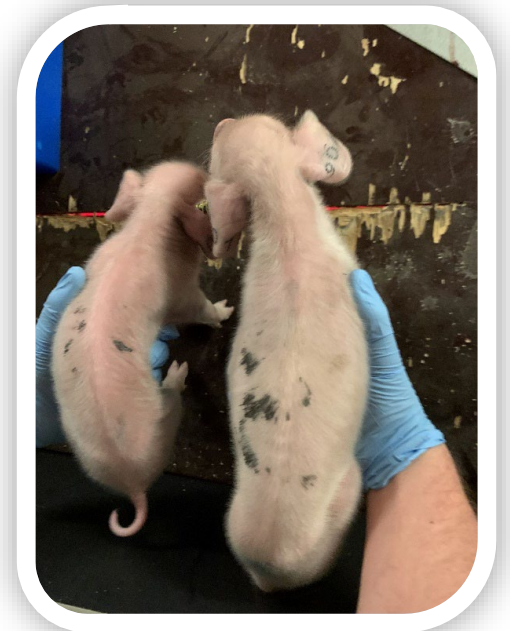
a. Impaired muscle development and subsequent lower carcass quality.

- Altered metabolism:^{7,8}

a. Impaired pre-weaning glucose tolerance – led to the identification of *myo-inositol* as a biomarker of low birthweight in pigs

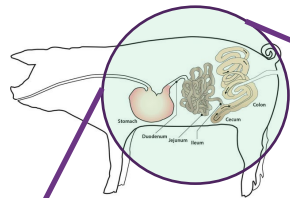
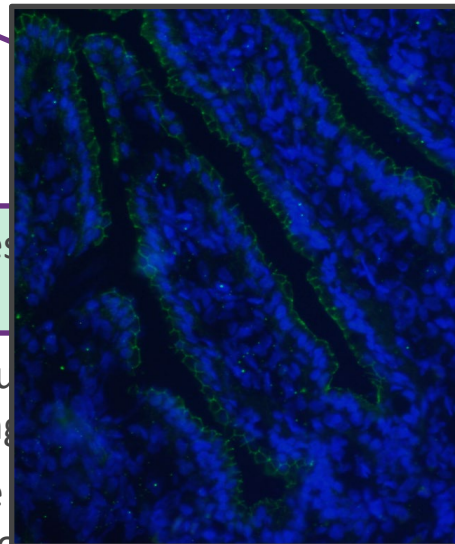
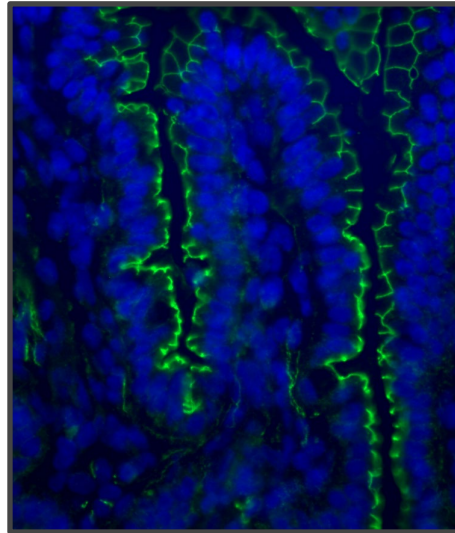
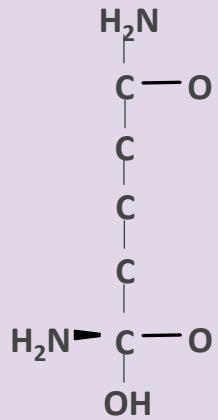
- Impaired gastrointestinal development:^{9,10} ✓

a. Impaired pre-weaning intestinal structure and lower enzymatic activity
b. Dysfunctional intestinal mitochondrial function and antioxidative defense at weaning.



✓ The amino acid Glutamine has been shown to improve the growth of pre-and post-weaning piglets and intestinal development in post-weaning piglets¹⁰⁻¹².

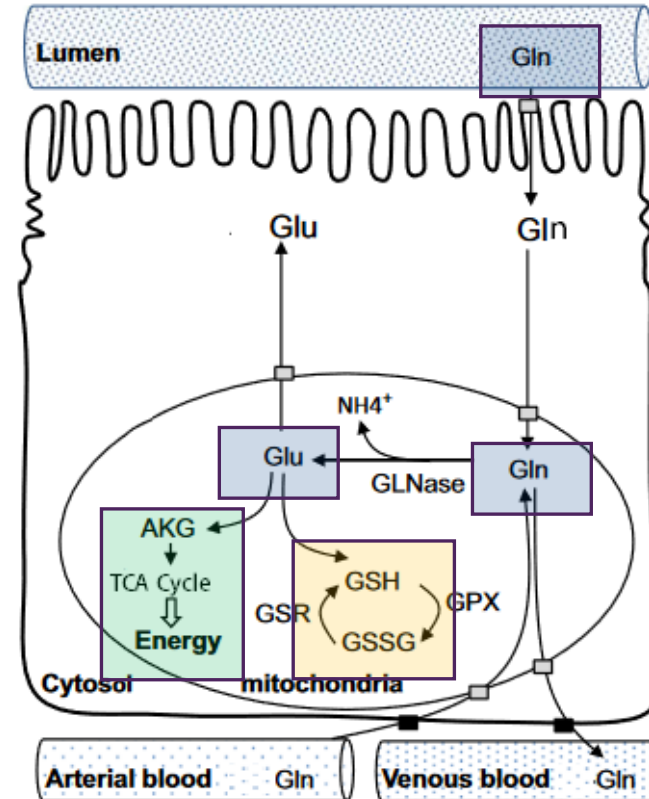
Glutamine – more than just an amino acid



In pigs the gastrointes rapidly after birth:

- The weight of the jejun almost doubles during
- The cells that line the (enterocytes) undergo rapid development

• Image 1



- Nutrient uptake
- Secretion of enzymes and immune molecules
- Passive barrier function

Glutamine (Gln) is the primary fuel for enterocytes of the small intestine¹³

Glutamine is taken up by the enterocyte, enters the mitochondria and is converted to Glutamate (Glu)

Glutamate is converted to Alpha-KetoGlutarate (AKG), enters the TCA cycle and used to generate ATP (energy)

Glutamate can also be converted to Glutathione (GSH), an antioxidant

Previous glutamine supplementation research in pigs

Most studies investigating glutamines effect on growth and development in pigs, have focused on the weaning period, results have reported that glutamine:



- Increased bodyweight
- Increased daily bodyweight gain
- Protected against intestinal atrophy by improving villus height and crypt depth
- Reduced intestinal permeability
- Prevented some negative metabolic changes associated with weaning¹⁶

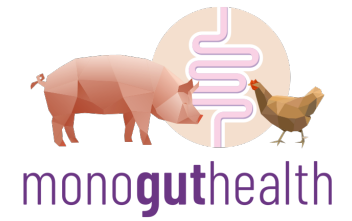
Glutamine supplementation studies in pre-weaning pigs have shown that:



- Increased bodyweight, when supplemented from 7 to 21 days of age¹⁸
- Protected against intestinal atrophy by improving villus height¹⁸
- Increased daily bodyweight gain when supplemented from birth to 21 days age¹⁰



The Q (Glutamine)-PIG1 Project - (2016 – 2020)



Hypothesis:

Oral glutamine supplementation during the early suckling period improves the bodyweight of low birthweight piglets, and is associated with changes in small intestine morphology/development and metabolism.

Aims:

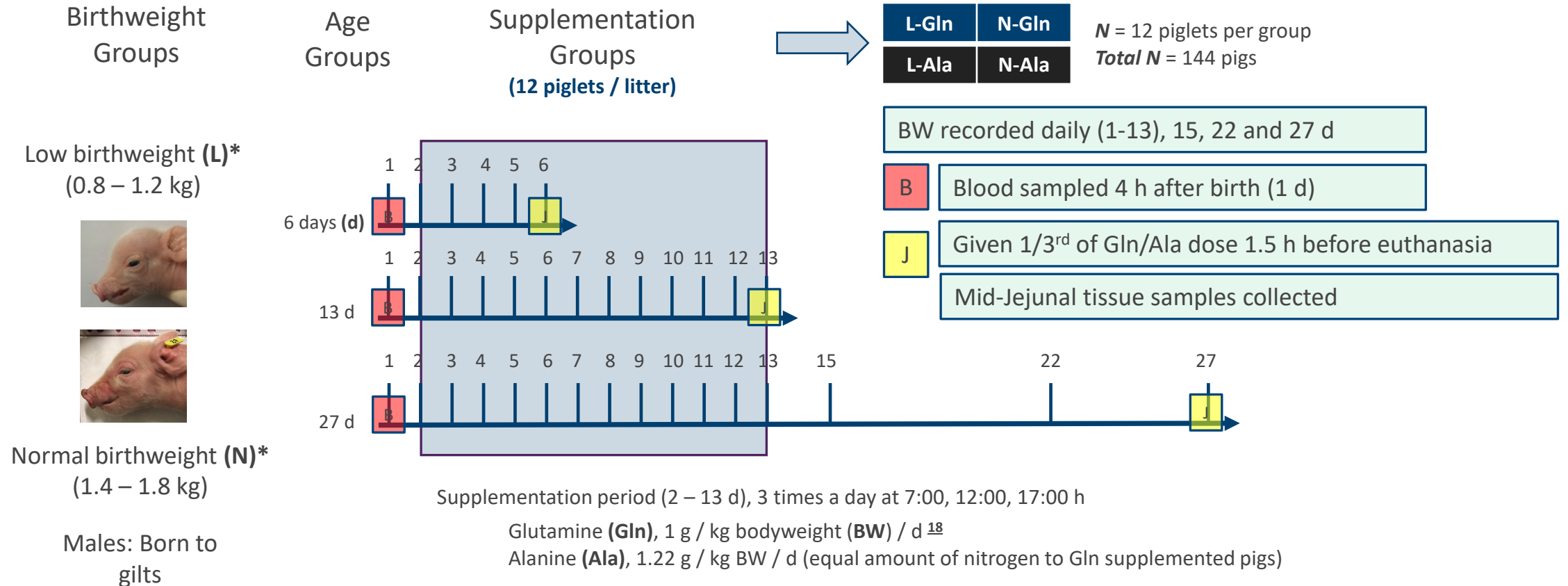
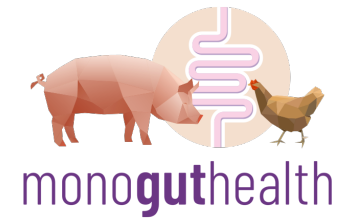
To determine if oral glutamine supplementation of low birthweight piglets, during the early suckling period is associated with:

1. Improved bodyweight,
2. Morphological / developmental changes of the small intestine,
3. Changes in glutathione synthesis.

To determine if the changes observed from 1-3 are still present after supplementation has stopped.



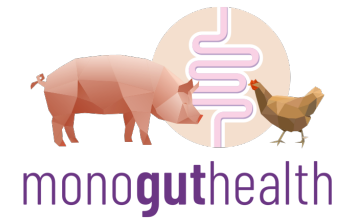
The Q-PIG1 Project – Trial design



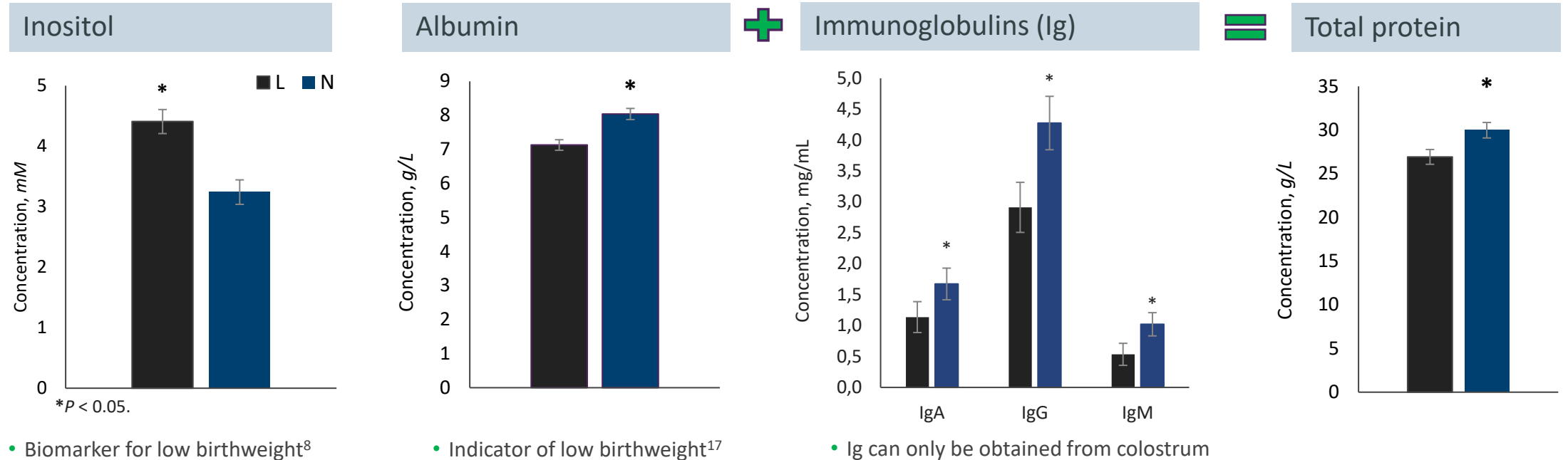
*(**L**), below the lowest birthweight quartile, (**N**); represents the middle 50th percentile, of the experimental pig facility



The Q-PIG1 Project - Results



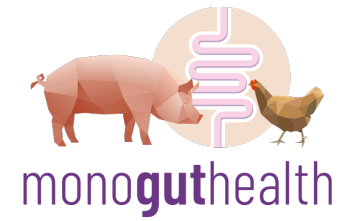
Plasma metabolites at 4 hours after birth



Taken together, these results provide strong evidence that the low birthweight piglets selected for this study are low birthweight

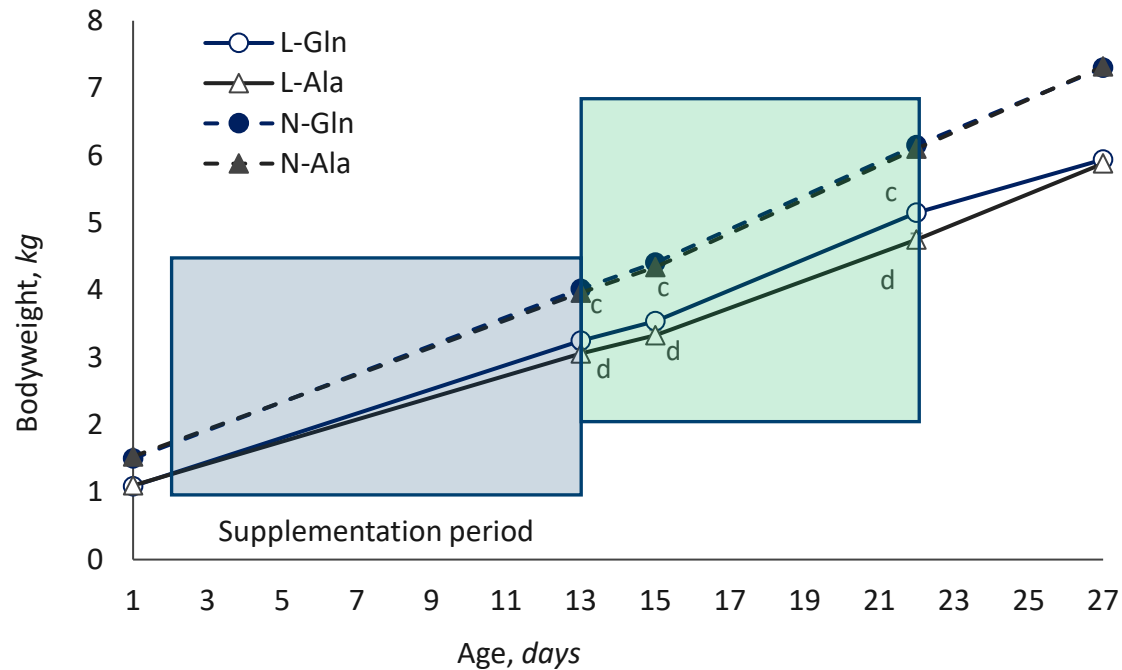


The Q-PIG1 Project - Results



Piglet growth^{11,12}

Bodyweight



^{a,b}Different from N piglets within Supplementation group ($P < 0.05$).

^{c,d}Different from Ala piglets within Birthweight group ($P < 0.05$).

Average Daily Gain

Period (g/d)	L-Gln	L-Ala	N-Gln	N-Ala	SEM
1 - 6 d	129 ^b	122 ^b	156 ^a	156 ^a	7.90
7 - 13 d	179 ^b	165 ^b	204 ^a	200 ^a	8.36
14 - 27 d	187 ^b	186 ^b	224 ^a	223 ^a	10.9

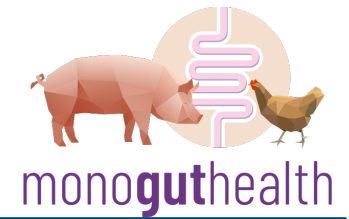
- Bodyweight and Average Daily Gain were always higher in N then L piglets

- At ages 13, 15 and 22 d L-Gln piglets were heavier than L-Ala

- ✓ Oral supplementation of Gln is associated with increased bodyweight in L piglets and this effect persists beyond the supplementation period

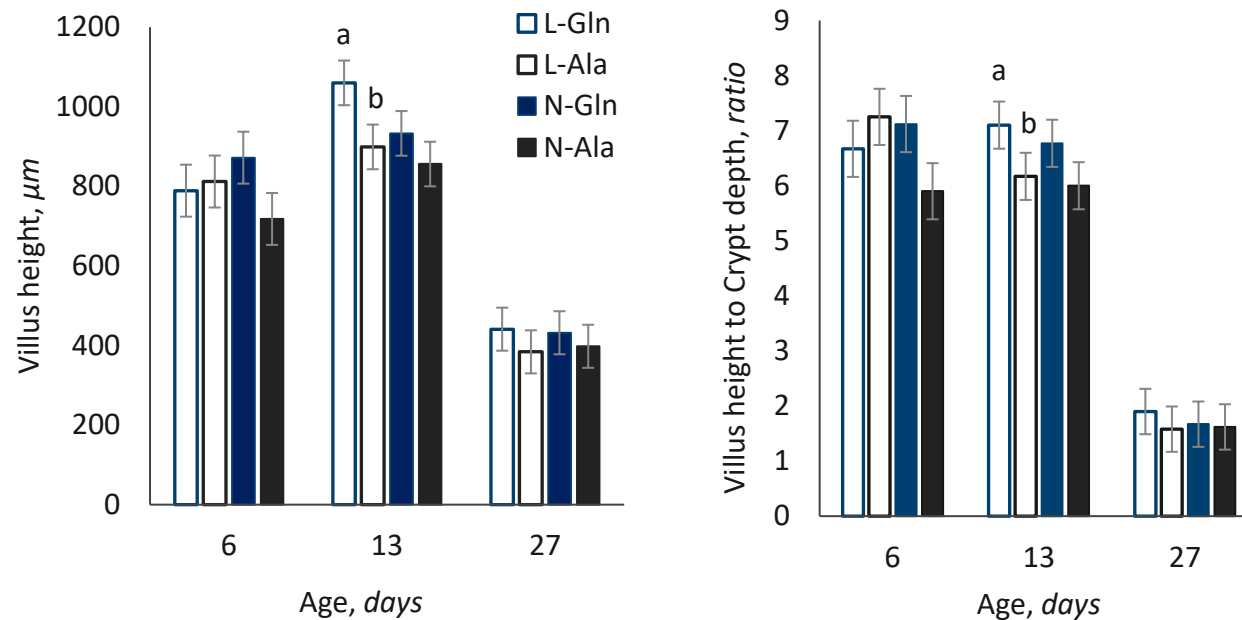


The Q-PIG1 Project - Results



Jejunal parameters¹²

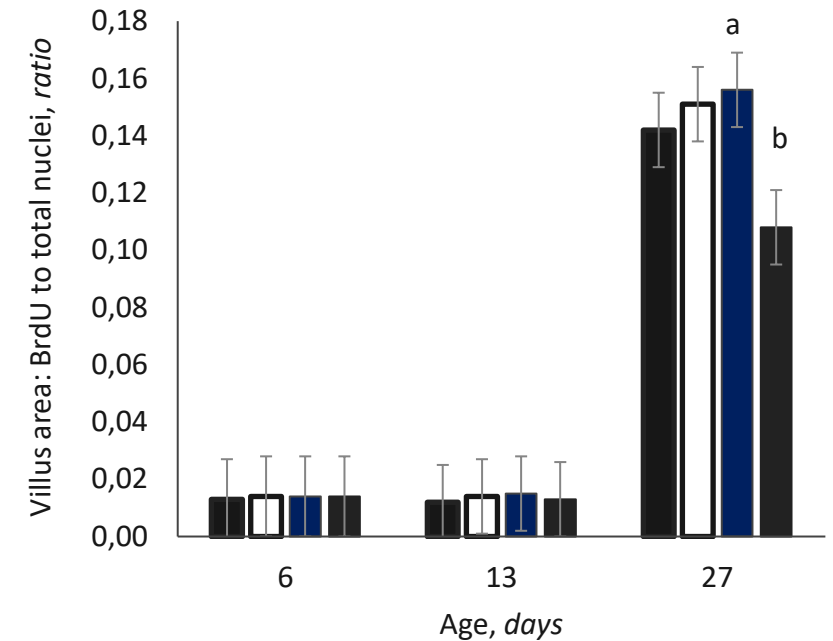
Villus height and Villus height to crypt depth ratio



^{a,b}Different from Ala piglets within Birthweight group ($P < 0.05$).

Jejunal glutathione concentrations were measured.
No differences were observed.

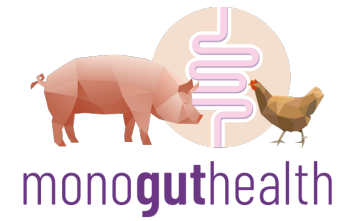
Cell proliferation



These results suggest increased jejunal digestive and absorptive capabilities in low birthweight male piglets supplemented with glutamine compared to low birthweight alanine piglets



The Q-PIG1 Project - Conclusions



Oral glutamine supplementation (1g/kg BW/day) from 2 – 13 days of life to male low birthweight piglets:

- ✓ Improved bodyweight from 13 – 22 days of life compared to low birthweight controls
- ✓ Improved jejunal digestive and absorptive capabilities at 13 days of life

Open questions:

How does glutamine improve low birthweight piglet bodyweight?

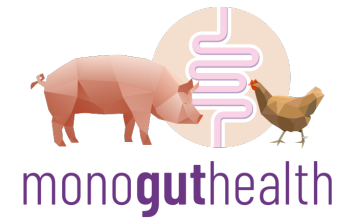
What would happen if piglets were supplemented longer?

What if controls were supplemented with water?

What if a more proximal section of the small intestine was analyzed?



MonoGutHealth Project – (2021 – 2023)



Hypothesis:

Oral glutamine supplementation during the early suckling period improves the bodyweight of low birthweight piglets, and is associated with changes in glutamine and/or glucose metabolism.

Aims:

To determine if oral glutamine supplementation of low birthweight piglets, during the early suckling period is associated with:

1. Improved bodyweight,
2. Changes in small intestine glutamine or glucose metabolism,
3. Changes in small intestine development.



Catheter study: Study design

Birthweight
Groups

Low birthweight (**L**)*
(0.8 – 1.2 kg)



Normal birthweight (**N**)*
(1.5 – 1.9 kg)

Males

Born to parity 2-9 sows

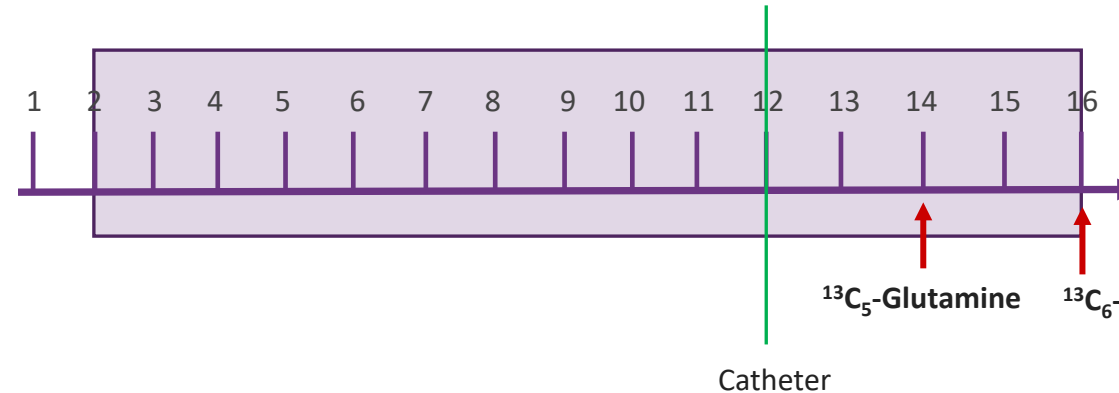
Supplementation
Groups
(14 piglets per litter)

Glutamine (**Gln**), 1 g / kg bodyweight (**BW**) / d
Water (**W**) (equal volume to Gln supplemented pigs)



L-Gln	N-Gln
L-W	N-W

N = 12 piglets per group
Total N = 48 pigs



BW recorded daily

1h separation from sow

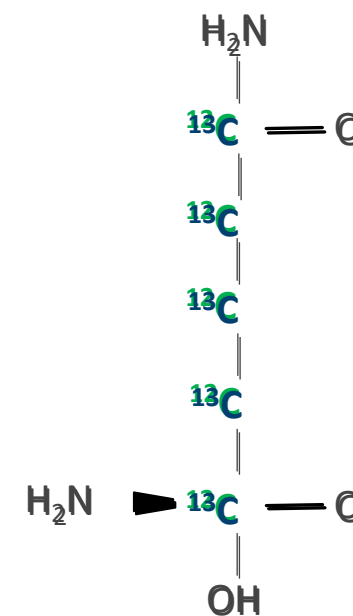
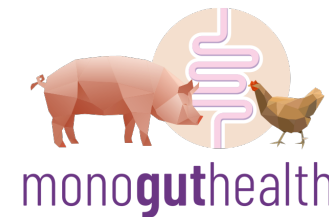
B Every 30 min for 5 hours

Stay with litter during
sampling (can suckle)

Supplementation period (2 – 16 d), 3 times a day at 7:00, 12:00, 17:00 h

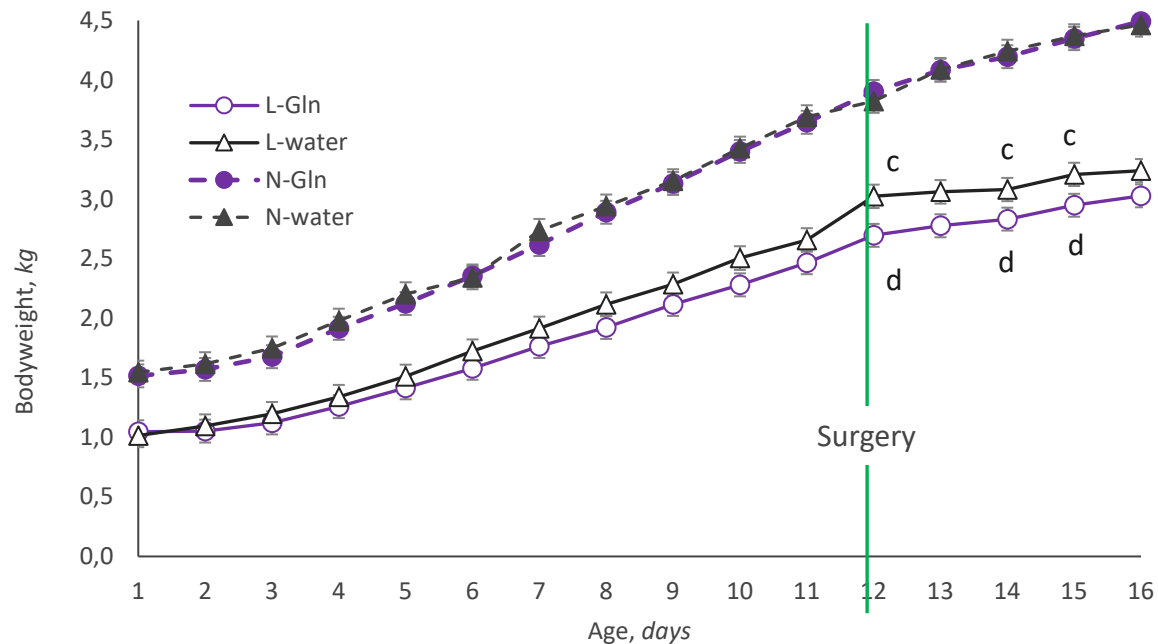
***(L)**, below the lowest birthweight quartile, **(N)**; represents the middle 50th percentile, of the experimental pig facility

Stable isotope tracers: Where are the carbons?



Catheter study: Piglet growth

Bodyweight



^{a,b}Different from N piglets within Supplementation group ($P < 0.05$).

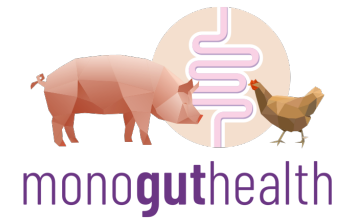
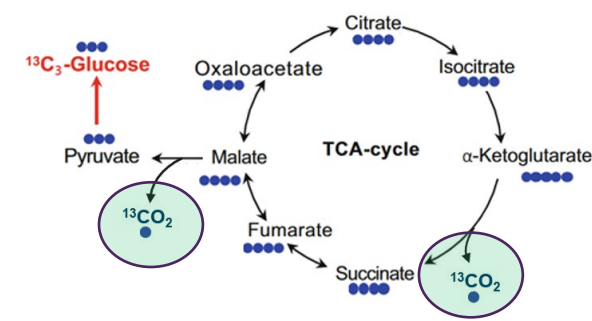
^{c,d}Different from Water piglets within Birthweight group ($P < 0.05$).

Average Daily Gain

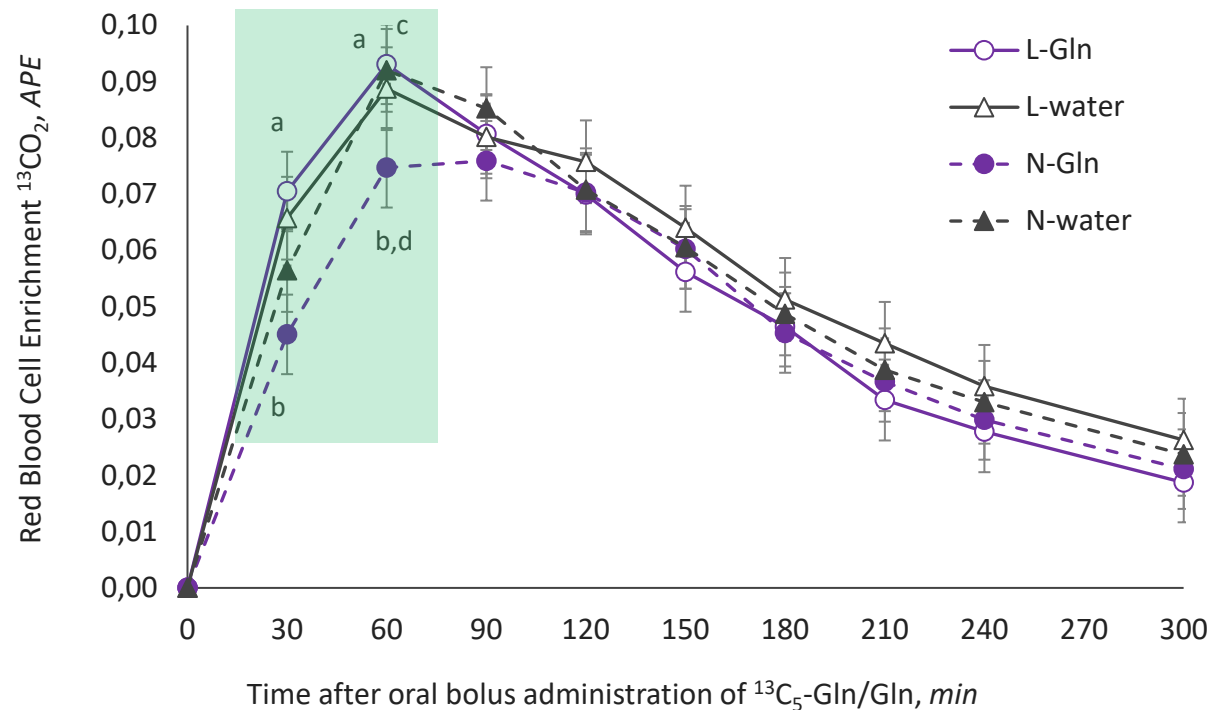
(g/d)	Period	L-Gln	L-water	N-Gln	N-water
Total	1 - 16 d	133 ± 9 ^b	144 ± 9 ^b	211 ± 9 ^a	190 ± 9 ^a
Pre-Surgery	1 - 12 d	158 ± 13 ^b	180 ± 13.4 ^b	222 ± 12.6 ^a	206 ± 13.4 ^a
Post-Surgery	13 - 16 d	25.2 ± 9 ^b	19.8 ± 9 ^b	39.3 ± 9 ^a	42.4 ± 9 ^a

- Bodyweight and Average Daily Gain were always higher in N then L piglets
- On the day of surgery (12), 14 and 15 days L-water piglets were heavier than L-Gln
- Surgery negatively impacted piglet bodyweight and average daily gain

Catheter study: $^{13}\text{C}_5$ -Glutamine metabolism



^{13}C -Carbon dioxide enrichment in Red Blood Cells



^{a,b}Different from N piglets within Supplementation group ($P < 0.05$).

^{c,d}Different from Water piglets within Birthweight group ($P < 0.05$).

Kinetic parameters

	L-Gln	L-water	N-Gln	N-water
AUC, APE×min	15.5 ± 1.66	16.7 ± 1.73	14.3 ± 1.66	15.8 ± 1.73
E _{max} , min	0.100 ± 0.01	0.09 ± 0.01	0.08 ± 0.01	0.09 ± 0.01
T _{max} , min	62.9 ± 5.54 ^b	75.1 ± 5.83	80.3 ± 5.54 ^a	65.0 ± 5.83

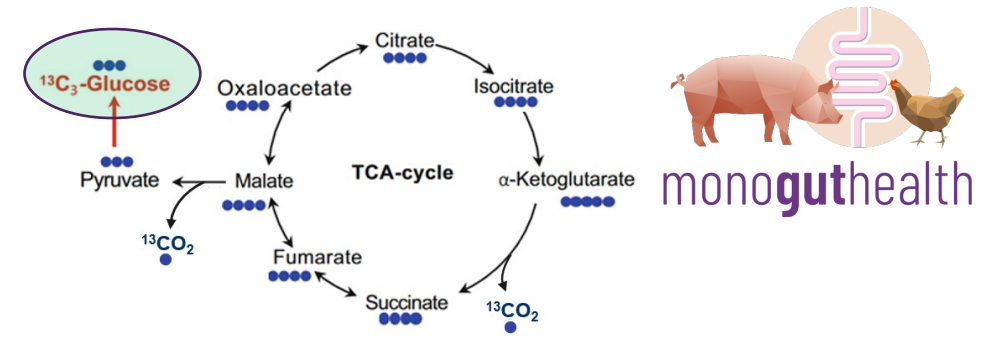
- N-Gln had a higher T_{max} than L-Gln

- N-Gln RBC $^{13}\text{CO}_2$ enrichment is lower than:

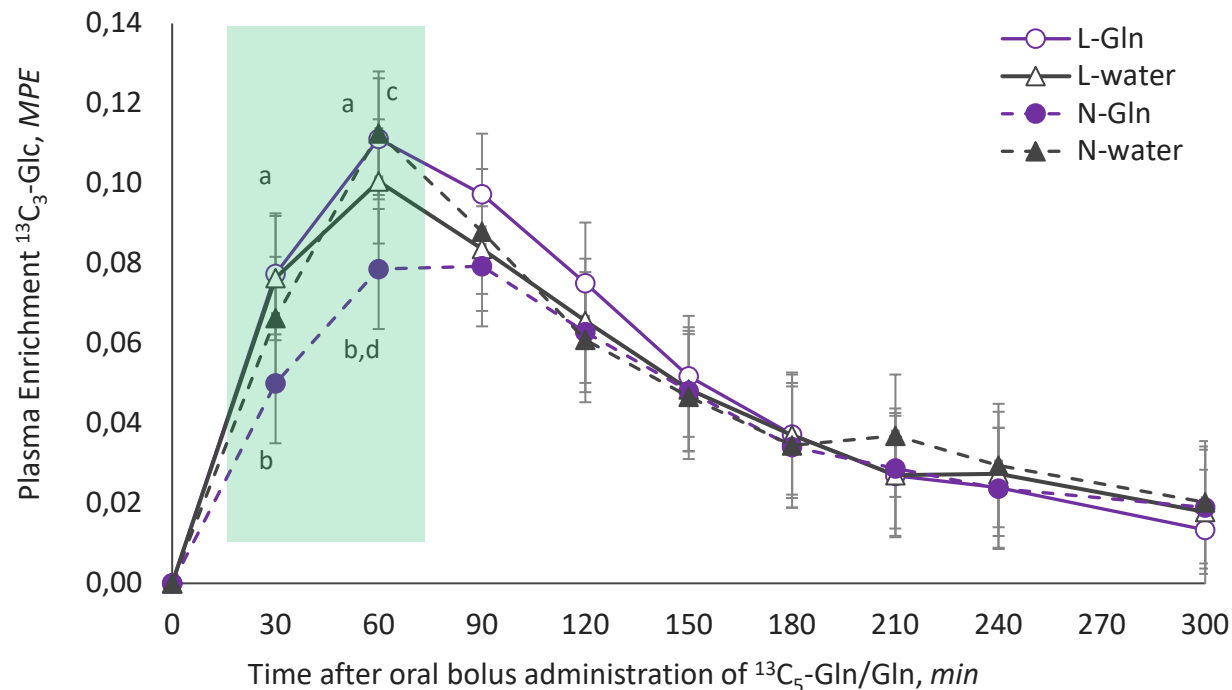
- L-Gln at 30, 60 min
- N-water at 60 min



Catheter study: $^{13}\text{C}_5$ -Glutamine metabolism



$^{13}\text{C}_3$ -Glucose enrichment in Plasma



^{a,b}Different from N piglets within Supplementation group ($P < 0.05$).

^{c,d}Different from Water piglets within Birthweight group ($P < 0.05$).

Kinetic parameters

	L-Gln	L-water	N-Gln	N-water
AUC, $\text{MPE} \times \text{min}$	15.4 ± 3.47	14.9 ± 3.53	12.9 ± 3.42	13.4 ± 3.60
E_{max} , min	0.118 ± 0.03	0.108 ± 0.03	0.086 ± 0.03	0.125 ± 0.03
T_{max} , min	59.6 ± 5.00^b	63.8 ± 5.34	73.6 ± 4.80^a	63.3 ± 5.09

- N-Gln had a higher T_{max} than L-Gln

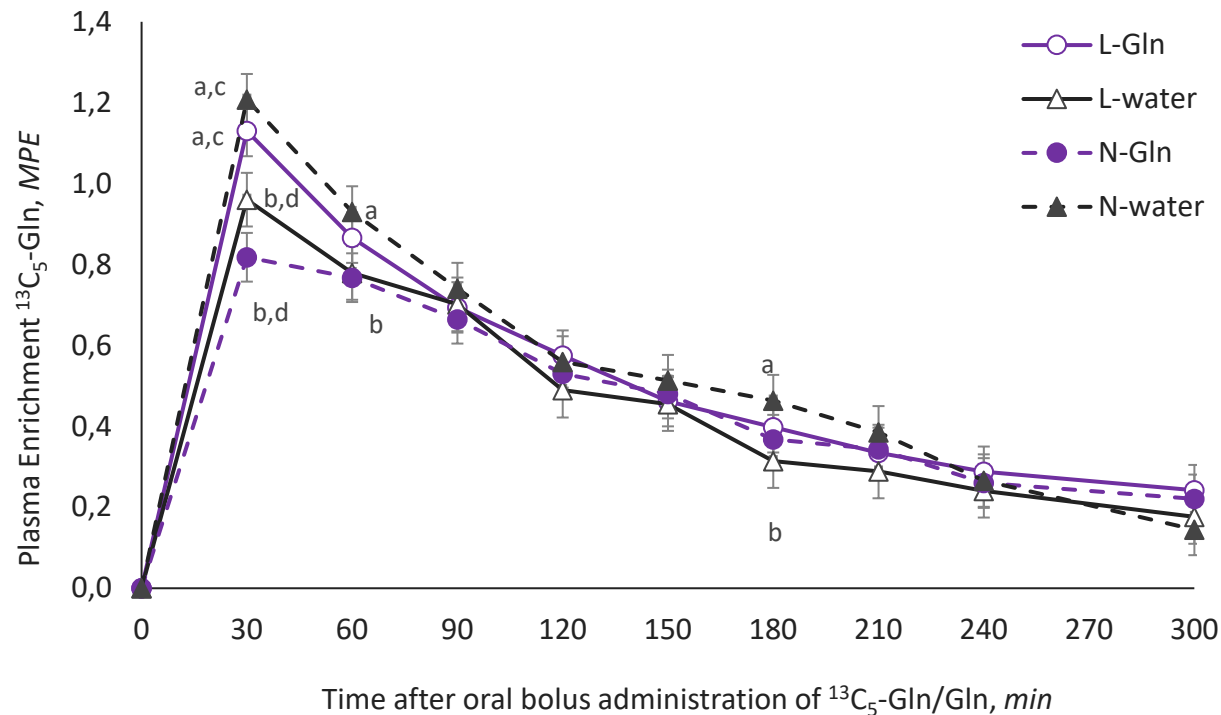
- N-Gln Plasma $^{13}\text{C}_3$ -Glucose enrichment is lower than:

—○— L-Gln at 30, 60 min

-▲- N-water at 60 min

Catheter study: $^{13}\text{C}_5$ -Glutamine metabolism

$^{13}\text{C}_5$ -Glutamine enrichment in Plasma



^{a,b}Different from N piglets within Supplementation group ($P < 0.05$).

^{c,d}Different from Water piglets within Birthweight group ($P < 0.05$).

Kinetic parameters

	L-Gln	L-water	N-Gln	N-water
AUC, $\text{MPE} \times \text{min}$	161 ± 14.3	141 ± 15.3	142 ± 13.8	171 ± 14.7
$E_{\text{max}}, \text{min}$	1.16 ± 0.09 ^a	0.983 ± 0.100 ^b	0.893 ± 0.086 ^{b,d}	1.24 ± 0.091 ^{a,c}
$T_{\text{max}}, \text{min}$	32.4 ± 3.71	36.5 ± 3.97	37.8 ± 3.57	28.3 ± 3.79

- N-Gln had a lower E_{max} than L-Gln and N-Water

- L-water had a lower E_{max} than N-Water

- L-Gln Plasma $^{13}\text{C}_5$ Glutamine enrichment is higher than:

N-Gln and L-water at 30 min

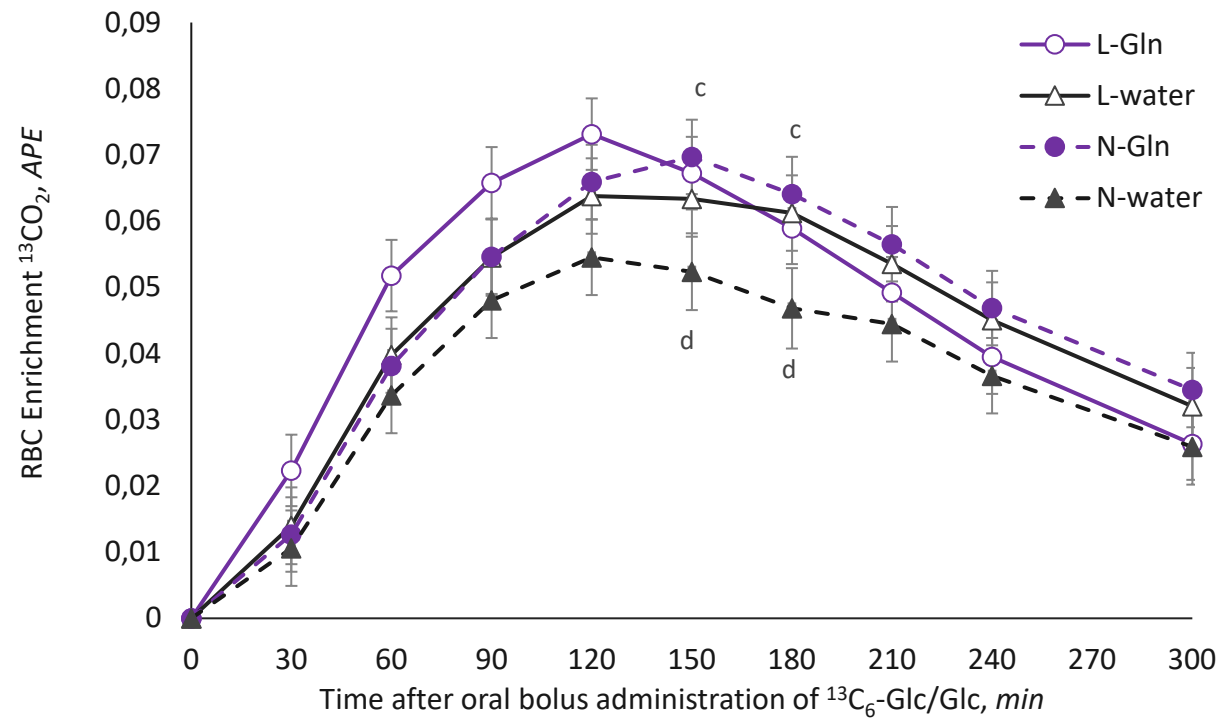
- N-water Plasma $^{13}\text{C}_5$ Glutamine enrichment is higher than:

N-Gln at 30, 60 min

L-water at 30, 60, 180 min

Catheter study: $^{13}\text{C}_6$ -Glucose metabolism

^{13}C -Carbon dioxide enrichment in Red Blood Cells



^{c,d}Different from Water piglets within Birthweight group ($P < 0.05$).

Kinetic parameters

	L-Gln	L-water	N-Gln	N-water
AUC, $\text{APE} \times \text{min}$	14.2 ± 7.80	13.6 ± 7.80	14.1 ± 7.80	11.2 ± 7.80
$E_{\text{max}}, \text{min}$	0.07 ± 0.008	0.07 ± 0.008	0.07 ± 0.008	0.05 ± 0.008
$T_{\text{max}}, \text{min}$	116 ± 8.02^b	125 ± 8.47	143 ± 8.32^a	122 ± 8.47

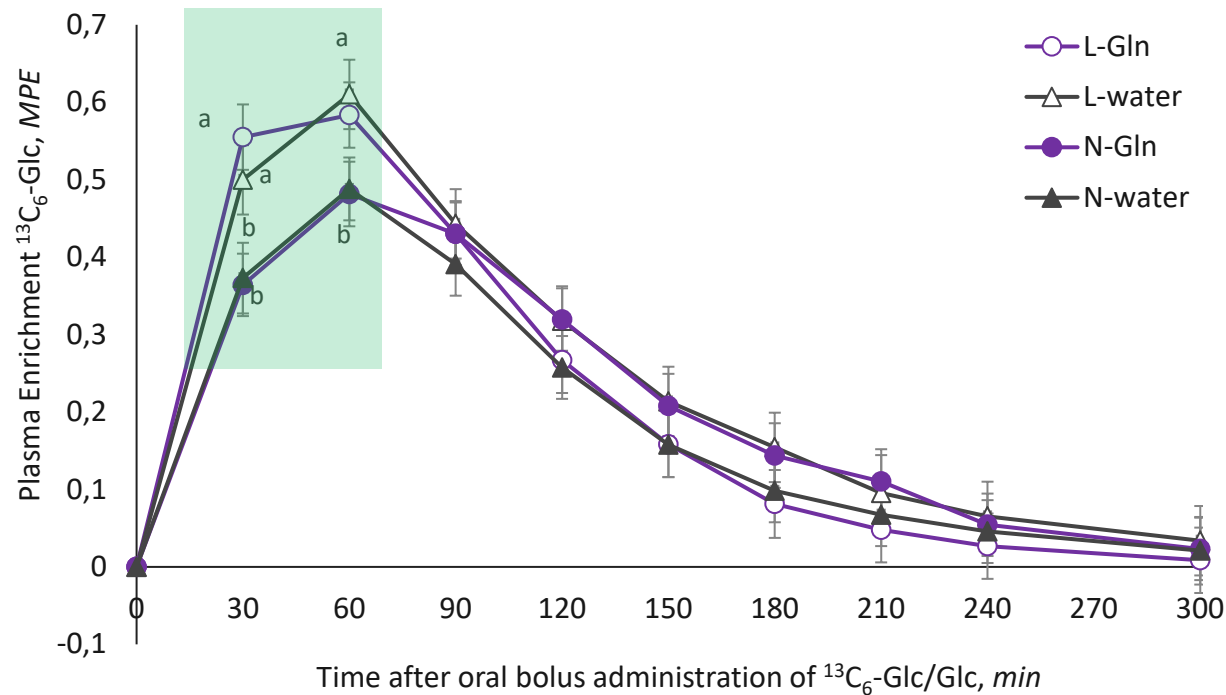
- N-Gln had a higher T_{max} than L-Gln

- N-Gln RBC $^{13}\text{CO}_2$ enrichment is higher than:

—▲— N-water at 150 and 180 min

Catheter study: $^{13}\text{C}_6$ -Glucose metabolism

$^{13}\text{C}_6$ -Glucose enrichment in Plasma



^{a,b}Different from N piglets within Supplementation group ($P < 0.05$).

Kinetic parameters

	L-Gln	L-water	N-Gln	N-water
AUC, $\text{MPE} \times \text{min}$	68.0 ± 7.80	75.4 ± 8.38	68.0 ± 7.50	59.7 ± 7.64
$E_{\text{max}}, \text{min}$	0.701 ± 0.07	0.656 ± 0.08	0.537 ± 0.07	0.558 ± 0.07
$T_{\text{max}}, \text{min}$	47.5 ± 5.28	50.3 ± 5.56	58.1 ± 5.05	50.7 ± 5.08

- No difference in kinetic parameters

- L-Gln Plasma $^{13}\text{C}_6$ -Glucose enrichment is higher than:

N-Gln at 30 min

- L-water Plasma $^{13}\text{C}_6$ -Glucose enrichment is higher than:

N-Water at 30, 60 min

Tissue study: Study design

Birthweight
Groups

Low birthweight (**L**)*
(0.8 – 1.2 kg)



Normal birthweight (**N**)*
(1.5 – 1.9 kg)

Males

Born to parity 2-9 sows

Supplementation
Groups
(14 piglets per litter)



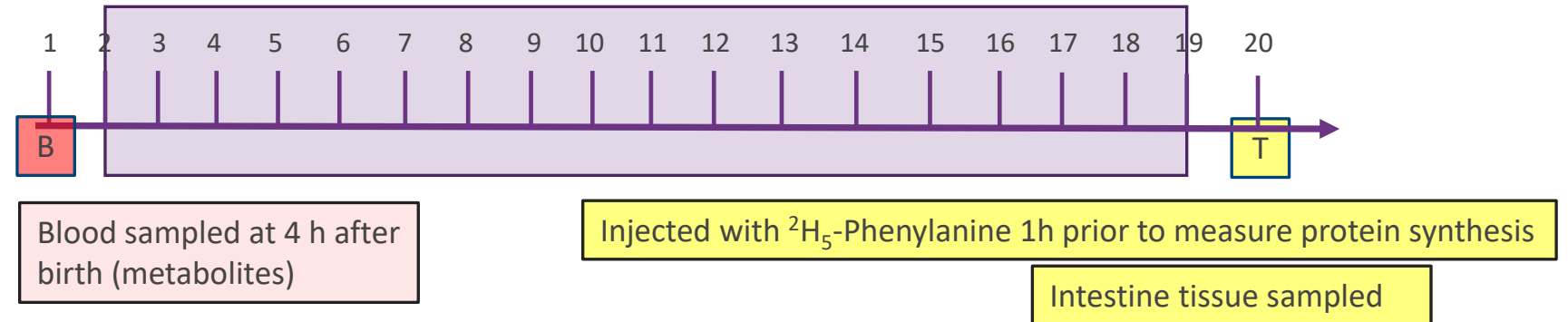
L-Gln	N-Gln
L-W	N-W

N = 12 piglets per group
N-Gln = 14 piglets
Total N = 50 pigs

Glutamine (**Gln**), 1 g / kg bodyweight (**BW**) / d
Water (**W**) (equal volume to Gln supplemented pigs)

Supplementation period (2 – 16 d), 3 times a day at 7:00, 12:00, 17:00 h

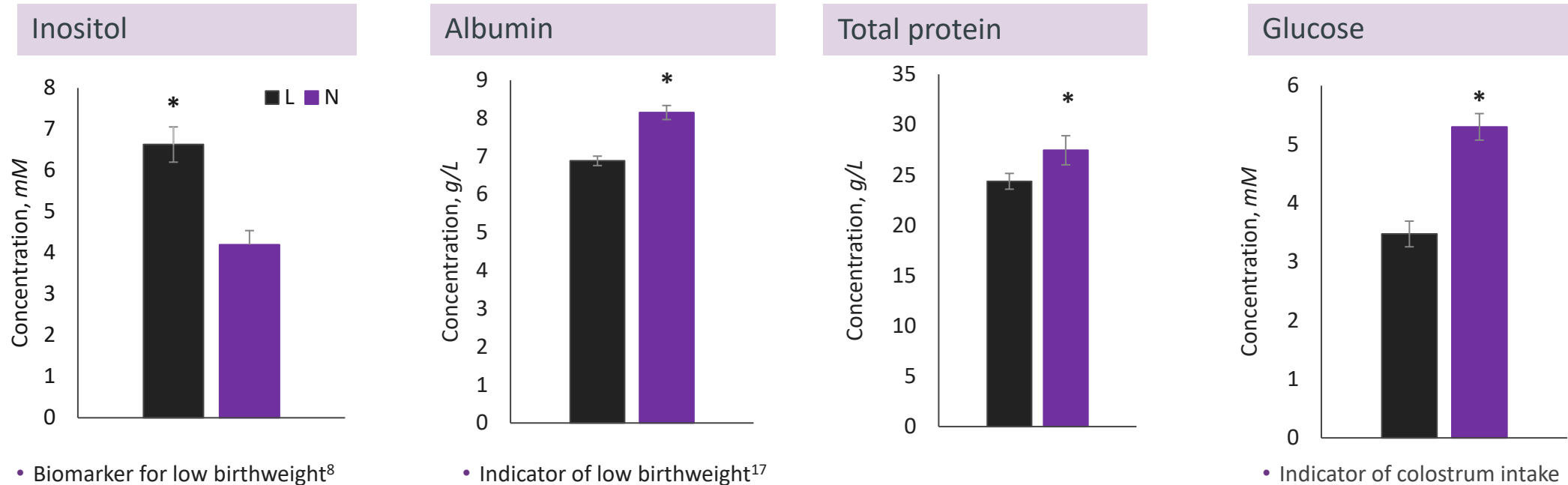
BW recorded daily



*(**L**), below the lowest birthweight quartile, (**N**); represents the middle 50th percentile, of the experimental pig facility

Tissue study: Plasma metabolites

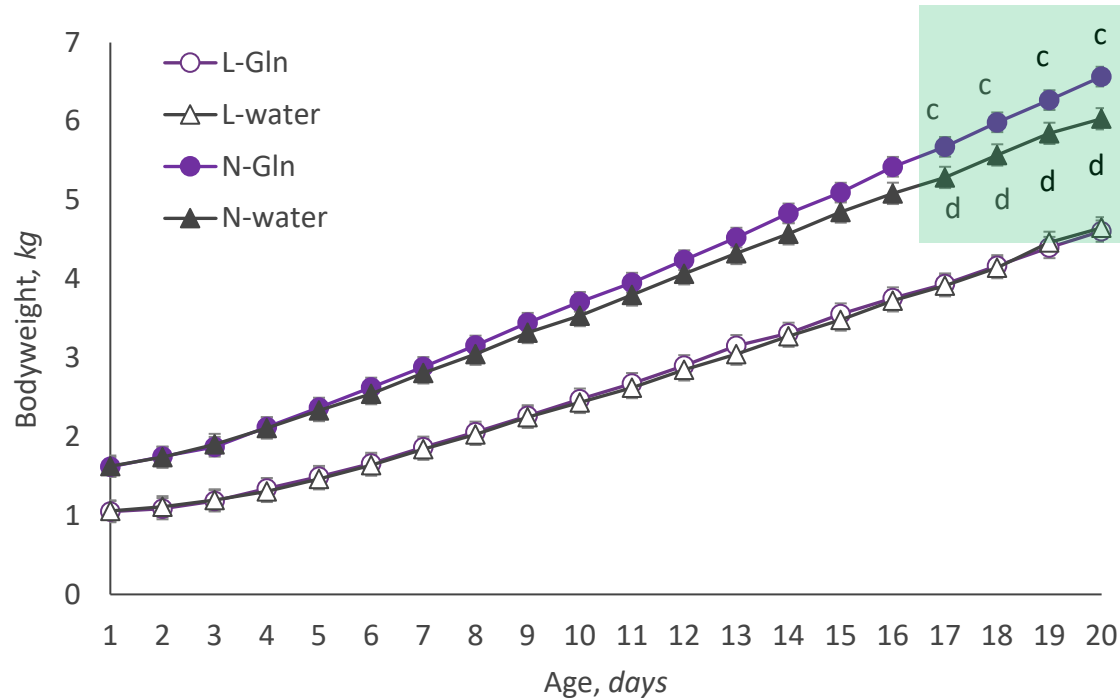
Plasma metabolites at 4 hours after birth



Taken together, these results provide strong evidence that the low birthweight piglets selected for this study are low birthweight

Tissue study: Piglet growth

Bodyweight



- From 17 – 20 d, N-Gln piglets were heavier than N-water

^{a,b}Different from N piglets within Supplementation group ($P < 0.05$).

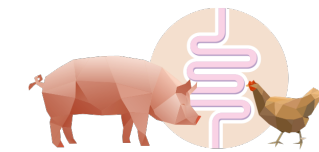
^{c,d}Different from Water piglets within Birthweight group ($P < 0.05$).

Average Daily Gain

Period (g/d)	L-Gln	L-water	N-Gln	N-water	SEM
1 - 20 d	190 ^b	200 ^b	264 ^{a,c}	236 ^{a,d}	13

- Bodyweight and Average Daily Gain were always higher in N then L piglets

- Average daily gain was higher in N-Gln than N-Water

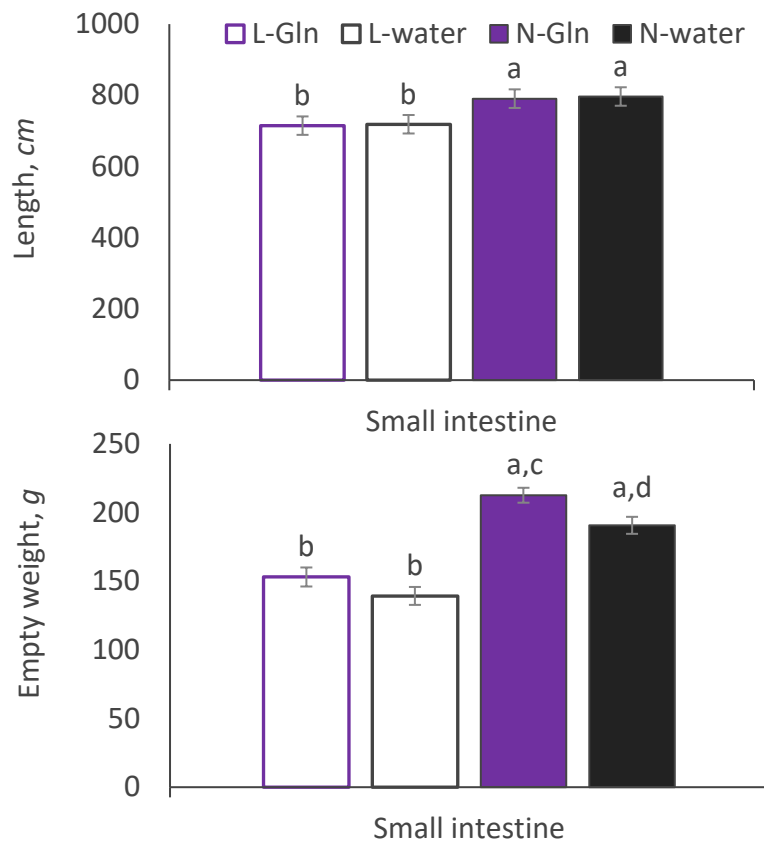


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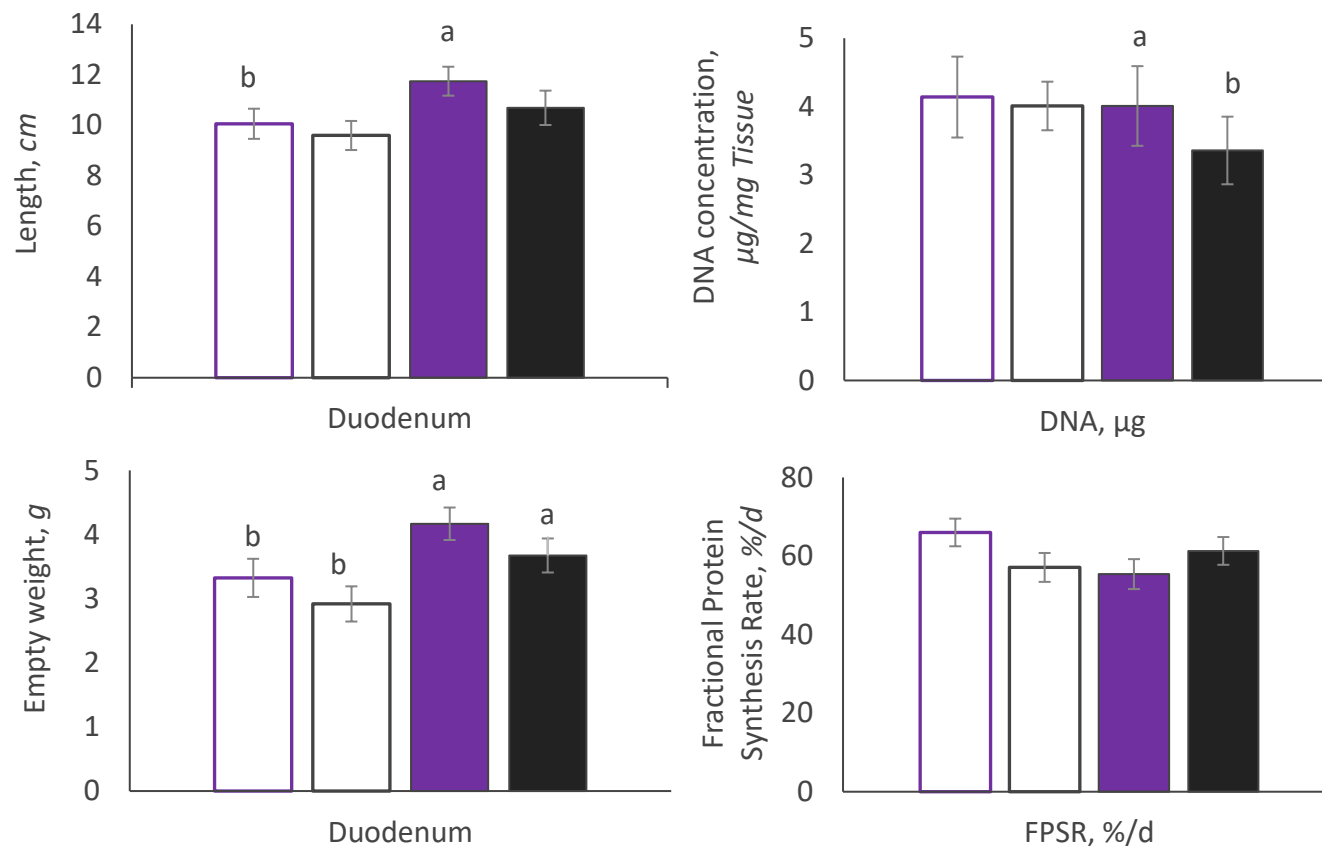
Tissue study: Intestinal parameters

Red blood Cell Glutathione concentrations were measured.
No differences were observed.

Small intestine (duodenum + jejunum + ileum)



Duodenum



^{a,b}Different from N piglets within Supplementation group ($P < 0.05$).

^{c,d}Different from Water piglets within Birthweight group ($P < 0.05$).



Overall Conclusions and Questions:

From the studies we have conducted:

1. Glutamine cannot be considered as a reliable supplement for improving low birthweight piglet growth

- **Q-PIG1**: L birthweight piglets supplemented with Gln were heavier than Ala supplemented

- **MGH Catheter**: L birthweight piglets supplemented with water were heavier than Gln supplemented

- **MGH Tissue**: N birthweight piglets supplemented with Gln were heavier than Water supplemented

2. **MGH Catheter**: Observed changes in glutamine and glucose metabolism were not associated with birthweight. Slower enrichment in N-Gln than L-Gln..... *Why?*

3. **MGH Tissue**: Increased bodyweight and average daily gain is associated with a heavier small intestine and higher duodenal DNA concentration in N-Gln than N-water..... *It worked?*

Acknowledgements

PhD Students

Q-PIG1



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Marianne Zenk



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15 – 18 September 2025
Rostock-Warnemünde, Germany



<https://isep2025-fbn.de/>



Research Institute for
Farm Animal Biology

Freie Universität



Berlin

Universität
Rostock



Traditio et Innovatio



Christian-Albrechts-Universität zu Kiel

THANK YOU

Do you have any questions?

Location: Workgroup „Nutritional Physiology“
Research Institute for Farm Animal Biology
Dummerstorf, Germany

Presenter: Quentin Sciascia, Ph.D.

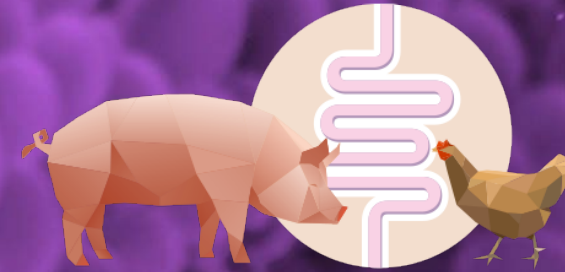
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Optimal gut function in monogastric livestock

References

1. Augere-Granier, Marie-Laure. "The EU pig meat sector." (2020).
2. Knap PW, Knol EF, Sørensen AC, Huisman AE, van der Spek D, Zak LJ, Granados Chapatte A and Lewis CRG (2023) Genetic and phenotypic time trends of litter size, piglet mortality, and birth weight in pigs. *Front. Anim. Sci.* 4:1218175. doi: 10.3389/fanim.2023.1218175.
3. Quiniou, Nathalie, J. Dagorn, and D. Gaudré. "Variation of piglets' birth weight and consequences on subsequent performance." *Livestock production science* 78.1 (2002): 63-70.
4. Declerck, Ilse, et al. "Long-term effects of colostrum intake in piglet mortality and performance." *Journal of Animal Science* 94.4 (2016): 1633-1643.
5. Stange, K., et al. "Low birth weight influences the postnatal abundance and characteristics of satellite cell subpopulations in pigs." *Scientific reports* 10.1 (2020): 6149.
6. Fix, J. S., et al. "Effect of piglet birth weight on survival and quality of commercial market swine." *Livestock Science* 132.1-3 (2010): 98-106.
7. Wellington, Michael O., et al. "Serum metabolomic characterization in pigs in relation to birth weight category and neonatal nutrition." *Journal of Animal Science* 101 (2023): skac386.
8. Nissen, Pia Marlene, et al. "Metabolomics reveals relationship between plasma inositols and birth weight: possible markers for fetal programming of type 2 diabetes." *BioMed research international* 2011.1 (2011): 378268.
9. Santos, Thaís Garcia, et al. "Intrauterine growth restriction and its impact on intestinal morphophysiology throughout postnatal development in pigs." *Scientific Reports* 12.1 (2022): 11810.
10. Wu, Guoyao, et al. "Triennial Growth Symposium: important roles for L-glutamine in swine nutrition and production." *Journal of animal science* 89.7 (2011): 2017-2030.
11. Li, Zeyang, et al. "Glutamine supplementation moderately affects growth, plasma metabolite and free amino acid patterns in neonatal low birth weight piglets." *British Journal of Nutrition* 128.12 (2022): 2330-2340.
12. Schregel, Johannes, et al. "Effects of oral glutamine supplementation on jejunal morphology, development, and amino acid profiles in male low birth weight suckling piglets." *Plos one* 17.4 (2022): e0267357.
13. Yang, X. F., et al. "Improved milk glutamine level and growth performance of suckling piglets by glutamine supplementation in maternal diet." *Annals of Animal Science* 18.2 (2018): 441-452..
14. Wu, Guoyao, et al. "Glutamine and glucose metabolism in enterocytes of the neonatal pig." *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology* 268.2 (1995): R334-R342
15. de Aquino, R. Santos, et al. "Glutamine and glutamate (AminoGut) supplementation influences sow colostrum and mature milk composition." *Livestock Science* 169 (2014): 112-117.
16. Xiao, Ying-ping, et al. "Response to weaning and dietary L-glutamine supplementation: metabolomic analysis in piglets by gas chromatography/mass spectrometry." *Journal of Zhejiang University Science B* 13 (2012): 567-578.
17. Quesnel, H., et al. "Physiological traits of newborn piglets associated with colostrum intake, neonatal survival and preweaning growth." *animal* 17.6 (2023): 100843.
18. Haynes, Tony E., et al. "L-Glutamine or L-alanyl-L-glutamine prevents oxidant-or endotoxin-induced death of neonatal enterocytes." *Amino acids* 37 (2009): 131-142.

Image 1: Beaumont, M., Blachier, F. (2020). Amino Acids in Intestinal Physiology and Health. In: Wu, G. (eds) Amino Acids in Nutrition and Health. Advances in Experimental Medicine and Biology, vol 1265. Springer, Cham. https://doi.org/10.1007/978-3-030-45328-2_1

Image 2: Nguyen, Trang TT, et al. "Methodological Approaches for Assessing Metabolomic Changes in Glioblastomas." *Autophagy and Cancer: Methods and Protocols* (2022): 305-328.