

Preservation of milk in liquid nitrogen during sample collection does not affect RNA quality for RNA-seq analysis

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INTRODUCTION

1.2. Why studying the mammary gland transcriptome?

1. Introduction

2. Objectives

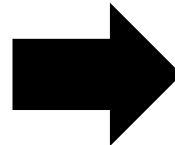
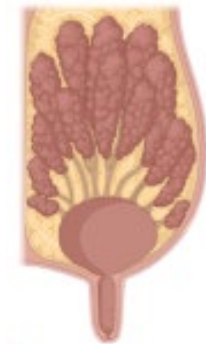
3. Materials and Methods

4. Results and Discussion

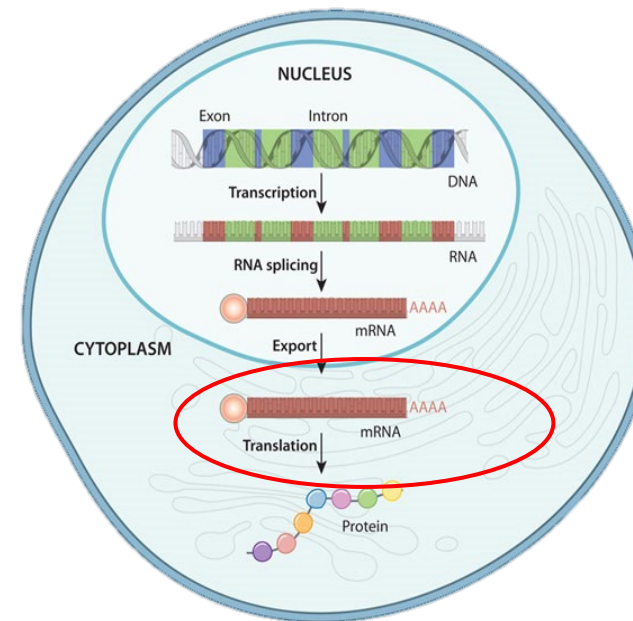
5. Conclusions

MAMMARY GLAND

In charge of **synthesis** and **secretion** of **MILK**



TRANSCRIPTOMIC



Will increase the BIOLOGICAL and PHYSIOLOGICAL knowledge of **LACTATION**

1.3. RNA sources for studying the mammary gland

1. Introduction

2. Objectives

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From TISSUE samples

- Mammary biopsies
- Laser microdissected mammary epithelial cells (LCMEC)

- ✗ Damage
- ✗ No opportunities for multiple sampling
- ✗ Disrupts normal lactation process

From MILK samples

- Milk Somatic cells (MSC)
- Antibody-captured milk mammary epithelial cells (mMEC)
- Milk Fat Globules (MFG)

- ✗ MSC: only 2% of the cells are mammary epithelial cells (MEC)
- ✗ mMEC: difficult to obtain

✓ **MFG** → A good representation of the mammary gland transcriptome (Yang et al., 2015)



1. Introduction

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MFG formation

The diagram illustrates the formation of a Milk Fat Globule (MFG) within a Mammary Epithelial Cell (MEC). On the left, a cow icon points to the MEC. Inside the cell, the nucleus and endoplasmic reticulum are shown. Triglyceride (TAG) cores are synthesized in the ER and move toward the apical plasma membrane. Lipid droplets, consisting of a TAG core and a phospholipid monolayer, are released into the cytoplasm. These droplets then acquire fragments of the apical plasma membrane bilayer to form the MFG envelope. A detailed view of an MFG shows it as a lipid droplet with an apical plasma membrane bilayer envelope. The envelope contains fragments of MEC cytoplasmic crescents, which may contain RNA. A final note states: 'In which different types of RNA from MEC could be found'.

MFG formation

Mammary Epithelial Cell (MEC)

Apical Plasma Membrane Bilayer

Lipid droplets
TAG core + Phospholipid Monolayer

Triglyceride (TAG) cores

Endoplasmic Reticulum

Nucleus

Milk Fat Globules (MFG)
Lipid Droplet + Apical Plasma Membrane Bilayer

The MFG envelop fragments of MEC (cytoplasmic crescents)

In which different types of **RNA** from MEC could be found

1.4. Milk sample collection for RNA-seq analysis

1. Introduction

2. Objectives

3. Materials and Methods

4. Results and Discussion

5. Conclusions

Standard procedures for **milk sample collection** in **transcriptome analysis**

ICE

- Require immediate milk sample processing
- **MILK**: challenging matrix
 - High abundance of ribonucleases
 - Low quantity of RNA



ALTERNATIVE MILK SAMPLE COLLECTION

- No immediate milk sample processing
- Inactivation of ribonucleases

Liquid Nitrogen

OBJECTIVES

The applicability of a method for milk preservation with **liquid nitrogen** during sample collection, subsequent **extraction of total RNA from milk fat globules** and its sequencing by **RNA-seq**

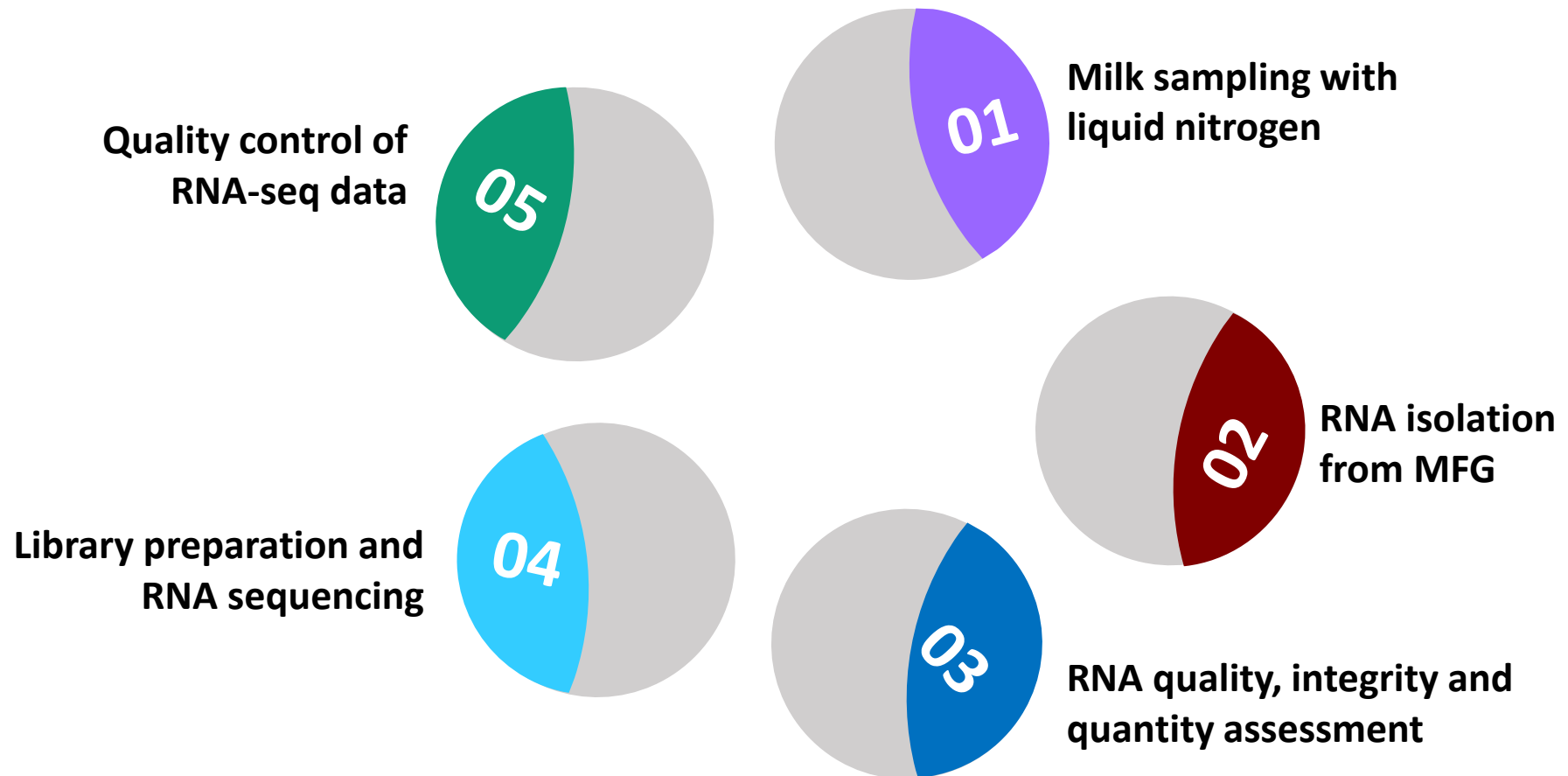
1. Introduction

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MATERIAL AND METHODS

3.1. Milk sampling with liquid nitrogen

1. Introduction

2. Objectives

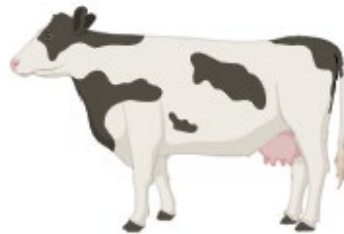
3. Materials
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❖ **Fifteen Holstein cows** from a commercial farm (Navarre, Spain) were selected for the study.

Milk was directly
collected **from the udder**
of the cow



...into **50 mL**
RNase-free
tubes...



...which were immediately snap-
frozen in **LIQUID NITROGEN** and then
stored at -80°C until further analysis.



3.2. RNA isolation from milk fat globules (MFG)

1. Introduction

2. Objectives

3. Materials and Methods

4. Results and Discussion

5. Conclusions

Milk sample pretreatment



To enable **MFG collection**



✓ MFG fraction

Skim milk

Somatic cell pellet

RNA isolation



Using **Trizol LS Reagent**

Sample lysis



Fat layer removal



Phase separation



RNA precipitation



Column-based method



RNA elution



3.3. RNA integrity, quality and quantity assesment

1. Introduction

2. Objectives

3. Materials
and Methods

4. Results and
Discussion

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Nanodrop 2000

- Concentration: A_{260}
- Quality: $A_{260/280}$
✓ RNA: 1.9-2.1
- Quality: $A_{260/230}$
✓ RNA: 2.0-2.2

*A= Absorbances

TapeStation 4200 RNA Screentape

- Integrity: RIN
✓ RNA ≥ 7

*RIN= RNA Integrity Number

Qubit 4 fluorometer

- Concentration: RNA HS
- Integrity: RNA IQ
✓ RNA ≥ 9

*HS=High Sensitivity; IQ=
Integrity and Quality

Statystical analysis:

- $A_{260/280}$: one sample equivalence test
- RIN and RNA IQ : one-sample non-inferiority test

3.4. Library preparation, mRNA sequencing and quality control

1. Introduction

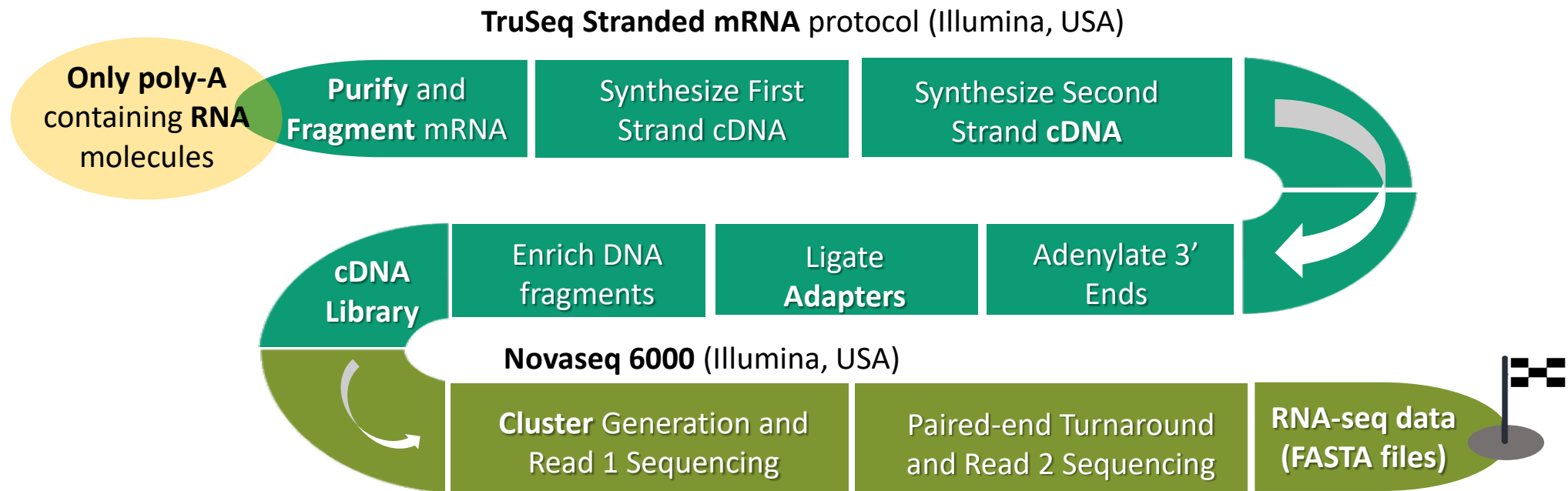
2. Objectives

3. Materials and Methods

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- Library preparation and mRNA sequencing



- Quality control of RNA-seq data

– Base quality scores of RNA-seq reads: **Phred scale (Q)**

✓ Q = 30 – 40
✗ Q < 20: low quality

+ FastQC Report

RESULTS AND DISCUSSION

4.1. RNA integrity, quality and quantity assesment

Quality and Integrity

1. Introduction

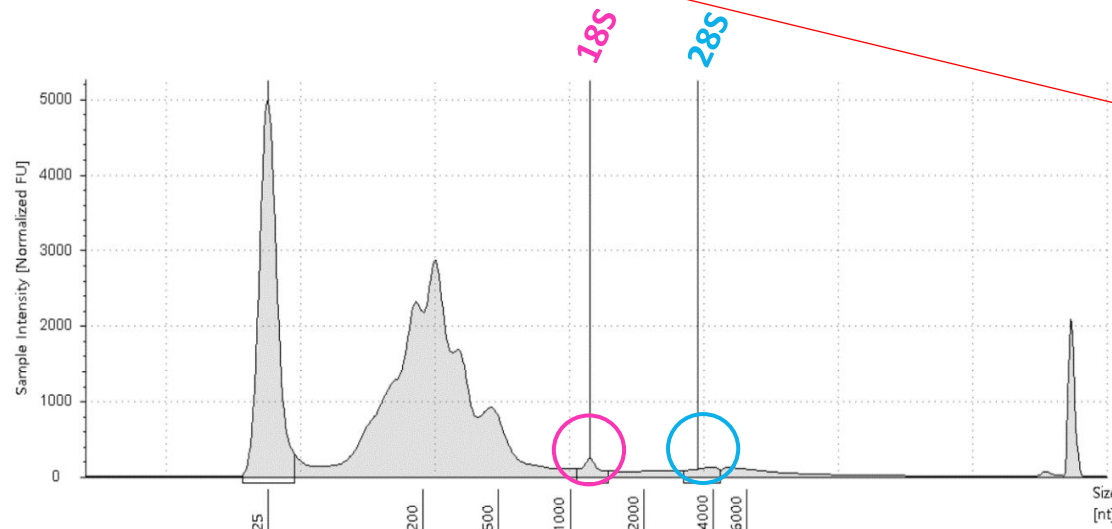
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Variable	Mean	SE	Reference value	Tested H_0	P-value
$A_{260/280}$	2.03	0.01	1.9-2.1	Mean-Reference ≤ -0.1 or Mean-Reference ≥ 0.1	$P_1 = 0.000 / P_2 = 0.000$
IQ	9.51	0.15	> 9	Mean-Reference ≤ 0	$P = 0.010$
RIN	3.59	0.27	> 7	Mean-Reference ≤ 0	$P = 1.000$



The **RNA from the MFG** contains a large amount of **low molecular weight RNA fragments** and a small amount of 28S and 18S rRNA (RIN).

4.1. RNA integrity, quality and quantity assesment

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Variable	Mean	SE	Minimum	Maximum	<i>n</i>
Nanodrop (ng/μL)	120.43	22.3	31.09	366.91	15
Qubit HS (ng/μL)	102.87	15.6	22.00	198.00	14

Quantity

Same initial MFG quantity per animal (20-25 milk fat), but...

Low RNA concentration & High variability between animals

4.2. cDNA library quantification

1. Introduction

2. Objectives

3. Materials and Methods

4. Results and Discussion

5. Conclusions

Variable	Mean	Standard error	Minimum	Maximum	<i>n</i>
Library concentration (nM)	8.64	5.0	1.60	39.70	15

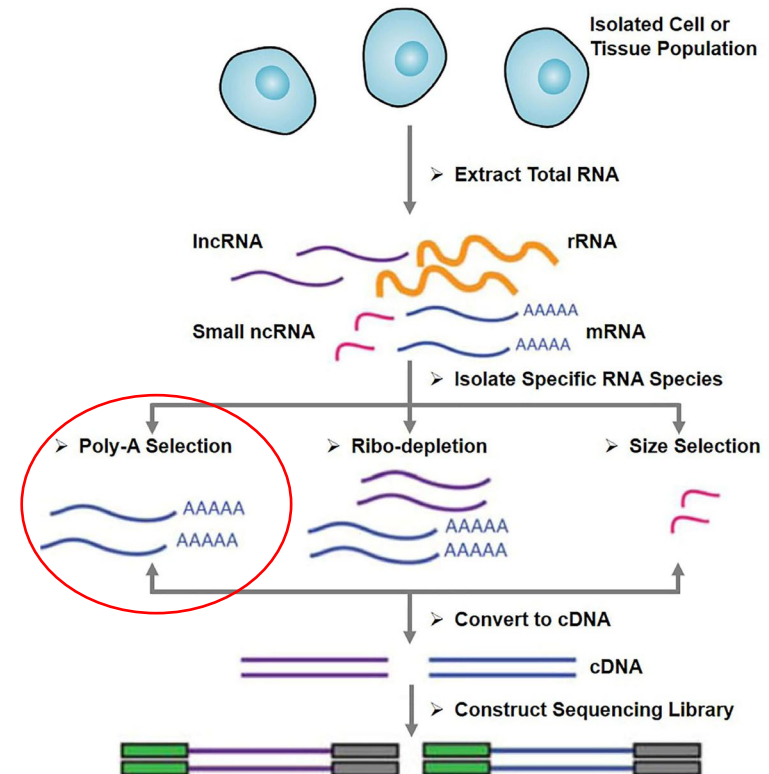
High variability among cDNA libraries

Same initial input

(1µg of total RNA/animal)

Why?

Different quantities of **mRNA molecules** within **total RNA**



4.3. Quality control of RNA-seq data

1. Introduction

2. Objectives

3. Materials
and Methods

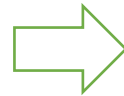
4. Results and
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A) Sequencing results

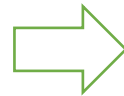
- A total of **791 million reads** were generated.
- An average read number of **52.7 million reads per sample**.

Q20



The **98%** of the nucleotides were identified with an error LESS than 1/100

Q30



The **94-95%** of the nucleotides were identified with an error LESS than 1/1000

Good quality of sequencing results

4.3. Quality control of RNA-seq data

1. Introduction

2. Objectives

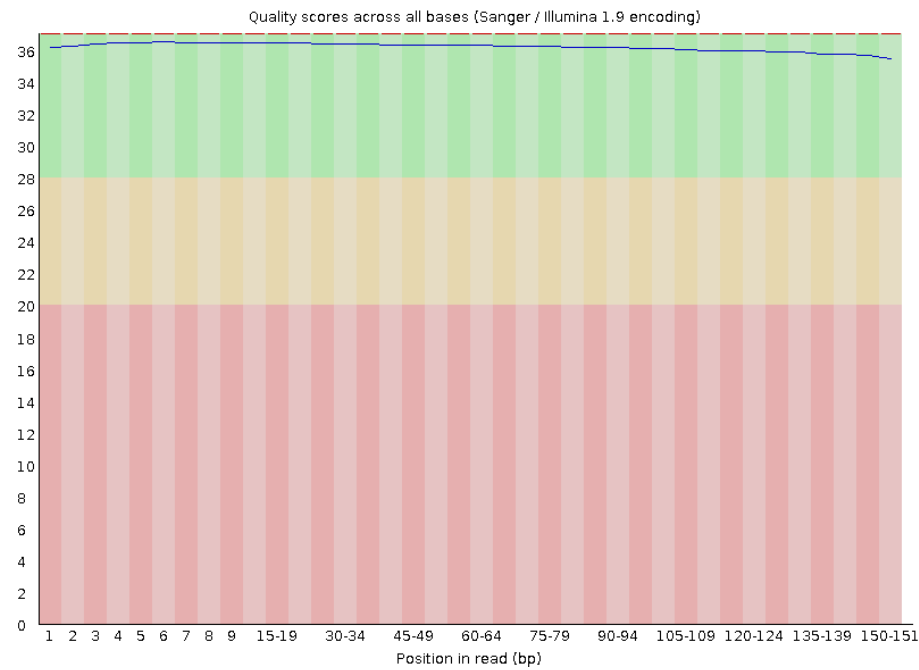
3. Materials
and Methods

4. Results and
Discussion

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B) FastQC results

- Quality score per base ✓ Q= 36



4.3. Quality control of RNA-seq data

1. Introduction

2. Objectives

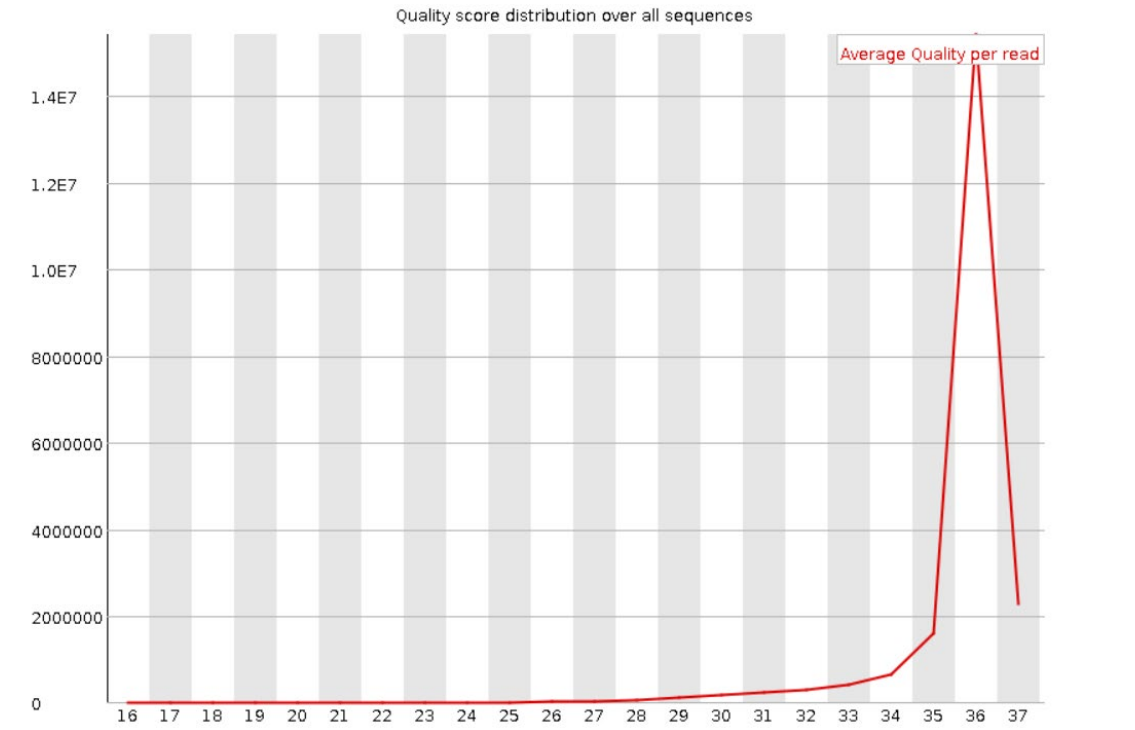
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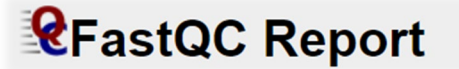
5. Conclusions

B) FastQC results

- Quality score per sequence ✓ Q= 36

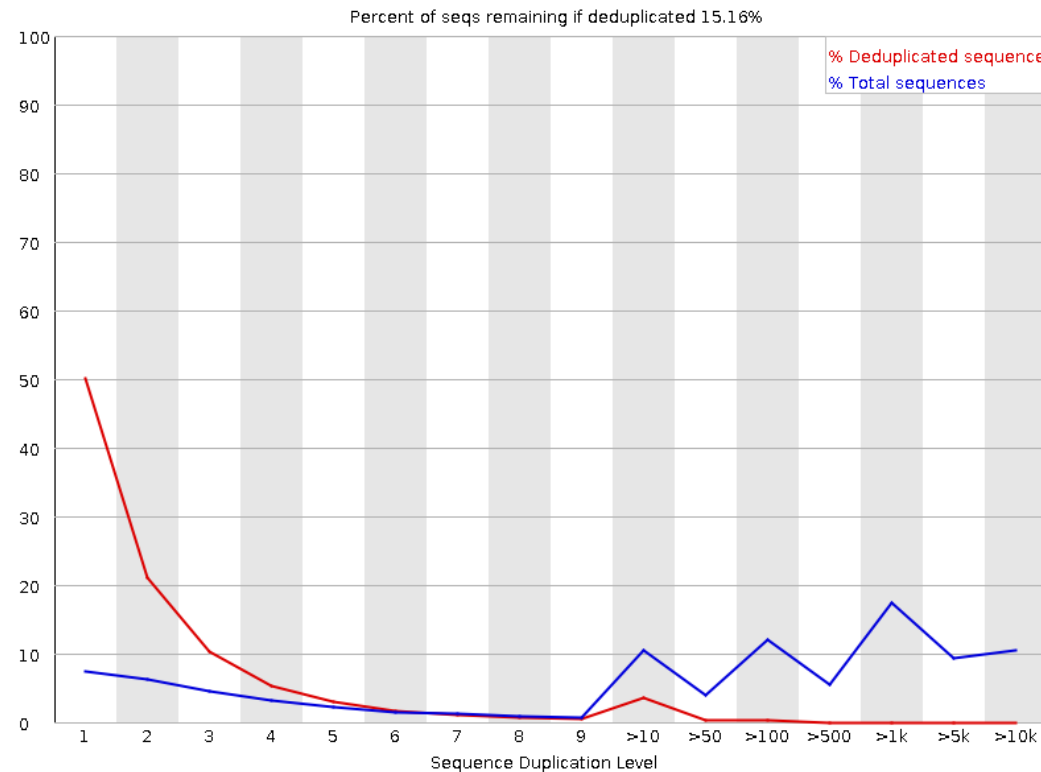


4.3. Quality control of RNA-seq data



B) FastQC results

- Sequence Duplication levels $\longleftrightarrow$ Overrepresented sequences



In a RNA-seq experiment is normal the presence of...

- Biologically relevant transcripts** for this RNA matrix.

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CONCLUSION

1. Introduction

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GENERAL CONCLUSIONS

- Results suggest that **milk preservation** using **liquid nitrogen** is a **suitable sample collection method** that prevents RNA degradation and overcomes the limitations of immediate sample processing required if using ice.
- The **quality, integrity and quantity of the RNA extracts** isolated from **MFG** were adequate allowing **successful downstream RNA-seq analysis**.

TAKE HOME MESSAGES

- This procedure could be considered a more **practical** and **non-invasive** means of **measuring the mammary epithelial cell transcriptome**.
- The **RNA** isolated from **MFG** contained **low molecular** RNA fragments and a very **low amount of 18S and 28S rRNA** due to the presence of small amounts of cytoplasmic material. Because of that **RIN values** are generally **below the benchmark of 7**.