

# Hepatic mitochondrial function in dairy cows with different levels of pasture inclusion during lactation

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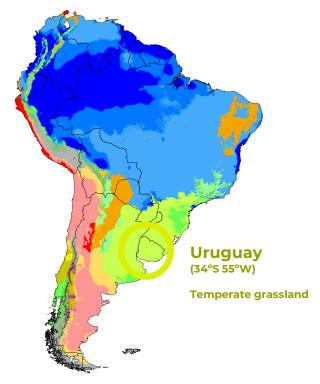








# **INTRODUCTION**

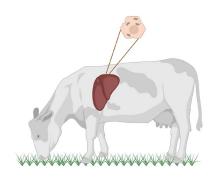


- Exports >70% of milk production (INALE, 2024)
- 9<sup>th</sup> exporter (INALE, 2024)
- Maximizing harvested pasture and minimizing reliance on supplementary feed are key to profitable pasture-based systems (Neal and Roche, 2019)
- Dependence on environmental conditions and management (Wales et al., 2013; Chilibroste et al., 2015; Chilibroste et al., 2005)
- Insufficient DMI and unstable nutrient availability (Chilibroste et al., 2012, Kolver and Muller, 1998)



Cows in grazing systems show a poorer metabolic status (†BHB, †NEFA \*Ins., \*IGF-I) and lower perfomance when compared to cows in confined systems (Meikle et al., 2013; Astessiano et al., 2015)

Cows in a TMR+Grazing systems *vs.* cows in a Grazing+Concentrate system have ↑DMI, ↑milk yield, ↑%fat, ↑%protein, ↓BCS loss (Wales et al., 2013).

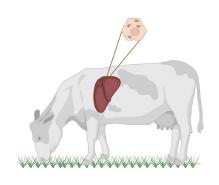


Liver plays a key role coordinating nutrient fluxes to support lactation and is central to the adaptation of metabolic pathways (Drackley et al., 2001).



The mitochondrion is the organelle responsible for most of the ATP produced in the cell and although it is dynamic, it is highly prone to damage (Nicholls and Ferguson, 2011).

Liver mitochondria are related to metabolic adaptations of lactation as higher mitochondrial function is associated with increased milk yield (Favorit et al., 2021).



Impaired hepatic mitochondrial function has been observed in dairy cows in pasture-based systems during early lactation (García-Roche et al., 2019, 2023).

Nutrient utilization for metabolic processes in the hepatocyte differs among feeding strategies and Holstein strains (García-Roche et al., 2021, 2022).



# **HYPOTHESIS**

Hepatic ATP-synthesis will be decreased in feeding systems with higher levels of pasture inclusion during early lactation

# **OBJECTIVE**

Study the association between stage of lactation and proportion of pasture on hepatic mitochondrial function.



# **MATERIALS AND METHODS: Animal trials**

2015-2016

Estación experimental

"Dr. Mario A. Cassinoni".

Facultad de Agronomía

Universidad de la República

- N = 24
- Multiparous NA Holstein
- Spring calving
- 664 ± 65 kgBW
- 3.0 ± 0.4 BCS
- Blocked according to BW, BCS, calving date, parity and previous annualmilk yield



TMR (0) 100% TMR (60:40, DM ratio)



Grazing (100) 85% Pasture<sup>1, 2</sup> 15% Commercial concentrate (offered basis)

2017-2018-2019



- N = 48
- Multiparous NA and NZ Holstein
- Fall calving





538 ± 63 kgBW 582 ± 59 kgBW 3.23 ± 0.19 BCS 3.03 ± 0.28 BCS



PMR (30) 31-33% Pasture<sup>1</sup> 67-69% TMR (55:45, DM ratio)



Grazing (100) 75-80% Pasture<sup>1</sup> + Forrage reserves<sup>2</sup> (40-50% grazed) 20-25% Commercial concentrate



DIM = -21, 35, 60, 110, 180, 250
Daily milk yield
Milk composition, BW, BCS - fortnightly

<sup>1</sup>Festuca arundinacea+ Medicago sativa. <sup>2</sup>Diet change due to <<ITH (113-180 DPP) cows grazed Festuca arundinacea (20 kgDM/cow/day) + TMR (15 kgDM/cow/day)



DIM = -45, 21, 100, 180
Daily milk yield
Milk composition, BW, BCS - fortnightly

<sup>1</sup>Festuca arundinacea, Dactylis glomerata + Medicago sativa.

<sup>2..</sup> Corn silage+ pasture haylage 75:25 (DM)



**MATERIALS AND METHODS:** 

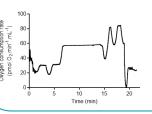


**DPAP** 

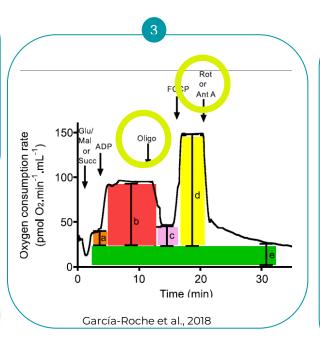
Levels of plasma and liver metabolites were determined with commercial kits



O2k-High Resolution Respirometry using mitochondrial substrates and inhibitors







Variables were grouped:

- Mitochondrial parameters related to ATP synthesis (n = 10)
- Mitochondrial parameters not related to ATP synthesis (n = 8)
- Energy balance markers (n = 5)

**PROC FACTOR** – Principal component analysis Previously confirmed with Bartlett's Sphericity Tests on the three groups (P < 0.001)

PROC CORR - Spearman's correlations, heatmap

PROC MIXED - Mixed models with repeated measures

Fixed effects: STAGE of lactation, PROP proportion of pasture, STAGE x PROP Random effect: Holstein strain

Covariate: Trial

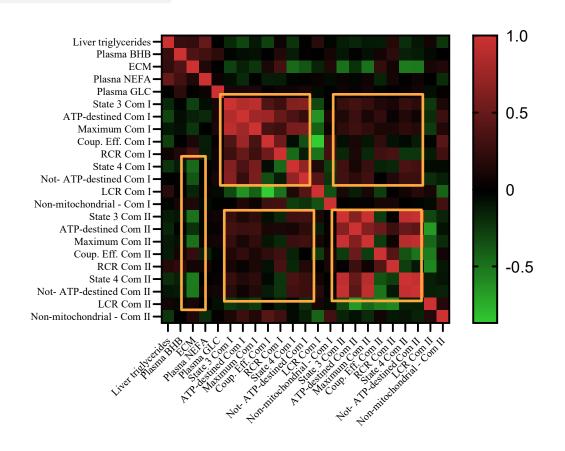
Least square means for mean separation Means were considered to differ when  $P \le 0.05$ and tendencies when  $0.05 < P \le 0.10$ 

PROC UNIVARIATE, REG - check normality and t student residuals

- Respiration destined to ATP-synthesis
- Proton leak
- Non-mitochondrial respiration



# **RESULTS**



Mitochondrial parameters when the same mitochondrial substrate was used correlated positively ( $\rho = 0.20 - 0.90$ , P < 0.01)

Did not correlate between complexes

Differences among mitochondrial substrates -different pathways that direct their reducing equivalents to either Complex I (TCA) or II (AA and fatty acids).

Weak to moderate negative correlations were observed between ECM and mitochondrial parameters ( $\rho$  = -0.20 to -0.52, P < 0.0001).







PC1 and 2 explain 58% of the variance

Factor Pattern		
	Factor1	
Plasma BHB	0.81231	
ECM	0.63578	
Liver TAG	0.62209	
Plasma NEFA	0.59463	
Plasma GLC	0.20657	



PC1 and 2 explain 60% of the variance

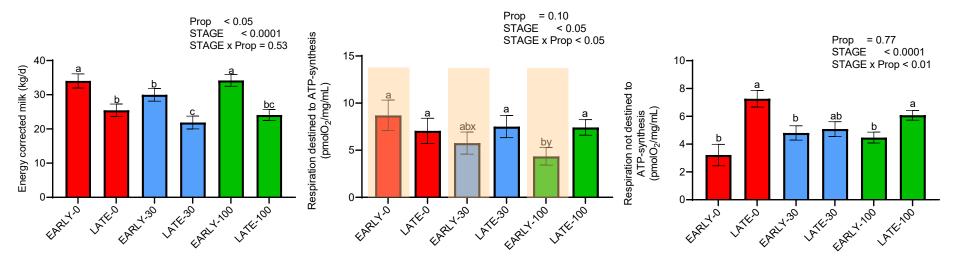
	Factor Pattern		
	Factor1	Factor2	Factor3
		1	
ATP-destined Com I	0.8955	-0.29247	-0.26165
State 3 Com I	0.88441	-0.12381	-0.27316
Maximum Com I	0.84251	-0.10784	-0.24736
Maxilliulli Colli I	0.84251	-0.10/64	-0.24/30
ATP-destined Com II	0.62715	0.45808	0.52388
	****		******
Coup. Eff. Com I	0.56942	-0.4043	-0.1677
State 3 Com II	0.38995	0.89202	0.0436
Maximum ComII	0.3873	0.88952	0.09494
Coup Eff Com!	0.22102	0.44107	0.60422
Coup. Eff. Com II	0.33183	-0.44197	0.68432
RCR Com I	0.26643	-0.379	0.22889
RCR Com II	0.15545	-0.25426	0.70378



PC1 and 2 explain 54% of the variance

	Factor Pattern	
	Factor1	Factor2
Not - destined ATP Com I	0.75877	-0.25411
Not - destined ATP Com II	0.74006	0.55841
State 4 Com I	0.72403	-0.14845
State 4 Com II	0.71724	0.58275
LCR Com I	0.48163	-0.50561
LCR Com II	-0.13119	0.07384
Non-mitochondrial Com I	-0.19076	0.6317
Non-mitochondrial Com II	-0.24114	0.65202

## **RESULTS**



Liver mitochondria are related to metabolic adaptations of lactation as higher mitochondrial function is associated with increased milk yield (Favorit et al., 2021).

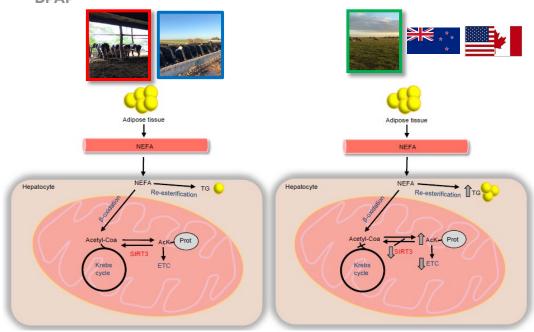
Uncoupling proteins (UCPs) 1, 2 and 3 and adenine nucleotide translocase induce proton leak in response to high fatty acid levels, superoxide or lipid peroxidation products (Toime and Brand, 2010).

UCPs may play a central rol in the adaptation to increased fatty acid supply by enhanced fatty acid oxidation (Cortez-Pinto and Machado, 2009)

UCPs regulate oxidative phosphorylation efficiency protecting against oxidative damage (Brand, 2005).



# **CONCLUSION**



García-Roche et al., 2019

 Greater mobilization leads to greater esterification of liver TAGs during early lactation

 Hepatocytes of dairy cows with different levels of pasture inclusion show different adaptive strategies during lactation

 Highlight the plasticity of hepatic mitochondria to adapt to different energy demands and nutrients

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Thank you